Primary follicular lymphoma of the testis in childhood: an entity with peculiar clinical and molecular characteristics

S A Pileri, E Sabattini, P Rosito, P L Zinzani, S Ascani, G Fraternali-Orchioni, B Gamberi, M Piccioli, D Vivenza, B Falini, G Gaidano

ORIGINAL ARTICLE

Background/Aims: Paediatric primary follicular lymphoma of the testis (PPFLT) is exceptional: the few reported cases seem to lack BCL-2 gene rearrangement and/or protein expression. The aim of this study was to characterise a PPFLT arising in a 4 year old boy.

Methods: This case was characterised using conventional histological analysis, immunohistochemistry, and a polymerase chain reaction based method for the detection of immunoglobulin V<sub>H</sub> chain rearrangements.

Results: The neoplasm was staged I<sub>A</sub>; left orchiectomy and chemotherapy were performed, producing complete remission. Histology showed a predominantly follicular lymphoid infiltrate mainly composed of centroblast-like cells. The phenotype was CD20<sup>+</sup>, CD79a<sup>+</sup>, CD10<sup>+</sup>, Bcl-2<sup>+</sup>, B cell specific activating protein<sup>+</sup>,<sup>−</sup> light chain, CD30<sup>−</sup>, interferon regulating factor 4<sup>+/−</sup>, c-myc<sup>−</sup>,<sup>+</sup> light chain, CD3<sup>+</sup>, Bcl-2<sup>−</sup>, p53<sup>+</sup>, cytokeratin<sup>−</sup>, and placental alkaline phosphatase.<sup>+</sup> Lymphomatous elements were found within a CD21<sup>+</sup> follicular dendritic cell network and 70% were positive for Ki-67/MIB-1. Molecu- lard analysis revealed monoclonal immunoglobulin heavy gene chain rearrangements and BCL-6 mutations, in the absence of BCL-2 major breakpoint and BCL-2 minor cluster region rearrangements, p53 mutations, and death associated protein kinase gene hypermethylation.

Conclusions: These findings suggest a different pathogenesis of PPFLT compared with adult follicular lymphoma and might explain its favourable course in spite of aggressive histology.

Follicular lymphoma (FL) is one of the most common lymphomas; it usually occurs in adults, involves lymph nodes, and may finally transform into a diffuse large B cell lymphoma.<sup>1,2</sup> Three morphological grades are recognised,<sup>1,2</sup> namely; grades I and II are indolent tumours that regularly carry the t(14;18) translocation with Bcl-2 protein overexpression; grade III presents aggressively with a greater tendency to evolve into diffuse large B cell lymphoma and can show different chromosomal aberrations, lacking detectable BCL-2 mutations and Bcl-2 protein expression in a minority of cases.<sup>1,12</sup> Paediatric testicular tumours most frequently display germ cell derivation, whereas gonadal stromal neoplasms, rhabdomyosarcomas, leukaemias, and lymphomas are much more rare.<sup>13–15</sup> In general, lymphomas occur in the elderly, with aggressive histology and poor prognosis.<sup>12</sup> Thus, the occurrence of a primary testicular lymphoma in children is noteworthy per se<sup>12,15</sup> and particularly unusual when of the follicular type, as shown by the few reports in the literature.<sup>12,15</sup>

We report an additional example of primary FL of the testis in a 4 year old boy.

CASE REPORT

In September 1999, a 4 year old boy was admitted to another hospital with left testis swelling without other signs and symptoms. On the incision biopsy a diagnosis of possible high grade lymphoma was made. The patient then entered the haematological oncology unit of the paediatric department of Bologna University. No history of infectious diseases was recorded, blood laboratory tests were unremarkable (including lactic dehydrogenase, α fetoprotein, and serology for Epstein-Barr virus, cytomegalovirus, hepatitis C virus, and Mycobacterium tuberculosis). Both total body axial computed tomography and nuclear magnetic resonance showed an enlarged (20 mm across) left testis with non-homogeneous structure of the upper pole and an 8 mm diameter cyst in the left kidney that remained unmodified in the follow up scans. The remaining organs and apparata were unremarkable. Bilateral bone marrow biopsies were slightly hypocellular and disease free; the cerebrospinal fluid was normal. Left orchiectomy and chemotherapy were performed, producing complete remission. Histology showed a predominantly follicular lymphoid infiltrate mainly composed of centroblast-like cells. The phenotype was CD20<sup>+</sup>, CD79a<sup>+</sup>, CD10<sup>+</sup>, Bcl-2<sup>+</sup>, B cell specific activating protein<sup>+</sup>,<sup>−</sup> light chain, CD30<sup>−</sup>, interferon regulating factor 4<sup>+/−</sup>, c-myc<sup>−</sup>,<sup>+</sup> light chain, CD3<sup>+</sup>, Bcl-2<sup>−</sup>, p53<sup>+</sup>, cytokeratin<sup>−</sup>, and placental alkaline phosphatase.<sup>+</sup> Lymphomatous elements were found within a CD21<sup>+</sup> follicular dendritic cell network and 70% were positive for Ki-67/MIB-1. Molecular analysis revealed monoclonal immunoglobulin heavy gene chain rearrangements and BCL-6 mutations, in the absence of BCL-2 major breakpoint and BCL-2 minor cluster region rearrangements, p53 mutations, and death associated protein kinase gene hypermethylation.

Conclusions: These findings suggest a different pathogenesis of PPFLT compared with adult follicular lymphoma and might explain its favourable course in spite of aggressive histology.

Abbreviations: APAAP, alkaline phosphatase antialkaline phosphatase; ISH, in situ hybridisation; MGMT, O6-methylguanine-DNA methyltransferase; PCR, polymerase chain reaction; PLAP, placental alkaline phosphatase; PPFLT, paediatric primary follicular lymphoma of the testis.
Figure 1  (A) The tumour shows a prevalent follicular growth pattern (haematoxylin and eosin stained; original magnification, ×100) and shows a certain degree of fibrosis, as revealed by Gomori silver impregnation for reticulin fibres (inset; original magnification, ×25). (B) At higher magnification, several centroblasts are seen within a neoplastic follicle, which lacks cell polarisation and a mantle zone, looks fibrotic, and contains hyperplastic follicular dendritic cells (one of which is arrowed) (haematoxylin and eosin stained; original magnification, ×400). (C) An area with a diffuse growth pattern; note the residual seminiferous tubule (haematoxylin and eosin stained; original magnification, ×300). The lymphomatous population mostly consists of centroblast-like cells (inset; Giemsa staining; original magnification, ×400). (D) Three neoplastic follicles laying “back to back” show a strong CD20 positivity (APAAP technique; Gill’s haematoxylin nuclear counterstaining; original magnification, ×250). (E) The same follicles display a prominent CD21+ follicular dendritic cell meshwork (APAAP technique; Gill’s haematoxylin nuclear counterstaining; original magnification, ×250). (F) Lymphomatous elements strongly express CD10 (APAAP technique; Gill’s haematoxylin nuclear counterstaining; original magnification, ×300) and the Bcl-6 protein (inset; original magnification, ×250). (G) Most of the tumour cells are in the cell cycle, as revealed by Ki67/Mib-1 staining (APAAP technique; Gill’s haematoxylin nuclear counterstaining; original magnification, ×300). (H) Negative staining of neoplastic follicles for the Bcl-2 protein; those cells that have stained are T cells (APAAP technique; Gill’s haematoxylin nuclear counterstaining; original magnification, ×300). (I) In an area with a diffuse growth pattern, lymphomatous elements strongly express the CD79a molecule and are admixed with some CD21+ follicular dendritic cells (inset) (APAAP technique; Gill’s haematoxylin nuclear counterstaining; original magnification, ×200). (J) No Bcl-2 gene rearrangement is seen; lane 1, molecular weight markers; C1–3, positive controls; N1–2, negative controls; P, formalin-fixed, paraffin wax embedded material; F1–2, frozen tissue.
**Table 1** Main clinical, pathological, and biological features of the cases of primary paediatric follicular lymphoma of the testis quoted in the literature compared with the ones of the reported case

<table>
<thead>
<tr>
<th>Case (ref)</th>
<th>Age (years)</th>
<th>Testis</th>
<th>Tumour size</th>
<th>Stage</th>
<th>Histo</th>
<th>Grade</th>
<th>Treat</th>
<th>Outcome</th>
<th>Bcl-2</th>
<th>CD10</th>
<th>Bcl-6</th>
<th>FL</th>
<th>p53</th>
<th>IgH/R</th>
<th>DAP</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 [13]</td>
<td>8</td>
<td>Left</td>
<td>2.0 cm</td>
<td>F</td>
<td>3</td>
<td>OR</td>
<td>-</td>
<td>NED/44 mo</td>
<td>-</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>2 [14]</td>
<td>3</td>
<td>Left</td>
<td>2.8 cm</td>
<td>F&amp;D</td>
<td>3</td>
<td>OR+CX</td>
<td>NED/18</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>3 [14]</td>
<td>3</td>
<td>Right</td>
<td>2.3 cm</td>
<td>F</td>
<td>3</td>
<td>OR+CX</td>
<td>NED/18</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>Clonal</td>
<td>NA</td>
<td>-</td>
<td>+</td>
<td>NA</td>
</tr>
<tr>
<td>4 [14]</td>
<td>10</td>
<td>Left</td>
<td>4.0 cm</td>
<td>F</td>
<td>3</td>
<td>OR+CX</td>
<td>NED/19</td>
<td>-</td>
<td>NA</td>
<td>+</td>
<td>Clonal</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>5 [14]</td>
<td>15</td>
<td>Right</td>
<td>4.0 cm</td>
<td>F</td>
<td>3</td>
<td>OR+CX</td>
<td>NED/24</td>
<td>-</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>6 [15]</td>
<td>6</td>
<td>Right</td>
<td>3.0 cm</td>
<td>F&amp;D</td>
<td>3</td>
<td>OR+CX</td>
<td>NED/7</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>7 [present case]</td>
<td>4</td>
<td>Left</td>
<td>2.0 cm</td>
<td>F&amp;D</td>
<td>3</td>
<td>OR+CX</td>
<td>NED/26</td>
<td>-</td>
<td>+</td>
<td>Clonal</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>NA</td>
<td></td>
</tr>
</tbody>
</table>

Bcl-2, Bcl-2 protein immunostaining; Bcl-6, Bcl-6 protein immunostaining; BCL-2/R, BCL-2 gene rearrangement; BCL-6/R, BCL-6 gene rearrangement; BCL-6/M, BCL-6 gene somatic mutations; CD10, CD10 immunostaining; CA, chemotherapy; DAP, DAP kinase gene hypermethylation; E, extranodal; F, follicular; F&D, follicular and diffuse; Histo, histology; IgH/M, IgH gene somatic mutations; IgH/R, IgH gene clonal rearrangement; NA, not available; NED, no evidence of disease; OR, orchiectomy; p53, p53 product immunostaining; Treat, treatment; -, negative result; +, positive result.

*Follow up updated in Finn et al.1,2

**DISCUSSION**

In adulthood, testicular lymphoma is not infrequent and mostly corresponds to the diffuse large B cell variety. In childhood, it is rare and when it is seen it usually represents a secondary involvement by an advanced stage Burkitt’s or lymphoblastic lymphoma. Only six reports of primary testicular lymphoma in children are quoted in the literature and in the most up to date ones the tumour was of the FL type (table 1).22–25

"The tumour seems to lack protection against apoptosis and to maintain efficient DNA repair mechanisms"

Our case represents a further example of PPFLT (table 1), which confirms the peculiar clinicopathological characteristics of the tumour, provides new molecular information, and (more in general) contributes to the existing debate on the biology of FL in childhood. The germinal centre cell origin of the tumour was supported by its prevalent follicular aggregation, the expression of the CD10 and Bcl-6 proteins, the detection of tight FDC networks, and the occurrence of somatic mutations of both the IgH and BCL-6 genes. Interestingly, CD10 positivity had previously been recorded in only one

alkaline phosphatase (PLAP), Ki-67 proliferation associated nuclear antigen (antibodies from Dako, Glostrup, Denmark), CD79a (from Professor DY Mason, Oxford, UK), CD68, interferon regulating factor 4 (IRF4), Bcl-6 protein (from Professor B Falini, Perugia, Italy), B cell specific activating protein (BAP) (from Santa Cruz, California, USA), c-Myc, and CD10 (from Novocastra, Newcastle, UK). Antigen retrieval was performed as previously reported.2 Positive and negative controls were always used. The positive neoplastic cells were graded semiquantitatively as: +, 75–100%; +/−, 50–75%; −/−, 25–50%; rare, 1–25%; and −, 0%. The staining intensity and pattern of positivity were recorded. In situ hybridisation (ISH) was performed as reported previously.17 Positive and negative controls were always used. The positive neoplastic cells were graded as described previously.

For molecular studies, DNA was extracted from fixed and frozen material according to previously reported methods and methods of Epstein-Barr virus (EBV) was performed with EBER 1 and 2. The positive neoplastic cells were graded as described previously.18–21

**PATHOLOGICAL FINDINGS**

The upper pole of the left testis was replaced by a tan/grey, fleshy, poorly demarcated tissue without macroscopic infiltration of the tunica albuginea, epididymis, and spermatic cord. Microscopically, the tumour infiltrated the parenchyma with displacement of seminiferous tubules, showing fibrosis, and mainly growing in “back to back” follicles, without a well defined mantle zone or germinal centre polarisation (fig 1A,B,D); some germinal centres looked fibrotic with follicular dendritic cell (FDC) hyperplasia (fig 1B). Areas with diffuse growth accounted for 25% of the neoplasm (fig 1C). Most tumoral cells were large with centroblastic morphology, although multilobular or centrocytic nuclei were seen (fig 1B,C). Mitoses were numerous. The tunica albuginea, epididymis, spermatocord, and the right incisional biopsy were negative.

Lymphomatous elements were strongly positive for CD20, CD79a, CD10, BAP, and CD68 protein with monotypic light chain Ig restriction and a MIB-1 value of 70% (fig 1D,F,G,I). Approximately 10% of neoplastic cells expressed CD30, IRF4, and c-Myc. Bcl-2, p53, cytokeratins, and PLAP were negative; the anti-CD21 antibody showed tight FDC networks and loosely dispersed FDCs in the follicular and diffuse areas, respectively (fig 1E,I). The reactive T cells admixed were positive for CD3 and Bcl-2 (fig 1H). EBV integration was not demonstrable by ISH.

Analysis of the IgH genes revealed the presence of a single PCR product, corresponding to a monoclonally rearranged V\_H gene. The DP-7/21-2 of the V\_H gene family was productively rearranged with the D2-15/D2 D segment and the JH6b gene. The V\_H gene utilised was somatically mutated, at a relatively low frequency (4.7 × 10^-3 bp). BCL-6 mutations in a heterozygous condition were detected at a frequency of 4.1 × 10^-3 bp at nucleotides 1069 (A → C), 1071 (A → C), 1072 (A → G), 1075 (C → A), 1085 (C → A), and 1109 (T → C). No p53 mutations or BCL-2 rearrangements were detected. Frozen and routine samples showed identical results (fig 1J). No methylation of the MGMT, DAP kinase, or p73 genes was observed.

Based on these findings, a diagnosis of peripheral B cell lymphoma, follicular, grade 3/3 (75%) with diffuse large B cell lymphoma (25%) was made, which was confirmed at the Workshop of the European Association for Haematopathology held in London, May 7–11, 2000.
instance11 (table 1): the variable results of the search for this molecule (which in conjunction with the Bcl-6 protein is a reliable marker of normal germinal centre B cells and germinal centre cell derived lymphomas)12 might result from the usual high grade histology of PPFLT (table 1). In fact, negative or weak CD10 expression has recently been detected in 83% of grade III Fls.12 The presence of somatic mutations of the IgVs and BCL-6 genes, not analysed in previous series (table 1), provides further evidence of the germinal centre cell derivation of the tumour. In fact, these mutations characteristically occur in antigenically stimulated B cells, which enter the germinal centre, carry out Ig class switching, rearrange their Ig V region to recognise specific antigenic determinants, and undergo selection based on the affinity of the antibodies produced.12–20

In agreement with previous reports11–15 (table 1), our case was molecularly characterised by the lack of BCL-2 gene rearrangement and p53 mutations, along with negativity for the corresponding protein products. In addition, our patient did not show MGMT, DAP kinase, or p73 gene hypermethylation.16–21 These findings—possibly in conjunction with the limited extent of the disease (all patients having been staged F)—might explain the excellent therapeutic response of PPFLT, in spite of the high grade histology and the detection of a diffuse component in three instances (table 1).

In fact, the tumour seems to lack protection against apoptosis and to maintain efficient DNA repair mechanisms. Furthermore, the results suggest that PPFLT and childhood FL in general might have a different pathogenesis to FL in adults. In particular, 83–100% of grade I/II and approximately 75% of grade III adulthood FLs carry the t(14;18)(q32;q21) translocation, with consequent BCL-2 gene rearrangement, Bcl-2 protein overexpression, and protection of neoplastic cells from apoptosis.14–21 Because of this, Bcl-2 staining is often used for the differential diagnosis between FL (Bcl-2+) and follicular hyperplasia (Bcl-2-). In adult FL, apoptosis is also compromised by the inactivation of the proapoptotic DAP kinase gene as a result of promoter hypermethylation.18 In addition, alterations of the p53 tumour suppressor gene, and BCL-6 gene rearrangements and/or the accumulation of mutations in its 5′ non-coding region seem to be involved in grade III adult FL development and/or in the transformation of FL into a diffuse large B cell tumour.14–17,19–21 With the exception of BCL-6 gene rearrangement, which was seen in one case by Finn et al.,19 and the prognostic value of which is debatable,22 none of these molecular alterations affecting tumour resistance and progression has so far been seen in PPFLT (table 1). In particular, the lack of BCL-2 gene rearrangement and Bcl-2 protein expression might not be exclusive to PPFLT, but also be a characteristic feature of childhood FL. In fact, Lorsbach et al.14 have recently reported their absence in 12 of 17 FLs in children, all of which occurred in patients younger than 12 years.24 Interestingly, similar to PPFLT, most of Lorsbach’s patients had high grade FL, were stage I, and had an excellent outcome, with the exception of the BCL-2 positive patients, who died of their disease or required aggressive treatment with peripheral blood stem cell transplantation. Based on these findings, Lorsbach et al.14 concluded that “in most instances FL in children has a pathogenesis that is distinct from its counterpart in the adult population.”22

Finally, PPFLT should always be differentiated from follicular hyperplasia in the course of chronic orchitis. Because the lack of Bcl-2 protein expression and BCL-2 gene rearrangement can make the diagnosis of FL less obvious, a series of parameters should always be considered, including the absence of previous infectious diseases, negativity for EBV at ISH, the lack of inflammation or granulomas in the epithidymis and spared testis, back to back follicular growth pattern, non-polarised germinal centres, almost exclusive B cell antigen expression, Ig light chain restriction, and the detection of a monoclonal Ig gene rearrangement. However, it should be emphasised that the absence of monoclonal Ig gene rearrangement using conventional PCR analysis on routine material does not exclude the diagnosis of PPFLT; in fact, a negative result was recorded in one of the reported cases (table 1), which might have been produced by the limitations of amplifying DNA extracted from paraflin wax blocks14 and/or the need for a large series of primers to detect rare forms of rearrangement, such as the one found in our case.

Furthermore, the results suggest that paediatric primary follicular lymphoma (FL) of the testis and childhood FL in general might have a different pathogenesis to FL in adults.

In conclusion, further studies based on the newly developed microarray techniques14 are needed to establish firmly whether childhood FL is indeed pathogenetically different from the adult form, irrespective of the primary site.

ACKNOWLEDGEMENTS

This paper was supported by grants from AIRC (Milan), MURST (Rome), and Fondazione “Piera Pietro e Giovanni Ferrero” (Alba). The authors thank Mr L. Chilli and Mrs F. Sandri for their skilled technical assistance.

Authors’ affiliations

S A Pileri, E Sabattini, P L Zinzani, S Ascani, G Fraternali-Orcioni, B Gamberti, M Piccioli, Pathology and Clinical Units of the Institute of Haematology and Clinical Oncology “L. and A. Seràgnoli”, University of Bologna, 40138 Bologna, Italy
P Rosito, Unit of Paediatric Haematology and Oncology, Department of Paediatrics, University of Bologna, Italy
D Vivenza, G Gaidano, Division of Internal Medicine, Department of Medical Sciences, University of Eastern Piedmont “Amedeo Avogadro”, 28100 Novara, Italy
B Falini, Haematopathology Laboratory, Institute of Haematology, University of Perugia, D6100 Perugia, Italy

REFERENCES

Want full text but don't have a subscription?

Pay per view

For just $8 you can purchase the full text of individual articles using our secure online ordering service. You will have access to the full text of the relevant article for 48 hours during which time you may download and print the pdf file for personal use.

www.jclinpath.com