hMSH6: a potential diagnostic marker for oral carcinoma in situ

Maryam Jessri,1 Andrew J Dalley,1 Camile S Farah1,2

ABSTRACT

Oral medicine specialists rely upon accurate assessment of pathology to rationalise lesion management, especially for high-risk oral epithelial dysplasia, carcinoma in situ (CIS) and oral squamous cell carcinoma. Cross-discipline cancer research has highlighted the role of genetic instability in neoplasia. Improved diagnostic stringency from translation of immunostaining for DNA repair defects into current pathology practice has potential to benefit pathologists, clinicians and patients. The focus of this study was the obligatory and non-obligatory components of the MutLα and MutSα mismatch repair heterodimers, namely hMLH1, hMSH2, hPMS2 and hMSH6, which were studied in 274 formalin-fixed paraffin-embedded sections. A readily apparent inverse correlation between oral disease severity and both obligatory and non-obligatory components of MutLα and MutSα was observed (hMLH1, p=−0.715; hPMS2, p=−0.692; hMSH2, p=−0.728; and hMSH6, p=−0.702), with particularly conspicuous loss of hMSH6 expression from the stratum basale of CIS.

INTRODUCTION

Mismatch repair (MMR) pathways encompass the strand-specific, postreplicative DNA repair mechanisms that are key to maintenance of genomic integrity and stability. Numerous MMR proteins intervene to repair DNA biosynthetic errors, single-base substitution mismatches and insertions/deletions in microsatellites.1 The most abundant MMR heterodimer, MutSα, initiates the repair mechanism and is composed of hMSH2 and hMSH6.2 Subsequently, the MutLα heterodimer is instrumental in excising and correcting mismatched nucleotides and is composed of hMLH1 and hPMS2.3

Oral squamous cell carcinoma (OSCC) accounts for 90% of head and neck cancer which globally represents the sixth highest rate of cancer mortality.4 OSCC can be attributed to both environmental and genetic factors. The presence and severity of oral epithelial dysplasia (OED) in oral potentially malignant lesions (OPML) is the most important indication of increased neoplastic transformation risk.5 Impaired expression of MMR genes has been demonstrated for OSCC;6 7 a process that can involve hypermethylation of their promoter regions.8 9 Genetic modification of the MMR pathway has also been investigated for OPML, although to a lesser extent.10 11

We hypothesised correlation between MMR protein immunoexpression and the pathology grading of OED in OPML and differentiation in OSCC. Accordingly, we compared MutSα and MutLα expression in pathologist-graded biopsies of OSCC (well-differentiated or poorly differentiated), OPML (low-risk (LD) or high-risk dysplasia (HD) including carcinoma in situ (CIS)) compared with normal oral mucosa.

MATERIALS AND METHODS

Patient samples

Two hundred and seventy-four archival formalin-fixed paraffin-embedded specimens: 113 OSCC, 34 HD including CIS, 37 LD and 90 normal oral mucosa were retrieved and rediagnosed according to WHO criteria by oral pathologist (CSF) (table 1).12

Immunohistochemistry

Five micrometre sections were incubated overnight at 4°C with mouse monoclonal antihuman antibodies (Biocare Medical, Concord, California, USA): hMLH1 (G168-15), hPMS2 (A 16-4), hMSH2 (FE11) and hMSH6 (BC/44). Secondary antibody and revealing agent were obtained from Biocare Medical (MACH1 Universal HRP polymer kit, M1U539L10, Biocare Medical, Concord California, USA). Positive controls were from colon, and routine negative staining was performed.

Scoring

All epithelial layers from 16 randomised microscopic fields (×400) were scored. Stain intensity was scaled: 0=no stain (equivalent to negative control) to 3=brown nuclear staining (equivalent to positive control). Percentage of positive cells was calculated as (numerator=product of antigen-positive cell count and intensity, denominator=total cell count).

All fields were evaluated if sample size precluded 16 fields.

Statistical analysis

Analyses were conducted with IBM SPSS Statistics V20 software (IBM Corporation, Armonk, New York, USA). MMR protein to lesion severity relationship was tested by Spearman’s rank correlation after ordinal grading of lesions: normal, 0; LD, 1; HD, 2; well-differentiated OSCC, 3 and poorly differentiated OSCC, 4. Age, sex and lesion site were control variables in the multinomial logistic regression analysis of each MMR protein. Backward stepwise elimination of variables from the saturated multinomial model was performed to achieve a best fit.

RESULTS

Representative photomicrographs for MMR antigen expression are presented (figure 1). In general, MMR protein expression was highest in normal samples and showed a decreasing trend as lesion severity increased (figure 2). Disease...
Table 1  Patient demographic and clinical information

<table>
<thead>
<tr>
<th>Variable</th>
<th>Normal mucosa</th>
<th>Dysplasia</th>
<th>OSCC</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Low risk</td>
<td>High risk</td>
<td>Carcinoma in situ</td>
<td>Well differentiated</td>
</tr>
<tr>
<td>Gender</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>52</td>
<td>13</td>
<td>9</td>
<td>7</td>
<td>55</td>
</tr>
<tr>
<td>Female</td>
<td>38</td>
<td>24</td>
<td>12</td>
<td>6</td>
<td>31</td>
</tr>
<tr>
<td>Age at diagnosis</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;45 years</td>
<td>57</td>
<td>28</td>
<td>16</td>
<td>11</td>
<td>20</td>
</tr>
<tr>
<td>≥45 years</td>
<td>33</td>
<td>9</td>
<td>5</td>
<td>2</td>
<td>66</td>
</tr>
<tr>
<td>Biopsy site</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tongue</td>
<td>9</td>
<td>3</td>
<td>2</td>
<td>1</td>
<td>14</td>
</tr>
<tr>
<td>Palate</td>
<td>9</td>
<td>4</td>
<td>1</td>
<td>0</td>
<td>6</td>
</tr>
<tr>
<td>Gingiva</td>
<td>21</td>
<td>9</td>
<td>4</td>
<td>2</td>
<td>15</td>
</tr>
<tr>
<td>Labial mucosa</td>
<td>12</td>
<td>2</td>
<td>1</td>
<td>0</td>
<td>10</td>
</tr>
<tr>
<td>Buccal mucosa</td>
<td>36</td>
<td>14</td>
<td>9</td>
<td>8</td>
<td>29</td>
</tr>
<tr>
<td>Floor of mouth</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>9</td>
</tr>
<tr>
<td>Not available</td>
<td>1</td>
<td>3</td>
<td>2</td>
<td>0</td>
<td>3</td>
</tr>
<tr>
<td>Smoking history</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Positive</td>
<td>9</td>
<td>4</td>
<td>6</td>
<td>4</td>
<td>3</td>
</tr>
<tr>
<td>Negative</td>
<td>14</td>
<td>4</td>
<td>0</td>
<td>2</td>
<td>7</td>
</tr>
<tr>
<td>Unavailable</td>
<td>67</td>
<td>29</td>
<td>15</td>
<td>7</td>
<td>76</td>
</tr>
<tr>
<td>Total</td>
<td>90</td>
<td>37</td>
<td>21</td>
<td>13</td>
<td>86</td>
</tr>
</tbody>
</table>

OSCC, oral squamous cell carcinoma.

Figure 1  Representative photomicrographs showing (A) normal oral mucosa, (B) low-risk oral epithelial dysplasia, (C) high-risk oral epithelial dysplasia, (D) well-differentiated oral squamous cell carcinoma (OSCC) cell nests and the underlying connective tissue and (E) poorly differentiated OSCC stained with (i) H&E, (ii) hMLH1, (iii) hPMS2, (iv) hMSH2 and (v) hMSH6. Magnification ×20.
severity correlated significantly with expression of: hMLH1 ($\rho = -0.715$), hPMS2 ($\rho = -0.692$), hMSH2 ($\rho = -0.728$) and hMSH6 ($\rho = -0.702$); (n=274). The majority (nine out of 13) of CIS samples exhibited localised loss of hMSH6 from the stratum basale (figure 3). The complementary obligatory and non-obligatory components of the MutS$\alpha$ and MutL$\alpha$ dimers correlated appropriately (hMSH2 and hMSH6, $\rho = 0.774$ and hMLH1 and hPMS2, $\rho = 0.847$).

When analysed individually by multinomial logistic regression, hMLH1, hPMS2 and hMSH2 were significantly associated with disease severity. Expression of hMSH6 was decreased in LD and OSCC but not for HD. In these analyses, MMR heterodimers were significantly decreased in all pathological conditions compared with normal, with an exception of hMSH6 in predicting the risk of HD (table 2). Table 3 presents the final results from backward stepwise elimination of lesion site, hMSH6 and sex from the saturated model. Each per cent increase in expression of hMLH1, decreased the odds of having LD (OR=0.92, CI 0.88 to 0.97) and well-differentiated OSCCs (OR=0.92, CI 0.88 to 0.96) by a factor of 0.92. Similarly, the odds of having poorly differentiated OSCCs were decreased by a factor of 0.86 (95% CI 0.81 to 0.91) with every 1% increase in expression of hMSH2. One intriguing finding which warrants further investigation was the localised loss of hMSH6 from the stratum basale of CIS specimens.

**DISCUSSION**

This study established correlation between loss of MMR protein expression with the severity of OED in OPML and with the degree of differentiation in OSCC. Three important findings emerged. First, a reduction in immunoexpression of the obligatory components of MutS$\alpha$ and MutL$\alpha$ during the progression of pathology from OPML to OSCC was observed. Second, a similar trend was found for expression of the non-obligatory components of MutS$\alpha$ and MutL$\alpha$. Third, there is conspicuous loss of hMSH6 expression from the stratum basale of CIS. These findings are consistent with the breadth of research surrounding MMR protein and specifically address the need for more rigorous techniques to assist with the grading of oral lesions, particularly CIS.
Expression of hMSH2 and hMLH1 have been investigated previously for OPML and OSCC, however, this study is the first to report simultaneous reduction in expression of both the obligatory and non-obligatory components. These results provide considerable weight to the established role of MMR process impairment during OSCC initiation and progression.

While the obligatory and non-obligatory components of MutSα and MutLα are complementary, they are, nonetheless, discrete gene products that are subject to independent regulation. Here, strong correlations between these obligatory and non-obligatory components have been reported (hMSH2 vs hMSH6, \( \rho = 0.774 \); hMLH1 vs hPMS2, \( \rho = 0.847 \)). These findings are indicative of a regulatory event that lies upstream of the individual gene expression levels, as opposed to discrete gene mutations or transcriptional modifications. Interestingly, based upon their predictive OR, only three out of the four MMR proteins were retained by the multinomial logistic model; the excluded MMR protein being hMSH6, the non-obligatory MutSα dimer. The low weight of hMSH6 in our statistical analyses can be readily accounted for by its anomalous expression in the majority of CIS cases, whereby hMSH6 expression was lost from the stratum basale of otherwise hMSH6-positive CIS epithelia (figure 3). This novel finding warrants further investigation, since localised loss of hMSH6 has potential to become a diagnostic marker in OED/OPML. Once validated, hMSH6 immunostaining may partially address the current need for additional diagnostic markers to assist with OED grading.

In this study, immunoexpression of MMR proteins in oral lesions was studied individually and as a panel. A stepwise logistic regression was used to (1) study these proteins as a panel and (2) eliminate those without a significant contribution to the model. Our results are suggestive of a diagnostic role for immunoexpression of hMLH1, hPMS2 and hMSH6 in grading of oral lesions individually, and preferably combined as a panel. Immunoexpression of hMSH6 was shown to particularly distinguish CIS samples. Our findings are independent of the statistical model used to prove their significance, and once validated would be of direct relevance to clinical pathologists as applicable biomarkers of oral disease severity to lessen possible diagnostic ambiguity among the more severe grades of OED, CIS and early invasive OSCC.

\[
\begin{array}{c|c|c|c|c}
\hline
\text{Parameter} & \text{OR} & \text{95% CI} \\
\hline
\text{Dysplasia} & & & \\
\hline
\text{hMLH1} & 0.88^* (0.84 to 0.91) & 0.95^{**} (0.92 to 0.99) & 0.84^* (0.81 to 0.88) & 0.74^* (0.74 to 0.83) \\
\text{hPMS2} & 0.89^* (0.86 to 0.92) & 0.96^{**} (0.92 to 0.99) & 0.87^* (0.84 to 0.90) & 0.82^* (0.78 to 0.86) \\
\text{hMSH2} & 0.91^* (0.87 to 0.94) & 0.95^{**} (0.92 to 0.99) & 0.86^* (0.83 to 0.89) & 0.80^* (0.76 to 0.84) \\
\text{hMSH6} & 0.92^* (0.89 to 0.95) & 0.96 (0.93 to 1.00) & 0.88^* (0.86 to 0.91) & 0.84^* (0.81 to 0.88) \\
\hline
\text{OSCC} & & & \\
\hline
\text{Well differentiated} & & & \\
\text{hMLH1} & 0.84^* (0.81 to 0.88) & 0.74^* (0.74 to 0.83) & \\
\text{hPMS2} & 0.87^* (0.84 to 0.90) & 0.82^* (0.78 to 0.86) & \\
\text{hMSH2} & 0.86^* (0.83 to 0.89) & 0.80^* (0.76 to 0.84) & \\
\text{hMSH6} & 0.88^* (0.86 to 0.91) & 0.84^* (0.81 to 0.88) & \\
\hline
\end{array}
\]

The reference group is normal; \( ^*p<0.0001, \ **p<0.05 \).

OSCC, oral squamous cell carcinoma.

Figure 3 Representative photomicrographs showing carcinoma in situ samples stained with (i) H&E, (ii) hMLH1, (iii) hPMS2, (iv) hMSH2 and (v) hMSH6. Magnification ×20. Image (vi) represents a high magnification (×40) of the insert indicated in image (v) demonstrating clear loss of hMSH6 in the stratum basale.
Table 3  Stepwise backward elimination method of multinomial logistic regression

<table>
<thead>
<tr>
<th>Variable</th>
<th>Mild (n=37)</th>
<th>Moderate/severe (n=34)</th>
<th>Well differentiated (n=86)</th>
<th>Poorly differentiated (n=27)</th>
</tr>
</thead>
<tbody>
<tr>
<td>HMLH1</td>
<td>0.92** (0.88 to 0.97)</td>
<td>0.97 (0.92 to 1.03)</td>
<td>0.92* (0.88 to 0.96)</td>
<td>0.92** (0.85 to 1.00)</td>
</tr>
<tr>
<td>HPMS2</td>
<td>0.94** (0.91 to 0.98)</td>
<td>0.98 (0.94 to 1.03)</td>
<td>0.94* (0.90 to 0.97)</td>
<td>0.89* (0.84 to 0.95)</td>
</tr>
<tr>
<td>hMSH2</td>
<td>0.96 (0.92 to 1.00)</td>
<td>0.97 (0.93 to 1.01)</td>
<td>0.93** (0.89 to 0.97)</td>
<td>0.86* (0.81 to 0.91)</td>
</tr>
<tr>
<td>Control variables</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age</td>
<td>1.02 (0.98 to 1.05)</td>
<td>1.04** (1.00 to 1.07)</td>
<td>1.06** (1.02 to 1.09)</td>
<td>1.07** (1.02 to 1.13)</td>
</tr>
</tbody>
</table>

The reference group is normal; *p<0.0001, **p<0.05. The general formula by which the model was designed is as follows: \( \ln \frac{\text{probability (mild dysplasia)}}{\text{probability (normal)}} = \frac{0.54 + 0.022 \text{age} - 0.075 \text{HMLH1} - 0.056 \text{HPMS2} - 0.034 \text{hMSH2}}{\text{probability (normal)}} \). Following equations were used for data shown in the table:

- \( \ln \frac{\text{probability (moderate to severe dysplasia)}}{\text{probability (normal)}} = 3.164 + 0.037 \text{age} - 0.028 \text{HMLH1} - 0.016 \text{HPMS2} - 0.032 \text{hMSH2} \)
- \( \ln \frac{\text{probability (well differentiated OSCC)}}{\text{probability (normal)}} = 13.808 + 0.050 \text{age} - 0.088 \text{HMLH1} - 0.070 \text{HPMS2} - 0.0569 \text{hMSH2} \)
- \( \ln \frac{\text{probability (poorly differentiated OSCC)}}{\text{probability (normal)}} = 16.659 + 0.079 \text{age} - 0.088 \text{HMLH1} - 0.102 \text{HPMS2} - 0.148 \text{hMSH2} \)

OSCC, oral squamous cell carcinoma.

Take home message

Although immunoexpression of all MMR proteins decreased with increasing lesion severity, isolated loss of expression of hMSH6 in the basal cell layer of carcinoma in situ lesions could potentially be a useful diagnostic biomarker for these lesions.

Acknowledgements  The authors thank the Australian Dental Research Foundation for funding this work.

Contributors  MJ: immunostaining; scoring; statistical analysis and preparing the first draft; help in designing the experiment and help in presenting the data. CSF: supervision, con

Funding  Australian Dental Research Foundation.

Competing interests  None.

Ethics approval  The Ethics Committees of the University of Queensland (2007001478) and the Royal Brisbane Hospital (HREC/10/QRBW336).

Provenance and peer review  Not commissioned; externally peer reviewed.

REFERENCES