MUC4 is increased in high grade intraepithelial neoplasia in Barrett’s oesophagus and is associated with a proapoptotic Bax to Bcl-2 ratio

D A Bax, J Haringsma, A W C Einerhand, H van Dekken, P Blok, P D Siersema, E J Kuipers, J G Kusters

ORIGINAL ARTICLE

Background: Patients with Barrett’s oesophagus (BO) are at risk of oesophageal adenocarcinoma. Because the pattern of mucosal mucins changes during neoplastic progression, it may serve as a marker of intraepithelial neoplasia.

Aims: To determine the expression pattern of mucins in neoplastic BO epithelium (high grade dysplasia) and correlate it with the expression of apoptosis markers Bax and Bcl-2.

Methods: Thirty seven patients with BO were studied: 16 without intraepithelial neoplasia, six with high grade intraepithelial neoplasia (HGN), and 15 with infiltrating adenocarcinoma. Biopsies were obtained from squamous epithelium, Barrett’s epithelium, and (when present) foci of suspected HGN or adenocarcinoma. MUC1–4, MUC5AC, MUC5B, MUC6, Bax, and Bcl-2 mRNA were determined by semiquantitative RT-PCR. MUC2, MUC5AC, and MUC6 protein was determined by immunoblotting.

Results: Mucin expression varied between neoplastic progression stages in BO. Mucin mRNA levels were low in squamous epithelium, except for MUC4, and were at least four times higher in BO and HGN (p<0.001), but less so in adenocarcinoma. MUC4 expression was significantly lower in BO than in normal squamous epithelium, whereas in HGN and adenocarcinoma, levels were significantly higher than in BO (p=0.037). The Bax:Bcl-2 ratio was increased in HGN compared with BO (p=0.04). MUC2, MUC5AC, and MUC6 protein values correlated with mRNA data.

Conclusions: Mucin expression varies during the development of oesophageal adenocarcinoma in BO. MUC4 could serve as a tumour marker in this process. In contrast to animal studies, upregulation of MUC4 in HGN is associated with increased apoptosis, suggesting that MUC4 plays a minor role in apoptosis regulation in BO.

The use of molecular markers in addition to normal endoscopical and histological evaluation could significantly enhance the detection of neoplasia in BO, both in vitro and in vivo. In particular, markers that indicate the presence of HGN could be helpful in the identification of patients at risk of malignant transformation.

Mucins are large glycoproteins that are the main components of the gel-like mucous layer on the surface of the intestine. This layer serves to protect the mucosa from mechanical damage. Each type of mucosa has a unique pattern of mucin expression, and this pattern changes during neoplastic progression. Therefore, mucins are putative molecular markers for the development of adenocarcinoma in BO. Currently, more than 10 MUC genes have been reported, the best characterised being MUC1–4, MUC5AC, MUC5B, and MUC6. Of these, only MUC1 and MUC4 are expressed in the normal squamous lining of the oesophagus, whereas BO predominantly expresses MUC2, and to a lesser extent MUC5AC and MUC6.

In addition to protection against mechanical damage, membrane bound mucins MUC1 and MUC4 have also been reported to be involved in oncogenic pathways. MUC1 can bind to β catenin, thereby altering its ability to bind to E-cadherin, which leads to decreased cell–cell adhesion and...
uncontrolled proliferation. MUC4, a mucin that is upregulated in several tumour types, acts as a ligand for the receptor tyrosine kinase, Erb-B2, thereby regulating the p27kip cell cycle inhibitor. MUC4 also inhibits apoptosis in a xenotransplanted melanoma model in nude mice.

The aim of our study was to evaluate the expression patterns of MUC1–4, MUC5AC, MUC5B, and MUC6 in high grade intraepithelial neoplasia and adenocarcinomas in BO, and to determine the association with the apoptosis markers Bax and Bcl-2.

METHODS

Patients

In total, 37 patients with BO were studied. Sixteen patients had BO without intraepithelial neoplasia, six patients had BO with high grade intraepithelial neoplasia, and 15 had an infiltrating adenocarcinoma. Patients were evaluated by standard oesophagoscopy and high magnification chromendoscopy. Biopsies were obtained from the squamous epithelium, Barrett’s epithelium, and (when present) from foci of suspected high grade intraepithelial neoplasia or adenocarcinoma. Two expert gastrointestinal pathologists independently evaluated haematoxylin and eosin stained sections for the presence of BO, grade of intraepithelial neoplasia, and ADC. All patients diagnosed with high grade intraepithelial neoplasia routinely underwent endomucosal resection. Endomucosal resection specimens were independently analysed by two pathologists to ensure that there were no indications for the presence of an (infiltrating) adenocarcinoma. Ten patients with adenocarcinoma were treated surgically and five patients received palliative treatment. All adenocarcinomas had infiltrated into the submucosa and three patients had local lymph node metastases. Eight tumours were well differentiated and seven were poorly differentiated. Our study was approved by the ethical committee of the Erasmus MC Rotterdam, The Netherlands, and informed consent was obtained before endoscopy.

RNA and protein isolation from biopsies

Biopsies were snap frozen in liquid nitrogen. Total RNA was isolated using TRIzol reagent (Invitrogen, Groningen, The Netherlands), according to the manufacturer’s instructions. The remaining chromosomal DNA was removed with the RNA free DNA kit (Zymo, Orange, California, USA). Total RNA concentrations were estimated by RNA electrophoresis on an agarose gel and band intensity was compared with that of a marker band with a known concentration. From the same biopsy specimen, a protein fraction was isolated according to the manufacturer’s instructions and denatured in 1% sodium dodecyl sulfate. Protein concentrations were determined with the BCA protein assay kit (Pierce, Rockford, Illinois, USA).

<table>
<thead>
<tr>
<th>Table 1 Polymerase chain reaction primers</th>
</tr>
</thead>
<tbody>
<tr>
<td>Amplified gene</td>
</tr>
<tr>
<td>----------------</td>
</tr>
<tr>
<td>MUC1</td>
</tr>
<tr>
<td>MUC2</td>
</tr>
<tr>
<td>MUC3</td>
</tr>
<tr>
<td>MUC4</td>
</tr>
<tr>
<td>MUC5B</td>
</tr>
<tr>
<td>MUC5AC</td>
</tr>
<tr>
<td>MUC6</td>
</tr>
<tr>
<td>Bax</td>
</tr>
<tr>
<td>Bcl-2</td>
</tr>
<tr>
<td>β-actin</td>
</tr>
</tbody>
</table>

Reverse transcriptase polymerase chain reaction
cDNA was synthesised with the use of avian myeloma virus reverse transcriptase (RT; Promega, Madison, Wisconsin, USA). Primers were annealed by cooling down from 70°C to room temperature. cDNA was synthesised by incubation of the mRNA for 30 minutes at 42°C. Polymerase chain reactions (PCRs) (total volume of 25 µl) contained 1 µl of the cDNA solution, 1× PCR core buffer (Promega), 2mM magnesium
chloride, 0.4 mM forward and reverse primer, 200 mM of each nucleotide (Promega), and 0.02 U/μl Taq polymerase (Promega). PCR conditions were 35 cycles at 94°C for 30 seconds, 55°C for 30 seconds, and 72°C for one minute. PCR products were visualised on a 1% agarose gel and stained with ethidium bromide. Band size and intensity were measured with Kodak 1D software (Kodak, Rochester, Minnesota, USA) version 3.5 by normalising against the

Figure 2. Mean relative expression of (A) MUC1, (B) MUC2, (C) MUC3, (D) MUC4, (E) MUC5AC, (F) MUC5B, and (G) MUC6. The measurement of mucin mRNA was performed by normalising the band area and intensity with that of β-actin. Sq, squamous epithelium (n = 32); BO, Barrett’s oesophagus (n = 30); HGN, high grade intraepithelial neoplasia (n = 6); and ADC, adenocarcinoma (n = 15).

Mean relative expression of (A) MUC1, (B) MUC2, (C) MUC3, (D) MUC4, (E) MUC5AC, (F) MUC5B, and (G) MUC6. The measurement of mucin mRNA was performed by normalising the band area and intensity with that of β-actin. Sq, squamous epithelium (n = 32); BO, Barrett’s oesophagus (n = 30); HGN, high grade intraepithelial neoplasia (n = 6); and ADC, adenocarcinoma (n = 15).
housekeeping gene β actin, as described previously. Table 1 lists the primers used.

**Protein crossblot**

Aliquots of 20 μg of protein were loaded in the lanes of a reverse line blotter (Immunetics, Cambridge, UK) on a PVDF membrane (Roche, Mannheim, Germany). After one hour of incubation at room temperature the samples were removed and the membrane was blocked with 5% Protifar non-fat milk (Nutricia, Cuijk, the Netherlands) in phosphate buffered saline containing 0.1% Tween. The primary antibodies anti-MUC2 (clone Cp58; Novocastra, Newcastle, UK), anti-MUC5AC (clone 45M1; Zymed, San Francisco, California, USA), and anti-MUC6 (clone CLH5; Novocastra) were loaded at an angle of 90° on to the protein samples. An alkaline phosphatase labelled goat anti-mouse IgG antibody (Southern Biotech Associates, Birmingham, Alabama, USA) was used as conjugate and 0.18 mg/ml 5-bromo-4-dichloro-indolyl phosphate (BCIP; Promega) and 0.35 mg/ml nitrobleue tetrazolium (NBT; Promega) in 0.1M Tris/HCl (pH 9.5), 0.1M NaCl, and 5 mM MgCl₂ were added as substrates. Immunohistochemical staining of gastric antrum and colonic mucosa confirmed that the primary antibodies detected the mature mucin protein in the case of MUC5AC and MUC6, or a precursor peptide, in the case of MUC2.

**Statistical analyses**

RT-PCR data are presented as mean (SEM). Statistical analyses were performed with the Mann-Whitney U test. A p value < 0.05 was considered significant.

**RESULTS**

**Mucin mRNA**

Figure 1 shows the results of RT-PCR testing of different mucins in squamous epithelium, BO, HGN, and ADC. Figure 2 shows the mean transcription levels of individual mucins relative to the transcription of β actin in the same sample. Overall, mucin transcription was low or absent in squamous epithelium (relative transcription level, < 0.2), except for MUC4 (relative transcription level, 1.1). Compared with squamous epithelium, MUC4 was downregulated in BO (relative transcription level, 0.62; p = 0.01), whereas the other mucins tested were considerably upregulated in BO (relative transcription level, > 0.4; p < 0.001). In HGN, mucin transcription levels were similar to those seen in BO, except for MUC4, which was significantly higher in HGN than in BO (relative transcription levels 1.27 v 0.62; p = 0.037). In most adenocarcinomas, the mucin mRNA expression was lower than in BO without intraepithelial neoplasia or with HGN. Amplification of residual chromosomal DNA was not found. Sequencing and comparison with previous published sequences confirmed the correct nature of the mucin PCR products.

**Mucin protein expression**

Protein fractions from the same biopsies as the mRNA fractions were analysed by protein crossblot to determine the mucin patterns at the protein level (fig 3). Commercial antibodies that recognise the denatured antigen were only available for MUC2, MUC5AC, and MUC6. Crossblot analysis revealed that in squamous epithelium MUC2 expression was preserved in two of 32 patients, for MUC5AC in nine of 32 patients, and for MUC6 in two of 32 patients. In the BO samples, MUC2 was expressed in 17 of 30 patients, MUC5AC in 28 of 30 patients, and MUC6 in 26 of 30 patients. In HGN, MUC6 was expressed in 15 of 30 samples, whereas expression of MUC2 and MUC5AC was comparable to that seen in BO (three of six and six of six, respectively). MUC5AC and MUC6 protein were found in nine of 15 and MUC2 protein in three of 15 of the ADC samples (fig 4). Protein data were found to correlate with the mRNA data.

**Correlation between Bax and Bcl-2 mRNA and mucin mRNA expression**

The correlation between mucin mRNA expression and the apoptosis markers Bax and Bcl-2 was determined by calculating the ratio of prosapoptotic Bax and antiapoptotic Bcl-2 mRNA. The amounts of Bax and Bcl-2 mRNA were determined by semiquantitative RT-PCR (fig 5). The Bax : Bcl-2 ratio was increased in HGN compared with BO (ratio 1.71 v 0.83; p = 0.04). In ADC, the ratio was lower than in HGN (0.95), but this difference was not significant (p = 0.22). The raised Bax : Bcl-2 ratio in HGN was associated with an increase in MUC4 mRNA.

**DISCUSSION**

Changes in the expression pattern of mucins have been associated with the development of early carcinomas of the colon, gallbladder, and the respiratory tract. The expression pattern of mucins in oesophageal adenocarcinoma has also been determined; however, the pattern of mucin expression in high grade intraepithelial neoplasia, the lesion that precedes infiltrating adenocarcinoma, has not been clarified. In our study, the transcription and expression of various mucins in biopsies of oesophageal squamous epithelium, BO, HGN, and ADC were determined. From each biopsy sample, both RNA and protein were purified, thus minimising sample discordance between RNA and protein analysis. Most mucins were upregulated in BO compared with squamous epithelium. In adenocarcinomas, a greater variety, with a trend towards lower mucin expression compared with BO, was seen. Compared with non-dysplastic BO, HGN contained higher amounts of MUC4 mRNA.

High expression of the secreted mucins MUC2, MUC5AC, MUC5B, and MUC6 has been reported previously in BO, as has their downregulation in adenocarcinoma of the oesophagus. In contrast to these reports, low amounts of these mucins were found in some squamous epithelial samples. Because RT-PCR is a very sensitive technique, these low amounts could have escaped detection in immunohistochemical studies.

However, the possibility that the squamous samples were contaminated with intestinal glands or subepithelial proper glands cannot be excluded. Adjacent biopsies were analysed for the purity of the epithelium because approaches designed for the extraction of mRNA from fixed tissue used for histological analysis have not been validated for the
quantitative comparison of mRNA. Furthermore, the presence of the MUC2, MUC5AC, and MUC6 proteins in the squamous epithelium could have been caused by reflux of gastric mucous components, including these mucins.

Most reports describe high expression of MUC1 in squamous epithelium and low expression in BO, whereas the opposite is shown in our present study. An explanation for this could be that most studies used unrelated squamous epithelium samples, whereas in our study paired squamous/BO samples from a single patient were used in which inflammation could have altered the mucin pattern. However, the most striking result is the upregulation of MUC4 in HGN compared with BO without intraepithelial neoplasia. This has not been reported before.

Duodenogastro–oesophageal reflux has been implicated in the development of BO and oesophageal adenocarcinoma.

Increased MUC4 expression has also been found in other early neoplastic lesions, such as early pancreatic carcinomas and dysplastic cervical lesions. When produced in normal quantities, MUC4 has been reported to function as a steric barrier that limits the access of other cells and large molecules to the epithelium, thereby protecting it from damage. In contrast, overexpression of MUC4 disrupts cell–cell and cell–matrix interactions, which could initiate the dissociation of tumour cells from the primary tumour. Finally, overexpression of MUC4 may result in the masking of tumour antigens, thereby effectively suppressing tumour cell killing by cytotoxic lymphocytes. Because oesophageal adenocarcinoma often metastasizes at an early phase, loss of cell adhesion probably occurs at an early stage of carcinogenesis. Increased MUC4 expression could contribute to the detachment of tumour cells from the primary adenocarcinoma.

MUC4 protein also acts as a ligand for the receptor tyrosine kinase, Erb-B2, a protooncogene that is abnormally expressed in the membranes of Barrett’s epithelium and associated adenocarcinoma. The MUC4–Erb-B2 complex is involved in the inhibition of apoptosis, thereby promoting tumour growth. Strong suppression of apoptosis was seen in MUC4 overexpressing tumours in mice. In our study, the mRNA levels of the apoptosis markers Bax and Bcl-2 were also determined. Inhibition of apoptosis was not found in the HGN samples with increased MUC4 expression. In contrast, the Bax : Bcl-2 ratio was increased in HGN compared with BO. This might indicate that MUC4 plays only a minor role in the regulation of apoptosis in HGN in humans.

An increased apoptotic rate does not fit in with the model of neoplastic progression. However, reduced Bcl-2 expression during the neoplastic progression of BO has been reported before. A recent report described clear Bax and Bcl-2 expression in both BO, HGN, and ADC, but results were not quantitative. Other apoptotic proteins, such as the Fas ligand and caspases, may have an antiapoptotic effect in BO with HGN, to compensate for the proapoptotic Bax : Bcl-2 balance.
In conclusion, the results of our study show that mucin expression is altered in BO. Membrane bound mucin MUC4 is associated with the development of high grade intraepithelial neoplasia in BO, but not with a decrease in apoptosis. This suggests a role for MUC4 as an early tumour marker in BO. In combination with other tumour markers, it could facilitate the early diagnosis of adenocarcinoma both in vitro and in vivo. However, additional studies are necessary to verify this hypothesis.

ACKNOWLEDGEMENTS

The authors acknowledge I Moons, A van Vliet, K Kwkeeboom, and J Dekker for helpful and stimulating discussions during the preparation of the manuscript.

Authors’ affiliations

D A Bax, J Haringsma, P D Siersma, E J Kuipers, J G Kusters, University Medical Centre, 3015 GD Rotterdam, The Netherlands

J Dekker for helpful and stimulating discussions during the

A W C Einerhand, Laboratory of Paediatrics, Erasmus MC – Sophia Children’s Hospital, Rotterdam, The Netherlands

H van Dekken, Department of Pathology, Erasmus MC – University Medical Centre, Rotterdam, The Netherlands

P Blok, Department of Pathology, Leyenburg Ziekenhuis, The Hague, The Netherlands

REFERENCES


22 Warner C, Van De Bovenkamp Jr, Kartelend Van Male AM, et al. Barrett’s esophagus is characterized by expression of gastric-type mucins (MUC5AC, MUC6) and TFF peptides (TFF1 and TFF2), but the risk of carcinoma development may be indicated by the intestinal-type mucin, MUC2. Hum Pathol 2002; 33: 660–8.


