Interstitial cells of Cajal, enteric nerves, and glial cells in colonic diverticular disease

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Background: Colonic diverticular disease (diverticulosis) is a common disorder in Western countries. Although its pathogenesis is probably multifactorial, motor abnormalities of the large bowel are thought to play an important role. However, little is known about the basic mechanism that may underlie abnormal colon motility in diverticulosis.

Aims: To investigate the interstitial cells of Cajal (the gut pacemaker cells), together with myenteric and submucosal ganglion and glial cells, in patients with diverticulosis.

Patients: Full thickness colonic samples were obtained from 39 patients undergoing surgery for diverticulosis. Specimens from tumour free areas of the colon in 10 age matched subjects undergoing surgery for colorectal cancer served as controls.

Methods: Interstitial cells of Cajal were assessed using anti-Kit antibodies; submucosal and myenteric plexus neurones and glial cells were assessed by means of anti-PGP 9.5 and anti-S-100 monoclonal antibodies, respectively.

Results: Patients with diverticulosis had normal numbers of myenteric and submucosal plexus neurones compared with controls (p = 0.103 and p = 0.516, respectively). All subtypes of interstitial cells of Cajal were significantly (p = 0.0003) reduced compared with controls, as were glial cells (p = 0.0041).

Conclusions: Interstitial cells of Cajal and glial cells are decreased in colonic diverticular disease, whereas enteric neurones appear to be normally represented. This finding might explain some of the large bowel motor abnormalities reported to occur in this condition.

METHODS AND MATERIALS

Patients
Colon specimens were obtained from 39 patients (17 men, 22 women; age range, 49–78 years) undergoing elective left hemicolectomy for diverticular disease (35 cases) or emergency surgery for acute diverticulitis with pericolic abscess (four cases) in the period January 1999 to January 2004. None of the patients had concomitant tumours, bowel obstruction, or other diseases of the colon. The studies were carried out in accordance with local ethical guidelines, following the recommendations of the Declaration of Helsinki (Edinburgh revision, 2000).

Abbreviations: ICC, interstitial cells of Cajal; IC-IM, muscle fibre interstitial cells of Cajal; IC-MY, myenteric region interstitial cells of Cajal; IC-SM, submucosal interstitial cells of Cajal; PGP 9.5, protein gene product 9.5
Controls
We used 10 specimens from age and sex matched subjects undergoing left hemicolectomy for non-obstructing colorectal cancer as controls. The control specimens were taken at least 5 cm from the resection margin in tumour free areas.

Methods
After removal, the surgical specimens were immediately fixed in 10% neutral buffered formalin for 24 hours, then 12 to 20 full thickness samples were taken from portions bearing diverticula and from those that were macroscopically normal. For conventional histology, 5 µm thick paraffin wax embedded sections were stained with haematoxylin and eosin, periodic acid Schiff, and trichrome stain.

Immunohistochemistry
At least 10 samples (five from diverticular and five from macroscopically normal portions) for each patient were processed for immunohistochemistry. To evaluate the enteric nervous system,21 we investigated PGP 9.5 (protein gene product 9.5), a cytoplasmic protein that acts as a marker of general neural tissue,22 23 and the glial marker protein S-100.24 Ganglion cells were assessed with a monoclonal anti-PGP 9.5 antibody (IgG2a; 1/100 dilution; Biomeda, Foster City, California, USA); Schwann cells, intragangliar glial cells close to ICC, and myenteric ganglia were assessed with a specific monoclonal antibody (anti-S-100; 1/50 dilution; Dako, Carpenteria, California, USA). S100 immunostaining highlights ganglion cells as prominent negatively stained cells surrounded by positive Schwann/glial cells.

The expression of Kit (CD117) was assessed using a rabbit polyclonal antibody (IgG; dilution 1/50; Dako) specific for ICC. In brief, consecutive formalin fixed, paraffin wax embedded sections were de waxed and rehydrated through a decreasing alcohol series up to distilled water. Sections were then subjected to heat induced epitope retrieval by immersion in a heat resistant container filled with citrate buffer solution (pH 6.0), placed in a pressure cooker, and microwaved for 20 minutes. Endogenous peroxidase activity was blocked by incubation with a 3% solution of H2O2 for five minutes. Kit immunostaining was carried out using a peroxidase based visualisation kit (Dako EnVision™), according to the manufacturer’s recommendations. PGP 9.5 and S-100 immunostaining was carried out by means of a peroxidase based visualisation kit (Dako LSAB™), according to the manufacturer’s recommendations. Diaminobenzidine tetrahydrochloride was used as the chromogen. The slides were then counterstained with Mayer’s haematoxylin for five seconds, dehydrated, and mounted in Clarion (Biomeda). To account for non-specific staining, peptides that blocked polyclonal antibody binding (passage with normal goat serum) were used, or sections were incubated in the absence of primary antibody. In these cases, no immunostaining was detected.

Kit positive mast cells served as an internal positive control. Not only nucleated cells but also Kit positive labelled elongated structures in the diverticular sacs of patients were compared with tumour free areas in the controls.

For PGP 9.5 and S100 staining, both the submucosal and the myenteric plexuses were assessed using optical microscopy at ×20 magnification (Olympus BX 40, Tokyo, Japan). For each patient, the number of immunopositive cells was calculated and expressed as the mean number of cells in 10 well stained and well oriented high power fields. The longitudinal muscle layer, intermuscular layer, and circular muscle layer were each assessed in 10 different fields.

Figure 1 Expression of PGP 9.5 in the submucosal and myenteric plexus of controls (A and C, respectively) and patients (B and D, respectively). Original magnification, ×20 (A, B) and ×40 (C, D).
Statistical analysis
The Kolmogorov-Smirnov test for normality was applied and showed that the data were normally distributed. Data from controls and patients were thus compared by means of the Student’s t test for unpaired data. Values of $p < 0.05$ were chosen for rejection of the null hypothesis.

RESULTS
Conventional histology
In both groups, the mucosa, submucosa, smooth muscle, and nerve plexus architecture appeared normal on haematoxylin and eosin, trichrome, and periodic acid Schiff staining. In the four cases in which diverticulitis was acute, an inflammatory infiltrate cytologically composed of neutrophils, lymphocytes, macrophages, and occasional giant plurinucleated cells was seen, localised in and around the boundaries of the affected diverticular sacs. Apart from these cases, and the presence of occasional neutrophils, plasma cells, and lymphocytes mainly within the luminal aspects of the lamina propria in both groups, no acute inflammatory cells (or intranuclear or viral inclusions) were seen in or around muscular or nervous structures (particularly at the myenteric plexus level).

Immunohistochemistry
The expression of PGP 9.5 was not significantly different between patients with diverticulosis and controls, either in the myenteric plexus (mean, 50 (SD, 17) v 60 (17) cells; $p = 0.103$) or in the submucosal plexus (mean, 41.2 (SD, 15) v 44.5 (11) cells; $p = 0.516$; fig 1). The number of S-100 positive cells was significantly lower in patients in both the myenteric plexus (mean, 171 (SD, 40) v 214.4 (33.3) cells; $p = 0.0041$) and the submucosal plexus (mean, 78.3 (SD, 27.4) v 127 (47) cells, $p = 0.0004$; fig 2). There was a significant decrease at each anatomical region in patients with diverticulosis for all three subclasses of ICC evaluated (IC-SM: mean, 7.6 (4.4) v 29 (8) cells, $p = 0.0001$; IC-MY: mean, 131.3 (SD, 43) v 214.2 (84) cells, $p = 0.0003$; IC-IM: mean, 10 (7) v 37 (12) cells, $p = 0.0001$; fig 3). No differences with regard to S-100 and Kit positive cells were found between the four patients with acute diverticulitis and the other patients with diverticulosis, or between diverticulosis and non-diverticular areas.

Scattered Kit positive cells (identified as mast cells and used as internal controls) were seen in the patients’ mucosa, and had a similar distribution to that seen in the control tissue. No alterations in size or shape were seen in the three ICC subpopulations.

DISCUSSION
The main finding of our study was that patients with colonic diverticulosis have significantly reduced numbers of colonic ICC and enteric glial cells compared with controls. To our knowledge, this is the first study to have revealed such abnormalities. We focused our attention on ICC because an increasing amount of recent evidence indicates that this cell population plays a pivotal role in the regulation of intestinal motor function. This is supported by the fact that, in addition to functioning as pacemaker cells that generate slow waves, ICC mediate neurotransmission from enteric motor neurones. In fact, these cells are an integral part of the enteric motor system, and may be the primary site of innervation; neural regulation of the musculature may also occur via the ICC.

“The reduction of interstitial cell of Cajal function might be responsible for the significant decrease in rhythmic colonic contractile patterns that we recently described in patients with diverticulosis”

The role of ICC as intestinal pacemakers has been clearly established in experimental animal models, which have shown that a lack of ICC networks leads to the absence of slow waves and is accompanied by delayed or absent intestinal motility. In the upper gastrointestinal tract a lack of ICC has been found in diseases associated with gastric and small bowel motility (diabetic gastroparesis, chronic intestinal pseudo-obstruction, etc). Data on human colonic ICC are still scarce, especially in pathological conditions, and chiefly limited to congenital diseases and slow transit constipation. We decided to study diverticular disease because it is a disorder with frequent and measurable alterations of colonic motility. We found that these patients consistently had a significant reduction of all subpopulations of ICC and of enteric glial cells, whereas the enteric neuronal population appeared to be normal. These alterations might explain the colonic motor abnormalities documented in patients with diverticulosis (increased overall motility, abnormal response to eating, retropropagation of mass movements, etc). We are presently unable to explain why decreased numbers of colonic ICC and S-100 positive cells and structures should be associated with motor abnormalities. It is possible that a reduction or loss of ICC function decreases or eliminates colonic electrical slow wave activity, thereby decreasing the contractile response and resulting in delayed transit. For example, the reduction of ICC function might be responsible for the significant decrease in rhythmic colonic contractile patterns that we recently described in patients with diverticulosis; in an experimental animal model these patterns appear to be driven by ICC.

Moreover, because enteric glial cells are thought to function as intermediaries in enteric neurotransmission, their decrease might further weaken the already precarious neuroenteric balance described thus far.

Although ICC drive spontaneous rhythmic motility in the gut, the enteric nervous system may also play a role. This is particularly true in diverticulosis, where loss of smooth muscle choline acetyltransferase activity, upregulation of M3 receptors, and increased in vitro sensitivity of the smooth muscle to exogenous acetylcholine have been documented, suggesting that cholinergic denervation hypersensitivity may occur in this condition. The association of such alterations with a decrease in ICC might contribute to the motor abnormalities described in diverticulosis.

The evidence is clearly insufficient to determine whether loss of ICC in diverticulosis is a primary event or is secondary to another lesion or to the rearrangement that occurs when...
changes take place in the colonic wall. A mechanical factor (for instance the push leading to diverticula formation with compression/atrophy of surrounding structures, including ICC and nerve structures) cannot be ruled out.

Similar considerations on whether these changes are primary or secondary to (still) unknown factors may be applied to other pathological conditions of the colon in which a decrease of ICC has been demonstrated, such as slow transit constipation\textsuperscript{15 16} and intestinal pseudo-obstruction. In fact, in about one third of cases of intestinal pseudo-obstruction limited to the colon a consistent reduction of ICC, similar to that found here in patients with diverticulosis, has recently been described.\textsuperscript{39}

However, increasing evidence from experimental animal models and reports in human subjects suggest that the ICC may play an important role in the pathophysiology of gastrointestinal motor abnormalities; these data shed new light on the mechanisms underlying diverticulosis and provide new directions for further studies on motor abnormalities in the gastrointestinal tract, with possible specific targeting of future therapeutic approaches.\textsuperscript{40}

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