Immunohistochemical study of neuroendocrine cells at the gastric cardia mucosa

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Background/Aims: The gastric cardia mucosa is a narrow band of tissue between the oesophagus and the stomach. The physiological role of this tissue is unknown. This study examined the presence and characteristics of neuroendocrine cells at this site.

Methods: Biopsy samples were obtained from across normal appearing squamocolumnar junctions. The cardiac mucosa was defined as the presence of special type mucosa composed of mucous secreting glands in the immediate vicinity of oesophageal squamous epithelium. Biopsy specimens were stained with haematoxylin and eosin, alcian blue (pH 2.5) periodic acid Schiff, and modified Giemsa. The chromogranin A and Fontana-Masson stains were used to identify neuroendocrine cells, which were also stained immunohistochemically for gastrin, serotonin, glucagon, pancreatic polypeptide, somatostatin, and vasoactive intestinal peptide.

Results: Chromogranin positive cells were seen in 18 cases with adequate biopsy specimens from the gastric cardia mucosa. These cells were all serotonin positive, but stains for other peptide hormones remained negative. Serotonin positive cells were detected only at the base of foveolae at the periphery of mucous secreting cardiac glands, giving a microscopic appearance resembling that of endocrine cells at the gastric antrum. The presence and numbers of serotonin positive cells did not correlate with chronic inflammation or intestinal metaplasia of the cardiac mucosa. These cells were seen both in Helicobacter pylori positive and negative patients.

Conclusions: Serotonin positive cells appear to be the sole neuroendocrine cell type at the gastric cardia mucosa. These cells may have a role in regulating the physiology of the gastric cardia mucosa and the lower oesophageal sphincter.

Neuroendocrine cells possess molecular machinery for the uptake and release of neurotransmitters and neuropeptide hormones. These cells enable autocrine communication both with adjacent (paracrine) and more distant (endocrine) cells. Gastric neuroendocrine cells comprise about 1% of the volume of the oxyntic mucosa, and at least six distinct cell types have been identified. The gastric mucosa contains enterochromaffin-like cells, G cells, and D cells, the respective secretory products of which include chromogranins and histamine, gastrin, and somatostatin.

The gastric cardia mucosa is a specialised type of mucosa on the gastric side of the squamous and columnar junction, which is composed of pits that occupy one half of the thickness of the mucosa; the glands are coiled and occasionally branched and secrete only mucous. Recent data indicate that the cardiac mucosa is congenital and gastric in origin. Electron microscopy has shown that the gastric cardiac contains numerous endocrine cells, which mainly occupy recesses within lateral surfaces of adjacent mucous or parietal cells. This study was published in the 1970s and endocrine cells were not identified by immunohistochemical methods.

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Neuroendocrine cells are involved in normal physiological processes at the gastric mucosa in addition to disease states. The role of gastrin secreted from antral G cells in the production of gastric acid is established. Hypochlorhydria may result in hyperplasia of gastric enterochromaffin cells or even in the development of carcinoid tumours of the gastric mucosa. Because only sparse data are available on gastric cardia neuroendocrine cells, we wanted to investigate the presence and characteristics of neuroendocrine cells at the gastric cardia mucosa.

MATERIALS AND METHODS
Biopsy specimens were obtained from gastric antrum and body, from the normal appearing squamocolumnar junction and from the columnar lined distal oesophagus (two specimens from each site). During endoscopy, the normal squamocolumnar junction or Z line was considered to be at the level of the proximal end of the gastric folds. Biopsy forceps were targeted across the squamocolumnar junction to obtain both columnar and squamous mucosa. If columnar appearing mucosa extended orally from the gastro-oesophageal junction, biopsy specimens were obtained separately from this mucosa to detect Barrett’s oesophagus. The presence of hiatal hernia was defined as the entire gastro-oesophageal vestibule remaining intrathoracic during both inspiration and expiration. Endoscopic examinations were performed by a single investigator (MV).

Histology
Formalin fixed biopsy specimens were embedded in paraffin wax and tissue sections were stained with haematoxylin and eosin, alcian blue (pH 2.5), periodic acid Schiff, and modified Giemsa. Only cases with typical gastric cardia mucosa in the

Abbreviations: PP, pancreatic polypeptide; VIP, vasoactive intestinal peptide
junctional biopsy specimens were included in our analysis. The presence of incomplete-type intestinal metaplasia in the biopsy specimens from columnar lined distal oesophagus was considered the diagnostic criterion for Barrett’s oesophagus.10 Chronic cardia inflammation was defined as infiltration of cardia mucosa by mononuclear cells.14 Gastric cardiac intestinal metaplasia was defined as the presence of goblet cells with (incomplete-type intestinal metaplasia) or without (complete-type intestinal metaplasia) acidic mucin in goblet and columnar-type cells in biopsy specimens obtained from normal appearing squamocolumnar junctions.12 Two pathologists examined the histological slides independently (MJ and PS).

Antibodies for immunohistochemical stains were supplied by Dako, Glostrup, Denmark (polyclonal anti gastrin, polyclonal anti glucagon, polyclonal antipancreatic polypeptide (anti-PP), polyclonal antisomatostatin, and monoclonal antiserotonin) and by Novocastra Laboratories, Newcastle upon Tyne, UK (monoclonal antivasoactive intestinal peptide; anti-VIP). The following dilutions were used: anti gastrin, 1/300; anti glucagon, 1/500; anti-PP, 1/1000; antisomatostatin, 1/10; antiserotonin, 1/10; anti VIP, 1/750; and anti VIP, 1/50. Before staining for somatostatin, the dewaxed sliced biopsy samples were treated with trypsin. Before staining for VIP, the dewaxed sliced biopsy samples were boiled in buffered citric acid (pH 6.0) in a microwave oven for four times five minutes. Immunohistochemical stains were performed using a Dako Technate 500, with an incubation time of 35 minutes. A Chemnate horseradish peroxidase/diaminobenzidine kit was used to detect neuroendocrine cells. Nuclei were stained with Mayer’s haematoxylin. For each peptide hormone stain, a known positive control tissue sample was used as an external control. The numbers of each type of neuroendocrine cell were classified as follows: 0, none, +, low number, ++, moderate number, ++++, abundant.

Ethics

The ethics committee of the Central Hospital of Central Finland approved the study.

RESULTS

Eighteen patients with biopsy specimens from normal appearing squamocolumnar junction were included. Two of the three patients with Barrett’s oesophagus also had typical cardiac mucosa in the biopsy specimens from the gastrooesophageal junction, bringing the number of studied patients with cardiac-type mucosa to 20 cases. These patients had been referred for endoscopy because of reflux disease symptoms (seven), dyspeptic symptoms (three), anaemia (four), coeliac disease (two), non-cardiac chest pain (one), dysphagia (one), surveillance of Barrett’s oesophagus (two), and surveillance of atrophic corpus gastritis (one). At endoscopy, hiatal hernia was detected in seven patients and signs of gastropathy (erythema and erosions) in nine, but none had erosions in the distal oesophagus. Histological examination of gastric antral and corpus biopsy specimens revealed chronic gastritis in seven patients (six *Helicobacter pylori* positive), whereas 13 had normal gastric histology.

Of the 20 patients with cardiac mucosa detected in biopsy specimens, four had normal non-inflamed cardiac mucosa, whereas 16 had chronic gastric cardia inflammation. In addition, six patients with “carditis” also had intestinal metaplasia at the cardia mucosa. Chromogranin positive cells at the gastric cardia mucosa were seen in 18 of the 20 patients. In the other two patients biopsy specimens were tangentially cut and superficial; these cases were deleted from the final analysis. Of the 18 patients in the final study population, six were *H pylori* positive, 14 had chronic cardiac inflammation (five with intestinal metaplasia), and four had normal non-inflamed cardiac mucosa.

All the chromogranin positive cells at the gastric cardia were serotonin positive when stained immunohistochemically; the numbers of serotonin positive cells were classified as low (+) in 12 and moderate (++) in six cases (fig 1). The mean age of the patients with serotonin positive cells at the cardiac mucosa was 52 years (9 to 74 years) and the male to female ratio was 10:8.

In contrast, the cardiac mucosa was negative for gastrin, glucagon, PE somatostatin, and VIP. Of the three patients with Barrett’s oesophagus, only one sample with metaplastic oesophageal epithelium contained chromogranin positive cells that were serotonin positive. All other immunohistochemical stains for Barrett’s mucosa were negative. Instead, biopsy specimens of gastric antral mucosa contained numerous gastrin positive cells, whereas specimens of oesophageal squamous epithelium (four) contained neither chromogranin nor serotonin positive cells. Gastric antral and corpus mucosa also contained endocrine cells that were positive for VIP and somatostatin, but negative for glucagon and PP.

DISCUSSION

Our results show that serotonin is the sole secretory product of the neuroendocrine cells at the gastric cardia mucosa. In this respect cardiac mucosa differs sharply from gastric mucosa, which contains several types of neuroendocrine cells with a large variety of secretory products.2 There are conflicting opinions over whether cardiac mucosa is a normal or a metaplastic phenomenon resulting from—for example, pathological gastrooesophageal reflux.4,5,15 Our present study revealed that serotonin positive cells occur in normal and inflamed cardiac mucosa with or without intestinal metaplasia. Our results are concordant with those of Stachura et al, who showed that gastric cardia endocrine cells were negative for gastrin, glucagon, somatostatin, VIE secretin, motilin, PE insulin, and substance P.14 Although we attempted to quantify the number of neuroendocrine cells, the results must be interpreted with caution. In our study, neuroendocrine cells were present only in the base of foveolae and at the periphery of mucous secreting glands. Endoscopic biopsy specimens are often cut tangentially, and the number of neuroendocrine cells in such specimens may be low as a result of inadequate tissue sampling. The failure to detect serotonin positive cells in two cases probably resulted from problems in biopsy sample collecting and orientation.

The physiological role of gastric cardia mucosa is unknown. Based on our present study with its very limited number of cases only hypotheses can be presented. The lower oesophageal sphincter system—the main mechanism for regulating the passage of food—is controlled mainly by cholinergic innervation.15 Selective reduction of cholinergic muscle

![Figure 1 Moderate immunohistochemical staining for serotonin in neuroendocrine cells at the gastric cardia.](http://jcp.bmj.com/10.1136/jcp.55.10.767)
Serotonin may be involved in the production and perception of non-cardiac chest pain, for which gastro-oesophageal reflux disease is probably an aetiological factor. 

Serotonin may stimulate cholinergic neurones to release acetylcholine, which in turn results in smooth muscle contraction. 

Furthermore, gastric mucosal functions such as secretion, growth, and repair after mucosal damage require complex interactions between neuronal and paracrine messengers, growth factors, hormones, and cytokines. We speculate that serotonin produced by neuroendocrine cells in the gastric cardia mucosa may contribute to the regulation of functions in both the gastric cardia mucosa and the lower oesophageal sphincter.

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References