

**UK survey of PD-L1 testing in NSCLC responses.**

Responses to questions requiring only selection of a pre-specified response are shown below.

<b>Survey question (total responses received, N)<sup>1</sup></b>	<b>Pre-specified response options (responses, n [%])</b>
Q1. Within your institution, are you the lead for thoracic pathology? (N=32)	<ul style="list-style-type: none"> <li>• <b>Yes</b> (26 [81.2])</li> <li>• <b>No</b> (6 [18.8])</li> </ul>
Q2. On average, how many PD-L1 tests for NSCLC patients are performed in your centre per week? (N=30)	<ul style="list-style-type: none"> <li>• <b>1–5</b> (11 [36.7])</li> <li>• <b>6–10</b> (9 [30.0])</li> <li>• <b>11–15</b> (4 [13.3])</li> <li>• <b>16–20</b> (3 [10.0])</li> <li>• <b>21–25</b> (2 [7.0])</li> <li>• <b>26–30</b> (1 [3.0])</li> <li>• <b>≥31</b> (0)</li> </ul>
Q3. How many pathologists in your institution are responsible for reporting on PD-L1 expression for NSCLC? (N=30)	<ul style="list-style-type: none"> <li>• <b>1</b> (4 [13.3])</li> <li>• <b>2</b> (7 [23.3])</li> <li>• <b>3–4</b> (14 [46.7])</li> <li>• <b>5–6</b> (4 [13.3])</li> <li>• <b>7–8</b> (1 [3.3])</li> <li>• <b>≥9</b> (0)</li> </ul>
Q4. If there are several pathologists involved, is the PD-L1 testing workload for NSCLC distributed evenly among these pathologists? (N=30)	<ul style="list-style-type: none"> <li>• <b>Yes</b> (18 [60.0])</li> <li>• <b>No</b> (8 [26.7])</li> <li>• <b>N/A (there is only one pathologist)</b> (4 [13.3])</li> </ul>
Q5. Does/Do the pathologist(s) reporting PD-L1 contribute to the service of thoracic pathology (i.e. are	<ul style="list-style-type: none"> <li>• <b>Yes</b> (30 [100])</li> <li>• <b>No</b> (0)</li> </ul>

they routinely reporting on thoracic material)? (N=30)	
Q6. Does your centre carry out reflex testing for PD-L1? (N=30)	<ul style="list-style-type: none"> <li>• <b>Yes</b> (27 [90.0])</li> <li>• <b>No</b> (3 [10.0])</li> </ul>
Q8. Which antibody clone are you using within your PD-L1 assay? (N=27)	<ul style="list-style-type: none"> <li>• <b>SP263</b> (16 [59.0])</li> <li>• <b>22C3</b> (11 [41.0])</li> <li>• <b>28-8</b> (0)</li> <li>• <b>SP142</b> (0)</li> <li>• <b>Other</b> (please specify) (0)</li> </ul>
Q9. Within what type of assay are you using this antibody clone? (N=27)	<ul style="list-style-type: none"> <li>• <b>A companion, trial-validated diagnostic assay</b> (26 [96.3])</li> <li>• <b>Any other form of laboratory-developed assay</b> (1 [3.7])</li> </ul>
Q10. At your centre, what interpretation training have the pathologists performing this assay received? (N=27)	<ul style="list-style-type: none"> <li>• <b>No training received</b> (0)</li> <li>• <b>Attended external certified training in person</b> (24 [88.9])</li> <li>• <b>Undertook distance learning/online certified training</b> (2 [7.4])</li> <li>• <b>Underwent internal training from a colleague who is certified for PD-L1 testing</b> (1 [3.7])</li> <li>• <b>Other</b> (0)</li> </ul>
Q11. Does your laboratory perform PD-L1 testing for any other centres? (N=27)	<ul style="list-style-type: none"> <li>• <b>Yes</b> (20 [74.1])</li> <li>• <b>No</b> (7 [25.9])</li> </ul>
Q12. What is your turnaround time (TAT) for PD-L1 testing? (N=27)	<ul style="list-style-type: none"> <li>• <b>≤1 day/24 hours</b> (3 [11.1])</li> <li>• <b>1–2 days</b> (8 [29.6])</li> <li>• <b>3–4 days</b> (8 [29.6])</li> <li>• <b>5–6 days</b> (4 [14.8])</li> </ul>

	<ul style="list-style-type: none"> <li>• <b>≥7 days</b> (4 [14.8])</li> </ul>
Q13. Do you subscribe to an external quality assessment (EQA) for your PD-L1 testing service? (N=27)	<ul style="list-style-type: none"> <li>• <b>Yes</b> (23 [85.3])</li> <li>• <b>No</b> (4 [14.8])</li> </ul>
Q14. Do you perform regular internal audits for PD-L1 positivity? (N=27)	<ul style="list-style-type: none"> <li>• <b>Yes</b> (19 [70.4])</li> <li>• <b>No</b> (8 [29.6])</li> </ul>
Q19. What type of cytology preparations do you test for PD-L1 IHC assays? (N=27)	<ul style="list-style-type: none"> <li>• <b>Do not test cytology preparations</b> (1 [3.7])</li> <li>• <b>Cell blocks</b> (25 [92.6])</li> <li>• <b>Cytology</b> (0)</li> <li>• <b>Both cell blocks and liquid-based preparations</b> (1 [3.7])</li> </ul>
Q20. Is the entire cytology sample used to create the cell block? (N=25)	<ul style="list-style-type: none"> <li>• <b>Yes</b> (14 [56.0])</li> <li>• <b>No</b> (11 [44.0])</li> </ul>
Q21. For PD-L1 expression in a tested sample, are you aware of the relevance of the ≥1% cut-off for IO therapy? (N=26)	<ul style="list-style-type: none"> <li>• <b>Yes</b> (26 [100])</li> <li>• <b>No</b> (0)</li> </ul>
Q22. For PD-L1 expression in a tested sample, are you aware of the relevance of the ≥50% cut-off for IO therapy? (N=27)	<ul style="list-style-type: none"> <li>• <b>Yes</b> (27 [100])</li> <li>• <b>No</b> (0)</li> </ul>
Q25. When reporting PD-L1 results, do you regularly comment on sample adequacy? (N=25)	<ul style="list-style-type: none"> <li>• <b>Yes, always (even when there are adequate numbers of cells for analysis, i.e., &gt;100 cells and staining is adequate)</b> (19 [76.0])</li> </ul>

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	<ul style="list-style-type: none"> <li>• <b>Yes, when there are inadequate numbers of cells (i.e., &lt;100 cells)</b> (4 [16.0])</li> <li>• <b>Yes, when the staining or fixation is not adequate for analysis despite sufficient cells being present in the sample</b> (0)</li> <li>• <b>No</b> (2 [8.0])</li> </ul>
Q26. When initially testing a tumour sample for PD-L1 expression, are you aware of the stage of disease in the NSCLC patient? (N=25)	<ul style="list-style-type: none"> <li>• <b>Yes, always</b> (2 [8.0])</li> <li>• <b>Yes, but only in some patients</b> (12 [48.0])</li> <li>• <b>No</b> (11 [44.0])</li> </ul>

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<sup>1</sup>Only questions with pre-specified responses are shown in this table; those requiring free-text responses are not shown.

IHC, immunohistochemistry; IO, immuno-oncology; N/A, not applicable; NSCLC, non-small cell lung cancer; PD-L1, programmed cell death ligand 1.