CASE REPORT

Transition of Sézary syndrome into mycosis fungoides after complete clinical and molecular remission under extracorporeal photopheresis

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Mycosis fungoides (MF) and Sézary syndrome (SS) are the most common clinical variants of cutaneous T cell lymphoma. Although thought to be closely related to mature T helper cells, the relation between the neoplastic cells in MF and SS is still not fully clarified. This report describes a patient with complete remission of SS under treatment with extracorporeal photopheresis (ECP), who subsequently developed typical plaques of MF and large cell lymphoma (LCL). Serial polymerase chain reaction analyses confirmed identical T cell receptor (TCR) gene rearrangements in SS, MF, and LCL, and complete disappearance of the circulating malignant T cell clone from the peripheral blood after ECP. These findings indicate that the neoplastic cells in SS, MF, and LCL are derived from a common precursor T cell, despite the change in clinical phenotype.

Primary cutaneous T cell lymphoma (CTCL) includes a heterogeneous group of lymphoproliferative disorders characterised by skin homing of malignant T cells. Mycosis fungoides (MF) and Sézary syndrome (SS) are the two major clinical variants of CTCL, both characterised by epidermotropic infiltrates of clonal malignant lymphocytes with cerebriform nuclei. SS shares many of the morphological and biological abnormalities of MF, including nuclear atypia, a mature T cell-like immunophenotype, frequent T cell receptor (TCR) gene rearrangements, and chromosomal aberrations, including PTEN gene abnormalities. However, SS shows some differences compared with MF, such as loss of the cutaneous lymphocyte antigen skin homing antigen that controls the ability of the malignant cells to circulate in the peripheral blood, and predominant secretion of T helper type 2 (Th2) cytokines. It is still a matter of controversy whether SS is simply a leukaemic phase of MF or a separate entity within the spectrum of CTCL.

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Here, we describe a patient with SS, who initially experienced complete remission under extracorporeal photopheresis (ECP), but then developed plaques and nodular lesions of MF, with subsequent transformation into large cell lymphoma (LCL). To clarify whether the malignant T cells of SS, MF, and LCL were derived from the same neoplastic precursor or, alternatively, corresponded to independent T cell clones, we analysed their TCR β and γ rearrangements by means of the polymerase chain reaction (PCR), GeneScan analysis, and subsequent DNA sequencing, and found that the different clinical phenotypes resulted from a common neoplastic cell precursor.

CASE REPORT

A 72 year old man presented to the dermatology department, Charité-Universitätsmedizin, Berlin, Germany, with generalised erythroderma, palm and plantar hyperkeratosis, enlarged lymph nodes, and recalcitrant itching (fig 1A). The patient had raised white blood cell counts (between 27 x 10⁹/litre and 21 x 10⁹/litre) with large numbers of Sézary cells (32–52%).

A skin biopsy revealed a moderately dense, perivascular lymphohistiocytic infiltrate in the upper dermis, with single cell epidermotropism of atypical lymphocytes showing hyperchromatic, convoluted nuclei (fig 1D). Histological examination of a palpable lymph node showed infiltration of atypical small and large lymphocytes with a cerebriform appearance. Serological studies for human T lymphotropic virus 1 and human immunodeficiency virus were negative. Flow cytometric immunophenotyping of peripheral blood lymphocytes revealed a CD3⁺CD4⁺CD7⁻CD8⁻ immunophenotype. Clonal TCR β and γ rearrangements of identical size and sequence were detected in all specimens from the skin, lymph node, and the peripheral blood. Further investigations excluded involvement of visceral organs or the bone marrow. All diagnostic criteria for typical SS according to the current World Health Organisation classification and the consensus conference of the International Society for Cutaneous Lymphomas were met.

Treatment with extracorporeal photopheresis, comprising two consecutive sessions every four weeks, resulted in complete resolution of the skin symptoms after six months, including remission of the lymph node enlargement and disappearance of the circulating Sézary cells. Furthermore, clonal TCR β and γ gene rearrangements were no longer detectable in the peripheral blood, indicating that the malignant T cell clone had disappeared. However, three months later the patient developed typical plaques and papulonodular lesions on his trunk (fig 1B). Histopathological examination showed features of MF with a band-like infiltrate in the upper dermis, atypical T cells with cerebriform nuclei, and the formation of intraepidermal microabcesses (Pautrier abscesses) (fig 1E). No extracutaneous involvement was found, including peripheral blood, where atypical cells were absent. In addition, the patient received 35 phototherapy sessions (psoralen and ultraviolet A irradiation of the skin) leading to major clinical improvement.

Abbreviations: CTCL, cutaneous T cell lymphoma; ECP, extracorporeal photopheresis; LCL, large cell lymphoma; MF, mycosis fungoides; PCR, polymerase chain reaction; SS, Sézary syndrome; TCR, T cell receptor; Th, T helper cell
Seventeen months later the patient developed progressive disease with multiple skin tumours, generalised lymphadenopathy, and infiltration of the bone marrow, liver, and tonsils (fig 1C). Histology revealed (D) an upper band-like infiltrate with epidermotropism of atypical lymphocytes and (E) a typical Pautrier microabscess (E). (F) LCL showing sheets of blast cells.

RESULTS AND DISCUSSION

Although MF and SS are generally thought to belong to the group of malignant CTCLs, their relation to each other has

Figure 1 (A) Typical erythroderma of Sézary syndrome, and (B) solitary mycosis fungoides plaque, (C) tumour nodule of large cell lymphoma (LCL). Histology reveals (D) an upper band-like infiltrate with epidermotropism of atypical lymphocytes and (E) a typical Pautrier microabscess (E). (F) LCL showing sheets of blast cells.

Figure 2 Putative model for the development of different cutaneous T cell lymphoproliferations: (A) linear transition of Sézary syndrome (SS) to mycosis fungoides (MF) and subsequently to large cell lymphoma (LCL), thus merely reflecting clinical variants of the same tumour; (B) SS and MF developing separately from a common precursor cell, with subsequent large cell transformation of the MF into LCL; (C) unrelated T cell clones as the origin of the different cutaneous T cell lymphoma phenotypes.

Figure 3 T cell receptor β polymerase chain reaction (6% polyacrylamide gel stained with ethidium bromide). Lane 1, positive control T cell line Hut; lane 2, skin in the Sézary syndrome (SS) phase; lane 3, blood in the SS phase; lane 4, blood after six cycles of extracorporeal photopheresis; lanes 5 and 6, skin in the mycosis fungoides (MF) phase; lane 7, lymph node in the large cell lymphoma (LCL) phase; lane 8, bone marrow in the LCL phase; lane 9, spleen in the LCL phase; lane 10, tonsil in the LCL phase; lane 11, negative control.
not yet been fully clarified. Patients with MF, who comprise 70% of all cases of CTCL, normally present with cutaneous patches and plaques and pursue a longer, indolent clinical course. However, in some patients there is a progression to erythrodermic disease, which may be associated with morphological changes in the peripheral blood. In contrast, SS is a leukaemic variant of CTCL, clinically characterised by early onset of erythroderma, pruritus, and peripheral lymphadenopathy. It is an aggressive clinical entity associated with poor prognosis and median survival of two to three years. In rare cases, MF can evolve into SS during progression of the disease. In addition, the incidence of large cell transformation in MF and less often in SS is a well known phenomenon, having been reported in 8% to 55% of cases. However, to our knowledge, the transition of SS into MF has not been reported yet.

There are three possible explanations of the unusual change in the clinical phenotype in our patient, namely: (1) the established SS progressed smoothly into MF and was then subsequently transformed into LCL; (2) SS and MF developed separately from a common precursor cell, with subsequent transformation of the MF into LCL; or (3) SS, MF, and LCL each arose separately from unrelated T cell clones (fig 2). To investigate the validity of these possibilities, we compared the rearrangement of the TCR genes in the three CTCL phenotypes, as related to the clinical course.

Clonal TCR rearrangements of the same size were detected in skin, blood, and lymph node at the initial diagnosis of SS, as shown by GeneScan analysis of the TCR β and γ PCR products. During clinical remission, no clonal T cell populations were detected in the peripheral blood. However, a T cell clone identical to that of SS could be identified in early MF lesions and during the LCL phase in lymph node, spleen, tonsil, and bone marrow (fig 3). Sequence analysis demonstrated that the clonal PCR products were not only of the same size but also had the same sequence (Vβ19D2Jβ2.7), thus demonstrating the identity of the clonal T cell populations involved.

The presence of the same T cell receptor rearrangements in the neoplastic cells from Sézary syndrome, mycosis fungoides, and large cell lymphoma found in our patient show that they were derived from a common T cell clone.

These results rule out the possibility that the clonal T cell populations of the three clinical entities originated from different or unrelated precursors. The presence of the same TCR rearrangements in the neoplastic cells from SS, MF, and LCL found in our patient show that they were derived from a common T cell clone. The clinical development of different lymphoma phenotypes harbouring the identical cell clone has already been described in B cell lymphoma (for example, Hodgkin disease in combination with follicular lymphoma), and lymphoproliferations, such as MF or anaplastic large cell lymphoma in combination with lymphomatoid papulosis.

The complete remission of the SS seen in our patient may have been the result of his good response to ECP. In addition to the low side effect profile of ECP, this treatment modality has shown to be effective and to induce complete remission in several patients with SS. Recent findings indicate that ECP induces a clone specific Th1-type immune response, with subsequent selective reduction of the malignant T cell population, as a result of T cell apoptosis and the modulation of dendritic cell differentiation. These findings were confirmed in our patient by the complete disappearance of circulating blood tumour cells and the complete regression of erythroderma after ECP treatment.

The underlying reasons for the relapse with a clinical picture of MF are unknown. However, the “pro Th-1 milieu” created by the ECP could have altered the activation pattern of the large number of surviving SS cells towards MF cells.

To our knowledge, this is the first report describing the transition of SS into MF. The neoplastic cells of the SS and the MF found in our patient were derived from a common T cell, which finally progressed to an LCL phenotype. Therefore, it seems reasonable to assume that SS, MF, and LCL represent a common genotype with different possible phenotypes of cutaneous lymphoma.

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