How should stainable iron in bone marrow films be assessed?

D A Hughes, S E Stuart-Smith, B J Bain

Aims: To identify how many particles should be examined to enable a confident assessment of the presence or absence of iron stores and the quantity of iron in a bone marrow aspirate to be made.

Methods: One hundred and ninety consecutive bone marrow aspirate samples were stained with Perls' stain and the iron content of 10 consecutive particles was recorded. The first particle found to be positive and the particle that was most positive were also noted.

Results: A minimum of seven particles must be examined to establish the absence of stainable iron. A minimum of nine particles must be reviewed to see the maximum iron stores in 100% of samples and therefore make a valid judgment of whether iron stores are reduced, normal, or increased. By these criteria, 46% of the samples tested here could not be optimally assessed for absence of iron or maximum iron stores.

Conclusions: The sensitivity of examination of bone marrow aspirates for iron stores can be optimised by increasing the number of particles reviewed to seven or more. This may require the staining of additional slides.

**METHODS**

**Selection of samples**

One hundred and ninety consecutive bone marrow aspirates obtained over a period of 12 months in the department of haematology, St Mary's Hospital, London, were selected for the assessment of iron stores. Bone marrow aspirates were obtained in the course of investigation of a range of haematological conditions, including anaemia and haematological malignancy.

**Perls' stain for iron**

Bone marrow was aspirated from the posterior iliac crest or, occasionally, from the sternum. The aspirate was spread on to glass slides, air dried, and fixed with methanol. Slides were stained in 10 g/litre potassium ferrocyanide for 10 minutes before being washed in tap water for 20 minutes and counterstained in 0.1% aqueous safranin.

**Assessment of iron stores**

The number of bone marrow particles available for each marrow sample was assessed. Where fewer than 10 particles were present on a single slide, further slides were stained to optimise the number of particles, wherever possible, to 10 or more. Each particle was assessed in turn and the iron content in 10 consecutive particles was recorded. The first iron

<table>
<thead>
<tr>
<th>Grade</th>
<th>Stainable iron</th>
<th>Results in 50 normal patients</th>
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<tbody>
<tr>
<td>0</td>
<td>None</td>
<td>0</td>
</tr>
<tr>
<td>1</td>
<td>Very slight</td>
<td>10</td>
</tr>
<tr>
<td>2</td>
<td>Slight</td>
<td>36</td>
</tr>
<tr>
<td>3</td>
<td>Moderate</td>
<td>4</td>
</tr>
<tr>
<td>4</td>
<td>Moderately heavy</td>
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</tr>
<tr>
<td>5</td>
<td>Heavy</td>
<td>8</td>
</tr>
<tr>
<td>6</td>
<td>Very heavy</td>
<td>6</td>
</tr>
</tbody>
</table>

"We aimed to answer the question of how many particles should be examined so that a confident assessment of the presence or absence of iron stores can be made."
Iron stores were graded according to the method of Rath and Finch (table 1).

RESULTS
Evaluation of bone marrow aspirate specimens for suitability for assessment of iron status
Of 190 consecutive bone marrow aspirate specimens, 87 (45.7%) contained 10 or more particles, 43 (22.6%) had no particles, and 60 (31.5%) had between one and 10 particles.

Figure 1 shows the distribution of the number of particles in each specimen. This distribution is polarised, with most preparations found to be completely aparticulate or to have an adequate number of particles. For the initial assessment, only samples with 10 or more particles were reviewed.

Minimum numbers of aspirate particles required to establish a diagnosis of iron deficiency
Eighty seven bone marrow aspirates with 10 or more particles were stained by Perls' method for iron and the iron content of each consecutive particle was assessed. The first particle to show an iron grading of 1 or more was recorded. Figure 2 shows the cumulative distribution of samples becoming positive with each consecutive particle. The distribution plateaus at seven particles, when 90% of the sample shows at least one particle positive for iron. No further samples became positive even when 10 particles were reviewed. This indicates that seven particles must be assessed before a confident report of zero iron stores can be made.

Of our original 190 specimens, 43 had no particles (22.6%) and 12 (6.3%) had one to six particles without stainable iron and, therefore, using these new criteria should be considered not assessable.

Minimum number of aspirate particles required to establish the maximum iron status of a sample
The 87 bone marrow aspirate samples with 10 or more particles were then assessed to determine the particle exhibiting the maximum iron staining (fig 3). This distribution plateaus at nine particles, with no further increase in the grading of iron stores achievable with the 10th particle. The mean (SD) variation in iron content between particles in individual patients was 1.33 (0.9) units. Most (98.7%) samples had reached their maximum positivity by the seventh sample. Of the original 190 samples, 33 (17.3%) had visible iron but had only one to six particles and could not therefore be optimally graded for maximum positivity.

DISCUSSION
Our study shows that for the optimum assessment of a bone marrow aspirate for the absence of iron or for maximum iron stores a minimum of seven particles is required. Any sample that has fewer than seven particles and that contains no particles with stainable iron cannot be confidently said to have absent iron stores. This may explain why other studies have reported discrepancies between the bone marrow aspirate iron and the clinical status of the patient. A total of 46% of the samples in our study would have been considered not optimally assessable: 22.6% were aparticulate, 6.3% had only one to six particles and were negative, and 17.3% had only one to six particles but were positive. We were surprised that nearly a quarter of the samples were aparticulate. We suspect that this is an increasing phenomenon related to the demands for samples to be available for immunophenotyping, cytogenetic and molecular genetic analysis, and other specialised tests. We would always advocate the use of a small syringe to obtain a sample for the preparation of films before other diagnostic samples are taken; 0.2–0.3 ml is generally sufficient for this purpose. If it is clear that particles are not numerous, excess blood should be drained off before spreading further films. If the initial slide stained does not contain storage iron, further slides...
should be selected to give a minimum of seven fragments. Although the clinical importance of discriminating between grades 5 and 6 iron storage may be limited, it is essential to be able to assess marrow specimens for absent iron stores when investigating anaemia. An attempt should be made to optimise the number of particles to seven; if necessary, more than one slide should be stained and reviewed. If this cannot be achieved and no iron is seen on the existing particles, we suggest that the sample should be reported as lacking stainable iron, but the assessment should be stated to be unreliable. However, if a small initial sample was obtained for film preparation, it is likely that a definitive report could be issued on a single film much more often.

“...we were surprised that nearly a quarter of the samples were aparticulate”

If fewer than seven particles are available for assessment in a thus far negative bone marrow, the examination of bone marrow trephine biopsy sections for haemosiderin might be helpful. This will be the subject of a further study.

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REFERENCES