Ultrastructural features of eosinophilic oesophagitis: impact of treatment on desmosomes

Kelley E Capocelli,1 Shahan D Fernando,2,3,4,5 Calies Menard-Katcher,2,3,4,5 Glenn T Furuta,2,3,4,5 Joanne C Masterson,2,3,4,5 Eric P Wartchow1

ABSTRACT

Aims A growing body of evidence suggests a role for altered epithelial barrier function in the pathophysiology of eosinophilic oesophagitis (EoE), but few have described the epithelial structure during inflammation. The purpose of this study was to define ultrastructural features of active, inactive EoE and control subject’s oesophageal epithelia.

Methods We prospectively enrolled patients undergoing diagnostic upper endoscopy for evaluation of EoE. Mucosal pinch biopsies were obtained from the distal oesophagus and processed for routine histology and electron microscopic assessment. Clinical features of enrolled subjects were analysed and subjects were divided into four groups: normal, gastroesophageal reflux disease (GERD), inactive EoE and active EoE. Representative photomicrographs of the basal and superficial epithelia were reviewed for abnormalities. Desmosomes were quantified on the surface of epithelia three to four prickle-cell layers above the basal layer.

Results Twenty-nine paediatric cases (ages 2–18 years) were enrolled in the study. We observed a significant decrease in the number of desmosomes per cell (DPC) of subjects with active EoE compared with inactive EoE, GERD and normal epithelia. With respect to DPC, no significant differences were found between inactive EoE compared with GERD or normal subjects. Additional ultrastructural features observed included epithelial microplicae and evidence of eosinophil transmigration, degranulation, and sombrero formation.

Conclusions Consistent with clinical and molecular findings, our ultrastructural data provide support for an altered oesophageal barrier in paediatric cases with active EoE, which may improve following treatment.

INTRODUCTION

Immunohistochemical and electron microscopic studies of oesophageal epithelium affected by eosinophilic oesophagitis (EoE) have focused on findings related to eosinophil activation and have identified a number of important characteristics including eosinophil degranulation, inverted granule protein cores and sombrero vesicle formations.1–4 While these findings document the impact of activated eosinophils as a functional endpoint, a growing body of evidence supports a role for altered epithelial barrier function in the pathophysiology of eosinophilic oesophagitis (EoE), but few have described the epithelial structure during inflammation. The purpose of this study was to define ultrastructural features of active, inactive EoE and control subject’s oesophageal epithelia.

Methods

We prospectively enrolled paediatric subjects between 1 year and 18 years of age undergoing upper endoscopy, to assess for EoE, from January 2011 to December 2012. Consecutive patients were approached to enrol in the study if they were undergoing initial endoscopic evaluation for EoE or repeat evaluation following treatment of EoE (topical steroids or diet). Clinical features of each subject were recorded and all underwent upper endoscopy with biopsy. Subject groups were defined as follows:

- Active EoE was defined as subjects with symptoms of oesophageal dysfunction and ≥15 eosinophils per high-power field (HPF) in whom alternative aetiologies of eosinophilia had been ruled out.
- Inactive EoE was defined as subjects with an established diagnosis of EoE who had undergone treatment with topical steroids or dietary exclusions, and whose symptoms resolved, and eosinophilia were found to be <15 eosinophils/HPF.
- Patients with gastroesophageal reflux disease (GERD) had symptoms related to reflux and <15 eosinophils/HPF.
- Normal persons had symptoms, such as abdominal pain, diarrhoea and vomiting significant enough to warrant an endoscopy, whose oesophageal histology was normal.

Tissue samples from the distal oesophagus were obtained and immediately fixed in cacodylate-buffered 2.5% glutaraldehyde and processed as previously described with samples being fixed in 2.0% osmium tetroxide then embedded in epoxy resin.5 Ultrathin sections were poststained with uranyl acetate and lead citrate and imaged using a Hitachi H-7650 TEM (Hitachi, Tokyo, Japan) equipped with an AMT XR-11 digital camera system (Advanced Microscopy Techniques, Massachusetts, USA). Samples were assessed without knowledge of their clinical diagnosis by two of the investigators (EPW and SDF).
As described previously, we enumerated desmosomes within the epithelium by examining one to nine individual epithelial cells per patient that were three to four cell layers above the basal layer. This region of interest was selected to maintain consistency in analysis. Desmosomes were identified and quantified as discrete, linear hyper-densities along the outer cellular membrane that make physical contact with an adjacent cellular membrane. Up to 43 cells were analysed within each of the four patient groups. Additional morphological features of the epithelia were identified independently by five investigators (EPW, KEC, JCM, SDF, GTF) who then reviewed their findings together.

Statistical significance was determined using a Kruskal–Wallis test and Dunn’s test for multiple comparisons. Statistical analysis was performed using GraphPad Prism V.6.0d (GraphPad Software, La Jolla, California, USA).

RESULTS
Twenty-nine persons were enrolled in the study and clinical features are shown in table 1.

Typical endoscopic features of EoE, including linear furrows and exudates, as well as histological findings such as inflammation and basal cell hyperplasia, are seen in figure 1. To minimise technical variability that may exist with assessing the stratified squamous epithelia in electron micrographs, we analysed the basal layer of the epithelia (figure 2A). The desmosome number was significantly reduced in epithelia with active EoE compared with normal epithelium (7.6±3.7 vs 14.1±5.9, active EoE vs normal, p=0.0001; figure 2B). Following standard EoE treatment, the desmosome number in inactive EoE (13.2±5.4) was virtually the same as normal epithelia. Additionally, desmosome numbers in epithelia of patients with GERD (12.0±4.0) were similar to inactive EoE and normal epithelia.

Further assessment of the intercellular space (ICS) revealed that the width of the ICS appears to be narrower in normal epithelial cancers, but ours is the first to identify epithelial ultrastructural findings in a prospective fashion in paediatric cases with active and inactive EoE.9 16–21 Our findings bear particular relevance and support for recent studies that identify a role for epithelial barrier dysfunction in EoE. Through the use of electrical impedance, transepithelial resistance and fluorescent flux, van Rhijn et al22 reported diminished epithelial barriers in adults with EoE compared with those with normal epithelium. Katzka et al23 demonstrated increased dilated ICS associated with decreased expression of filaggrin, zonula occludens-1 and claudin-1 in oesophageal biopsies from patients with untreated EoE. Sherrill et al24 examined molecular regulation of barrier dysfunction and found that desmoglein-1 expression was decreased via a type-2 cytokine microenvironment, and was associated with diminished barrier function in an ex vivo squamous epithelial model. In an electron micrograph, the number of desmosomes appears to be diminished, but enumeration was not presented. Here, we provide quantitative, morphological evidence that the oesophageal epithelial ultrastructure is indeed altered in EoE, a finding consistent with these functional and molecular studies. Additionally, we show that desmosome number returns to that found in the normal epithelium following treatment. This observation suggests that morphological evidence of barrier dysfunction occurs secondary to inflammation and not as a primary defect. Alternatively, treatments may repair underlying inherent epithelial barrier defects.

Our results also identify a number of interesting and important features related to relationships between the epithelium and eosinophils. First, while our findings identify the morphological

<table>
<thead>
<tr>
<th>N</th>
<th>Age (years)</th>
<th>Symptoms</th>
<th>Endoscopic findings†</th>
<th>Eos/HPF</th>
</tr>
</thead>
<tbody>
<tr>
<td>Active</td>
<td>9</td>
<td>2–15</td>
<td>Vomiting-5, Abdominal pain-4</td>
<td>F-6, E-4, LOVP-4</td>
</tr>
<tr>
<td>Inactive</td>
<td>9</td>
<td>4–7</td>
<td>None</td>
<td>F-1, E-1, LOVP-1, N-6</td>
</tr>
<tr>
<td>GERD</td>
<td>4</td>
<td>14–18</td>
<td>Abdominal pain-2, Dysphagia-1, Regurgitation-1</td>
<td>N-4</td>
</tr>
<tr>
<td>Normal</td>
<td>7</td>
<td>9–17</td>
<td>Abdominal pain-5, Diarrhoea-1, Vomiting-1</td>
<td>F-1, LOVP-1, FR-1, N-5</td>
</tr>
</tbody>
</table>

†Selected based on previous studies of epithelial features in EoE.22

Table 1 Clinical features of patients
disruption of the normal stratified epithelia, our assessment also reveals the inherent nature of the oesophageal epithelium to maintain an intact barrier. We show that the superficial epithelia associated with active and inactive EoE, similar to that found in GERD and normal subjects, maintain a robust pattern of interdigitating microplicae or intercellular ridges. This ‘zipper-like’ morphology emphasises the importance of maintaining a barrier, which appears different than the basal layer, and further illustrates the pleotropic nature of the oesophageal epithelium. Therefore, the superficial surface possesses an infrastructure comprised of flattened epithelial cells, microplicae, tight junctions and narrow ICS that creates a network to prevent luminal contents from penetrating the underlying immunomicromilieu. In EoE, the epithelia may limit exposure of the immune system to food allergens that initiate and perpetuate eosinophilic inflammation. Additionally, we provide evidence of the dynamic nature of eosinophil chemotaxis within the epithelia as seen in the figures demonstrating eosinophil transmigration. Despite a

Figure 1  Endoscopic and histological assessment of the oesophagus between normal patients and those with active eosinophilic oesophagitis (EoE), inactive EoE and GERD. H&E stained images obtained at ×10 magnification. Scale bars represent 100 μ.

Figure 2  Desmosome quantification. (A) Illustration of the oesophageal epithelium depicting the basal epithelial layer near the bottom and squamous epithelial layer near the top. The black box highlights the third prickle cell layer used to quantify desmosomes. (B) Number of desmosomes per cell in normal patients compared with those with GERD, active eosinophilic oesophagitis (EoE) and inactive EoE. Results are expressed as mean±SD.
number of studies defining the chemotactic pathways associated with EoE, none have captured an eosinophil actively migrating between the stratified squamous epithelia in EoE.\textsuperscript{23–25} Lastly, our ultrastructural analysis confirms recent observations that report the presence of eosinophil degranulation, sombrero vesicle formation and reversal of core proteins in EoE.\textsuperscript{1}

While this ultrastructural report provides a high level of detail, it is difficult to provide a more expansive quantitative assessment or deeper mechanistic understanding than can be obtained through routine histological or molecular analysis, respectively.\textsuperscript{52 62 7} For instance, since eosinophils are unevenly distributed within the epithelia, quantification of their features and patterns of distribution in such a small field of view may not truly represent the overall mucosal inflammatory state. Therefore, we were limited to observational assessments only. Similarly, we did not report exact measurements of the width of the ICS due to the high degree of variability in cell membrane shapes and forms.

Although enumeration of desmosomes within superficial squamous epithelia has been used in a number of previous studies to assess epithelial integrity and morphology,\textsuperscript{9 11 19 20 28} our enumeration focused on the deeper, basal epithelial cells primarily due to technical concerns about the integrity of the superficial mucosa. Because of mechanical stresses on the epithelial surface caused by the pinch biopsy procedure, we could not consistently assess the uppermost superficial surfaces of the epithelia. However, we felt confident that the basal layer could be identified by allowing the underlying lamina propria to serve as a landmark. Thus, the numbers of desmosomes may appear to be sparse in the basal layer (figure 3) when compared with

\begin{itemize}
\item \textbf{Figure 3} Electron micrographs of basal epithelial cells from normal patients (A–C) and patients with active eosinophilic oesophagitis (B and D). A and B illustrate relative intercellular spaces. Arrowheads indicate the basal lamina separating basal cells from the lamina propria. (C and D) depict desmosomes (arrow) encircling basal cells located within the third prickle cell layer. Scale bars represent 10 μ (A and B) and 2 μ (C and D).
\end{itemize}

\begin{itemize}
\item \textbf{Figure 4} Electron micrographs of the stratified squamous epithelia in normal patients (A) and patients with active eosinophilic oesophagitis (B). A resting eosinophil (arrow) is depicted near the luminal surface. Scale bars represent 2 μ.
\end{itemize}
those in other more superficial parts of the epithelia (figure 4). These findings may indicate that the more superficial epithelia require a larger number of desmosomes than the more basal epithelia to maintain barrier function for reasons discussed above, but this speculation would require further studies. Notwithstanding the technical challenges, we feel that the present study using transmission electron microscopy provides unique ultrastructural insights into features of EoE that would not be permitted through either routine histological or molecular techniques.

In summary, we report that the basal layer of stratified squamous epithelia appears to be altered in paediatric subjects with EoE. Future studies using laser capture microscopy to determine the exact molecular patterns leading to this finding will provide more mechanistic insights and potential identification of novel therapeutic targets.

**Take home messages**

- Oesophageal epithelial barrier dysfunction may play a critical role in the initiation and/or perpetuation of eosinophilic oesophagitis (EoE), a chronic immune-mediated inflammatory condition.
- At the ultrastructural level, a significant decrease in the number of desmosomes on the surface of the oesophageal epithelia of paediatric subjects with active EoE is observed. This provides support for recent works identifying evidence of altered barrier function in EoE.
- Eosinophils actively transmigrate through the oesophageal epithelium where they intimately adjoin with squamous epithelial cells prior to degranulation.

**Figure 5** Electron micrographs of oesophageal epithelial eosinophils in various stages of activity including (A) resting state, (B) epithelial transmigration, (C) partial degranulation and (D) complete degranulation. Inverted granule protein cores (arrow) and sombrero formation (arrowhead) are seen within the degranulating eosinophils. Scale bars represent 4 μm.

**Correction notice** A sentence in Abstract section has been corrected since published Online First.

**Acknowledgements** The authors acknowledge and thank the physicians (Robert Kramer, Edward Hoffenberg, Edwin Liu, Edwin de Zoeten, Shikha Sundaram, Cara Mack, Michael Narkewicz, Ronald Sokol, Jason Soden, Deborah Neigut, David Brumbaugh and Christine Waasdorp Hurtado), nurses (Amy Kopp and Jo Anne Newton), research coordinators (Wendy Moore and Dresden Whitehead), technical staff (Bill Marcovich and Luann Goin), biostatistician (Zhaoxing Pan) and EM Laboratory Director (Gary Miera) who contributed to this work. We are grateful to our patients and families who consented to be a part of this study.

**Contributors**

- Study concept and design (KEC, SDF, GTF, JCM, EPW); acquisition of data (KEC, SDF, CM-K, GTF, EPW); analysis and interpretation of data (KEC, SDF, GTF, JCM, EPW); drafting of manuscript (SDF, GTF); critical revision of manuscript for important intellectual content (KEC, SDF, CM-K, GTF, JCM, EPW); statistical analysis (SDF, JCM); obtained funding (GTF, JCM); study supervision (GTF, JCM, EPW).

**Funding** This study was supported by grants from the US National Institutes of Health K24-DK100303 (GTF) and T32-DK067009-09 (SDF), and American Partnership for Eosinophilic Disorders (JCM).

**Competing interests** None.

**Ethics approval** This study was approved by the Colorado Multi-Institutional Review Board.

**Provenance and peer review** Not commissioned; externally peer reviewed.
REFERENCES