



OPEN ACCESS

Genetic associations with polycystic ovary syndrome: the role of the mitochondrial genome; a systematic review and meta-analysis

Almira Moosa, Meeladah Ghani, Helen Claire O'Neill

► Additional supplemental material is published online only. To view, please visit the journal online (<http://dx.doi.org/10.1136/jcp-2021-208028>).

Department of Maternal and Fetal Medicine, UCL Institute for Women's Health, University College London, London, UK

Correspondence to

Dr Helen Claire O'Neill, Institute for Women's Health, UCL, London, UK; helen.oneill@ucl.ac.uk

Received 25 October 2021

Accepted 26 July 2022

ABSTRACT

Background Polycystic ovary syndrome (PCOS) remains the most common female reproductive endocrine disorder. Genetic studies have predominantly focused on the role of the nuclear genome, while the contribution of mitochondrial genetics in PCOS remains largely unknown.

Aim This study aims to systematically evaluate the literature regarding the associations between the mitochondrial genome and PCOS.

Methods A literature search focused on PCOS and mitochondrial genetics was conducted on (1) MEDLINE, (2) EMBASE and (3) The Cochrane Library (CENTRAL and Cochrane Reviews). Search results were screened for eligibility, and data involving genetic variants of mitochondrial DNA (mtDNA) were extracted. Quantitative data were presented in forest plots, and where this was not possible, data were analysed in a qualitative manner. Quality of studies was assessed using the Q-Genie tool.

Results Of the 13 812 identified studies, 15 studies were eligible for inclusion, with 8 studies suitable for meta-analysis. Women with PCOS showed higher frequencies of a 9 bp deletion, and aberrant single nucleotide polymorphisms (SNPs) in the ND5, A6 and 7 transfer RNA-encoding genes. They also showed lower frequencies of two SNPs in the D-loop of the genome. Women with PCOS also exhibited significantly lowered mtDNA copy number.

Conclusion Women with PCOS harbour genetic variants in coding and non-coding regions of the mitochondrial genome. This may disrupt the electron transport chain and lead to oxidative stress, causing apoptosis of cells and further genetic damage. However, further studies of higher quality are required to confirm these associations.

PROSPERO registration number CRD42021267991.

INTRODUCTION

Polycystic ovary syndrome (PCOS) remains the most common female reproductive endocrine disorder, affecting 15%–20% of women of reproductive age.^{1,2} Typically showing signs of anovulation, hyperandrogenemia, dyslipidaemia and insulin resistance (IR), women with PCOS often experience infertility, and are predisposed to a multitude of conditions, including type 2 diabetes mellitus, cardiovascular disease, metabolic syndrome (MetS) and, in later years, endometrial cancer.^{2–4} As such, PCOS, although rooted in ovarian pathology, evolves gradually

TAKE HOME MESSAGES

- ⇒ There is a well established pool of literature exploring the associations between PCOS and genetic variants in the nuclear genome, but far less research focused on the influence of the mitochondrial genome.
- ⇒ Genetic variations in multiple parts of the mitochondrial genome may contribute to structural and functional differences in mitochondrial proteins, leading to oxidative stress in cells that then predisposes them to further damage.
- ⇒ Further research is required to appreciate the complexities of PCOS and its inheritance pattern.

to encompass complex multiorgan and multi-system dysfunction.

The pathogenesis of PCOS remains largely unknown, however two of the most influential factors thought to contribute to the development of PCOS are genetics and oxidative stress. As such, the next step towards understanding this multifaceted disorder is to consider the interface between these two systems, by evaluating the role of the mitochondrial genome in PCOS.

Hyperglycaemic states in women with PCOS, largely attributed to IR, place the mitochondria under a non-physiological amount of strain, exposing it to high levels of reactive oxygen species (ROS). Its close proximity to the electron transport chain (ETC) therefore leaves mitochondrial DNA (mtDNA) vulnerable to damage by oxidative stress, particularly given that mtDNA lacks protection from histones and chromatin.⁵ The mitochondrial genome is vulnerable to acquiring genetic changes, and it possesses a lowered capacity to manage them, by way of low-efficiency DNA repair mechanisms,⁶ given its dependence on a single DNA polymerase, DNA polymerase-gamma, for both replication and repair.^{7,8} Consequently, oxidative stress in PCOS can lead to mtDNA mutagenesis, and an array of metabolic changes. Given the vital role of the organelle, many regions of the mitochondrial genome remain evolutionarily conserved,⁹ providing scope for small-scale genetic variants to have large implications in the pathogenesis of this disease.

While it has become accepted that genetic associations exist with PCOS, the majority



© Author(s) (or their employer(s)) 2022. Re-use permitted under CC BY-NC. No commercial re-use. See rights and permissions. Published by BMJ.

To cite: Moosa A, Ghani M, O'Neill HC. *J Clin Pathol* Epub ahead of print: [please include Day Month Year]. doi:10.1136/jclinpath-2021-208028

of research thus far has focused on the nuclear genome, resulting in a sizeable number of studies exploring nuclear-encoded factors and their influence on PCOS. Despite this large pool of literature, however, few definitive associations have been documented, leaving many questions unanswered about the true genetic basis of the disease. Owing to the lack of answers therefore, attention in recent years has been redirected towards the mitochondrial genome, which, up until now, remained largely overlooked.

Following this redirection in research, there has been an increase in the number of studies considering the role of mitochondrial genetics in PCOS, providing new insights into PCOS as a disease, and warranting a systematic review of the evidence on this emerging concept.

As such, this study aims to systematically evaluate the literature regarding the associations between the mitochondrial genome and PCOS. Outcomes of interest for this study are:

- ▶ Genetic variants in coding regions of the mitochondrial genome.
- ▶ Genetic variants in the non-coding region of the mitochondrial genome.
- ▶ Changes in mitochondrial genetic content.
- ▶ Quality of included studies.

To the authors' knowledge, this is the first systematic review to consider the mitochondrial genome in the context of PCOS.

METHODS

This study was conducted in accordance with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses guidelines,^{10 11} and is registered in the International Prospective Register of Systematic Reviews (PROSPERO), with registration number CRD42021267991.

Search strategy

A literature search was carried out via (1) MEDLINE, (2) EMBASE and (3) The Cochrane Library (CENTRAL and Cochrane Reviews) electronic databases, using subject headings and keywords relating to "polycystic ovary syndrome" and "mitochondrial genetics" (online supplemental 1). Limits were applied to return only English language and human studies.

Searches were conducted by two reviewers independently, with disagreements being resolved by discussion. Publications spanning all years were considered, with the last search being conducted on 19 July 2021. Citations were exported to EndNote X9 for management, and duplicates were removed. Search results were then screened for eligibility.

Reference lists of included studies and review articles were manually searched to identify articles not sought through electronic searches, and authors were contacted regarding papers with restricted access.

Eligibility criteria and study selection

Two reviewers independently assessed eligibility of studies for inclusion, with discrepancies being resolved by discussion. Publications were initially screened by title and abstract, with seemingly relevant papers then being considered in full.

Studies were included if they met the following criteria:

1. *Population* considered women with a clinical diagnosis of PCOS.
2. *Interventions* included genetic investigations of the mitochondrial genome.

3. *Comparison* was made to a suitable control group of women without PCOS.
4. *Outcomes* included any measures relating to genetic content.
5. *Study design* of any type.

Thus, studies focused on animals, the nuclear genome, other reproductive conditions apart from PCOS, and those which did not carry out genetic techniques were excluded. Review articles, abstract-only articles, reply letters and inaccessible papers were also excluded.

Data extraction

Data regarding the presence and frequencies of mutations/polymorphisms in both the coding and non-coding regions of the mitochondrial genome, as well as data reporting mtDNA copy numbers (mtDNA CN), were extracted from studies. Information regarding authorship, geographical location and sample size was also extracted.

In cases where data were only available in graphical form, Automeris-WebPlotDigitizer, a web-based software tool designed specifically for graph reading, was employed to improve the accuracy of the values extracted.

In order to optimise the completeness of data, study authors were contacted regarding missing datasets, and supplementary data sources were searched where possible.

Data synthesis and statistical analysis

All data were managed by the author, with professional advice sought from an independent statistician. Of the 15 studies included, 8 had sufficient data to be considered for various meta-analyses. Forest plots and graphical representation of figures were carried out using Review Manager V.5.4.1 statistical analysis software. Heterogeneity between studies was quantified by Cochran's Q scores and inconsistency square (I^2) measures, with values of $I^2 > 50\%$ indicating significant heterogeneity.

Quantitative data regarding genetic variants were processed to represent data as 'percentage frequency of a given polymorphism (%)' in both PCOS and control groups. Where possible, these data were shown in forest plots, and reported as ORs, along with respective 95% CIs, p values and heterogeneity (I^2) scores.

Data regarding mtDNA content were processed to represent data as 'log-transformed mean mtDNA CN' (log (mean mtDNA CN)), and studies showing suitable measures of spread, such as SD or SEM, were used to construct a forest plot. Subgroup analyses were performed to consider separately studies which assessed mtDNA CN in blood cells, compared with one study which assessed mtDNA CN in granulosa cells. A pooled estimate of the standardised mean difference (SMD) in log (mean mtDNA CN) was calculated, again with a 95% CI, p value and I^2 score.

Where quantitative measures were not feasible, data were reported and analysed in a qualitative manner. Despite original intentions, funnel plots could not be constructed to assess risk of publication biases, owing to the limited numbers of studies considering each outcome.

Quality assessment

The quality of all included studies was assessed using the Q-Genie Tool for Quality Assessment of Genetic Association Studies,^{12 13} considering the following 11 categories:

1. Rationale for study
2. Selection and definition of outcomes of interest

3. Selection and comparability of comparison groups
4. Technical classification of exposure
5. Non-technical classification of exposure
6. Sources of bias
7. Sample size and power
8. A priori planning of analyses
9. Statistical methods and control for confounding
10. Testing of assumptions and inferences for genetic analyses
11. Appropriateness of inferences drawn from results

Studies were awarded scores between 1 (poor) and 7 (excellent) for each category. Total quality scores were calculated for each study, with scores ≤ 35 indicating poor quality, scores >35 and ≤ 45 indicating moderate quality and scores >45 indicating good quality. Median scores and IQRs for all 11 categories were also calculated.

RESULTS

Search results

The overall search results are summarised in figure 1. A total of 13 812 records were identified through searching, spanning years 1964–2021. Following the removal of 3767 duplicates, 10 045 records were screened for eligibility, first by title, excluding 5938 records, then by abstract, excluding a further 4026 records. After considering the remaining 81 full-text records, a further 67 records were excluded, thus leaving 14 studies eligible for inclusion. After searching reference lists, one additional study was added. Overall, this systematic review included 15 studies (shown in online supplemental table, online supplement 2),^{14 15} considering a total of 2053 participants (1125 PCOS, 928 control). Of these studies, eight had suitable data to be included in the meta-analysis.

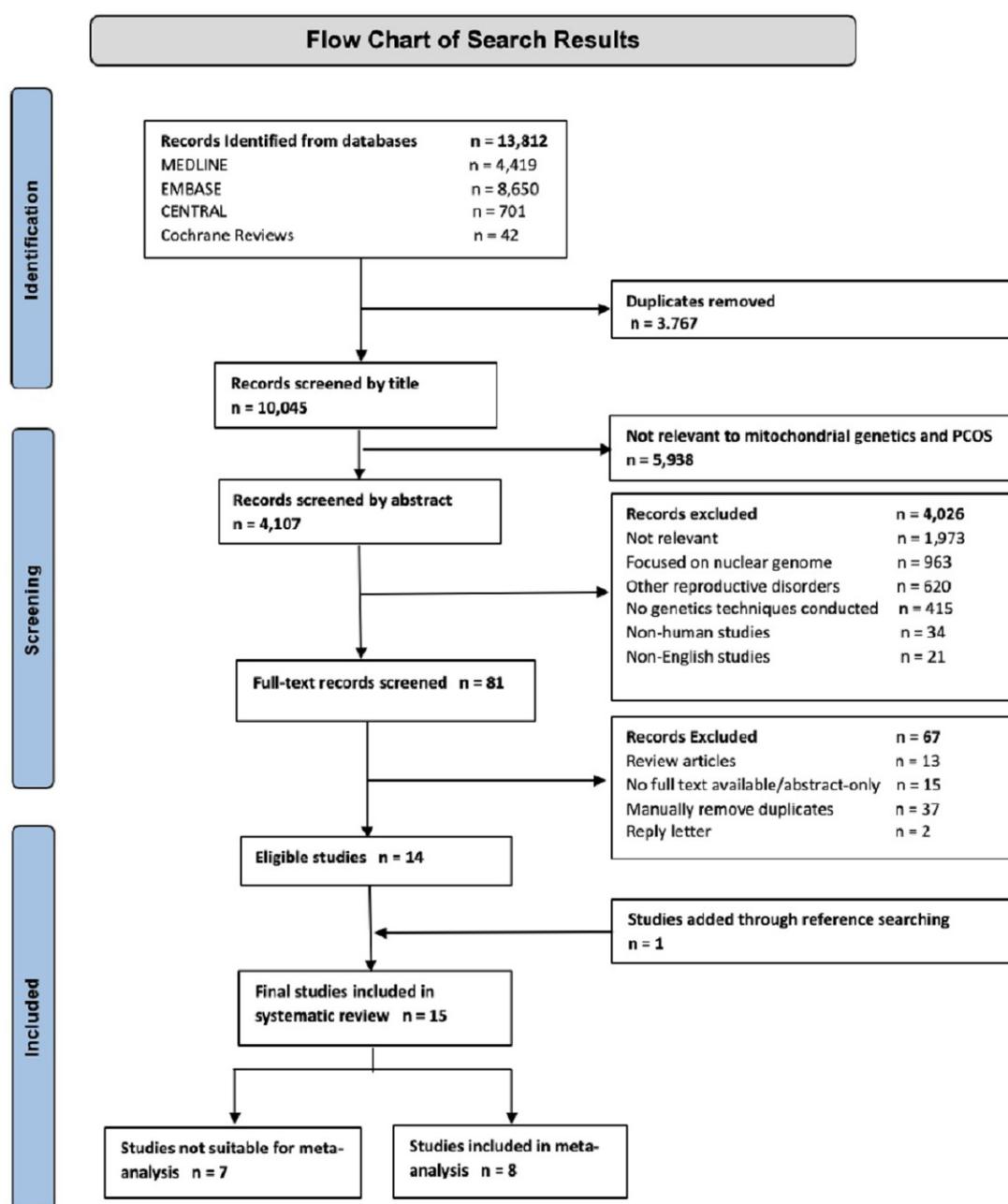


Figure 1 Flow chart of search results and included studies. PCOS, polycystic ovary syndrome.

Genetic variants in coding regions of the mitochondrial genome

Polymorphisms in the coding region of the mitochondrial genome were reported by seven studies.

Nine base pair deletions

Four studies documented the presence of a 9 bp deletion (CCCCCTCTA) in region V of the mitochondrial genome.^{16–19} While all four studies agreed that this deletion occurred more frequently in women with PCOS than in controls, only one study quantified this effect, showing this deletion to occur in 23.5% of cases compared with 7.1% of controls, and reported a p value of 0.02, indicating a significant relationship.¹⁸

Polymorphisms in the ND5 and A6 genes

The full list of all the genetic polymorphisms detected in other coding regions is available in online supplemental 3.

Frequently reported variants included the T12811C and T12338C polymorphisms in the ND5 gene,^{17 20 21} and the

G8584A and C8684T polymorphisms in the A6 gene.^{17 18 20}

While sufficient numerical data were not available to calculate the frequencies of these polymorphisms, these mutations were detected only in PCOS cases, and not in controls, eluding to their potential significance.

Polymorphisms in genes coding for mitochondrial tRNAs and rRNAs
All data extracted considering mt-tRNA genes are available in online supplemental 4.

Polymorphisms appeared to occur most frequently in genes coding for the following seven mt-tRNAs: tRNA^{Cys}, tRNA^{Leu}, tRNA^{Glu}, tRNA^{Gln}, tRNA^{Lys}, tRNA^{Arg} and tRNA^{Asp}. The three studies with sufficient data were used to determine the frequency of polymorphisms in these tRNAs, which are summarised in figure 2A.^{17 20 22} tRNA^{Cys} gene polymorphisms were more frequent in PCOS cases than in controls, occurring in 2.9% of cases vs 1.1% of controls. This was also the case for polymorphisms in genes coding for tRNA^{Leu}, which occurred in 2.7% of cases, but only in 0.6%

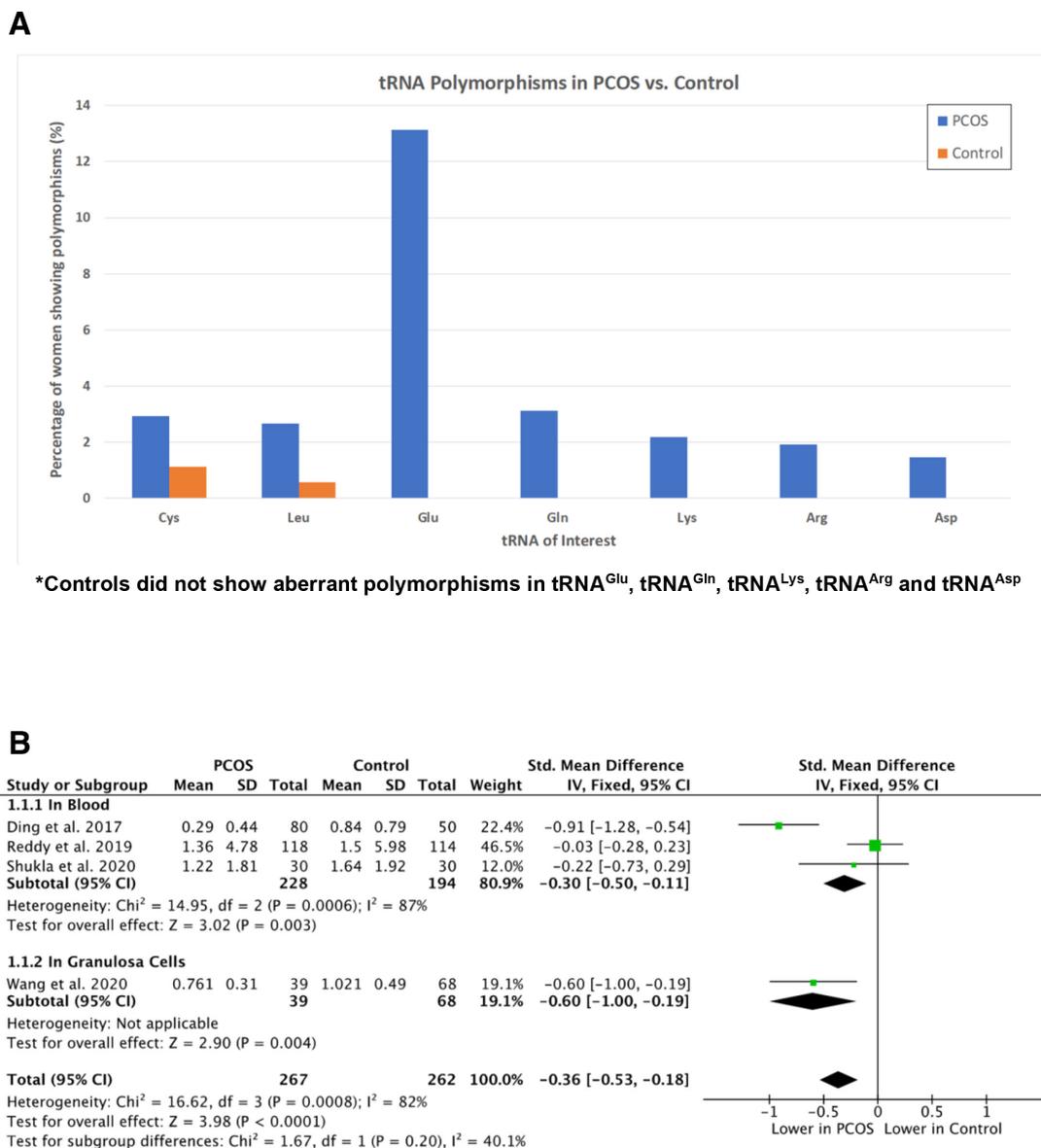


Figure 2 (A) Bar graph of mitochondrial transfer RNA (tRNA) gene polymorphisms. (B) Log-transformed mean mitochondrial DNA (mtDNA) copy number in polycystic ovary syndrome (PCOS) cases versus controls.

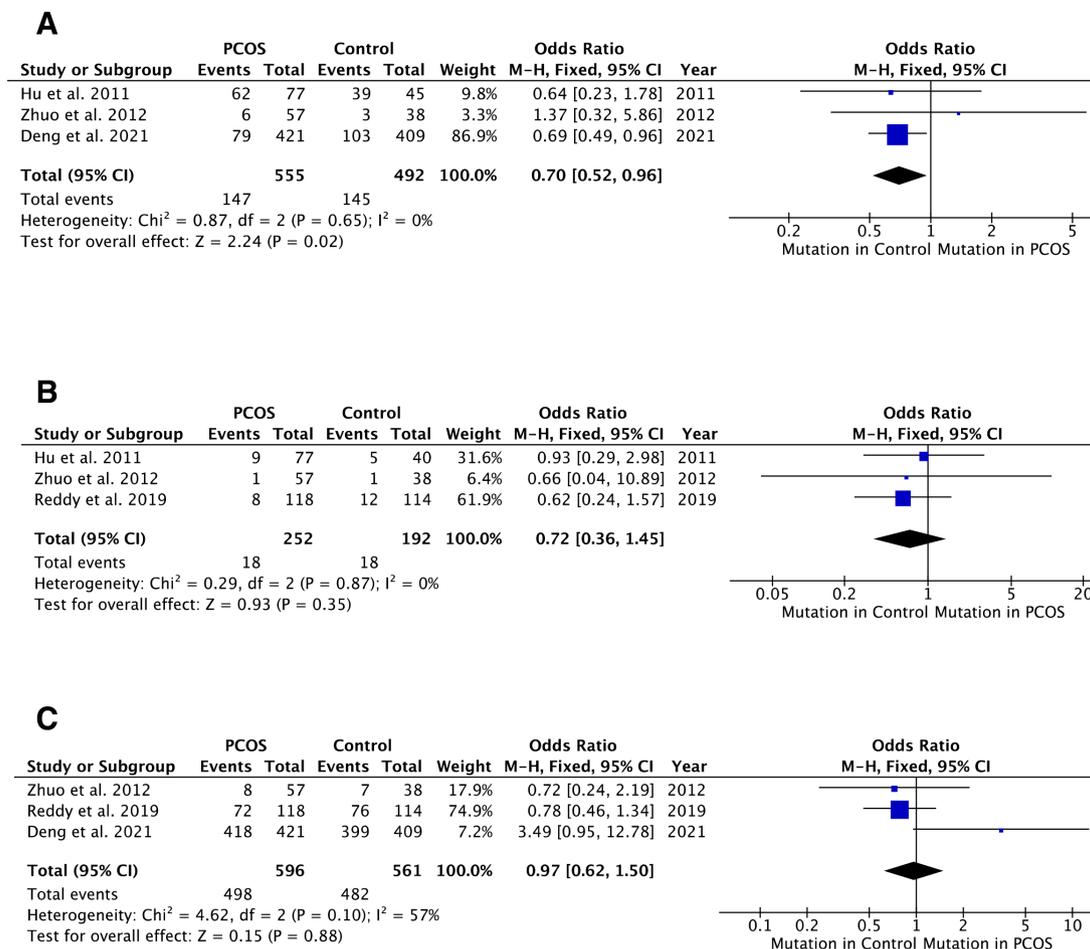


Figure 3 (A) C150T D-loop polymorphisms in polycystic ovary syndrome (PCOS) cases versus controls. (B) T146C D-loop polymorphisms in PCOS cases versus controls. (C) A263G D-loop polymorphisms in PCOS cases versus controls.

of controls. Polymorphisms in genes coding for tRNA^{Glu}, tRNA^{Gln}, tRNA^{Lys}, tRNA^{Arg} and tRNA^{Asp} were detected only in PCOS cases and not in controls, occurring at frequencies of: 13.1%, 3.1%, 2.2%, 1.9% and 1.5%, respectively. Overall, mt-tRNA gene polymorphisms reported in PCOS cases were either absent, or less frequent, in controls.

Polymorphisms in genes coding for both the 12S and 16S rRNAs were also detected in women with PCOS (online supplemental 5).^{17–19 23}

Genetic variants in non-coding regions of the mitochondrial genome

Polymorphisms in the mitochondrial D-loop

D-loop polymorphisms were considered by eight studies, with all extractable data shown in online supplemental 6. The following five polymorphisms were reported in two or more studies: C150T, T146C, A263G, A189G, D310. However, only three of these polymorphisms, C150T, T146C and A263G, had sufficient data available to be considered for meta-analysis (figure 3).

As shown in figure 3A, the C150T D-loop polymorphism occurred at a significantly lower frequency in PCOS cases compared with controls, with an overall OR of 0.70 (95% CI 0.52 to 0.96, $p=0.02$). While the T146C polymorphism (figure 3B) also occurred at a lower frequency in PCOS cases than in controls, with an overall OR of 0.72 (95% CI 0.36 to 1.45, $p=0.35$), this result is not considered to be significant.

As shown by the OR of 0.97 (95% CI 0.62 to 1.50, $p=0.88$), the A263G polymorphism (figure 3C) in the D-loop of the mitochondrial genome occurred at similar frequencies in both cases and controls, again indicating a result that is not significant.

Overall, women with PCOS showed lower odds of possessing the C150T polymorphism, potentially lower odds of possessing the T146C polymorphism and almost the same odds of possessing the A263G polymorphism, as controls.

One study also detected the A189G and D310 polymorphisms almost twice as often in women with PCOS compared with controls, with the A189G polymorphism present in 26.3% of PCOS cases vs 13.2% of controls, and the D310 polymorphism present in 54.2% of cases vs 28.9% of controls.⁶ Interestingly, women with PCOS who showed these two specific polymorphisms appeared to have significantly lower mtDNA CNs than those who did not show these polymorphisms.

Changes in mitochondrial genetic content

Mitochondrial genetic content, reported as log (mean mtDNA CN), was assessed by nine studies (online supplemental 7).

While all nine studies showed lowered mtDNA CNs in women with PCOS, one study did not report data for the control group and four did not report suitable measures of spread to be considered for meta-analysis. The data from the four remaining studies,^{6 20 24 25} considering a total of 529 subjects (267 PCOS,

262 control), are summarised in figure 2B. Subgroup analyses were adopted to consider separately three studies which assessed mtDNA CN in blood leucocytes, compared with one study which assessed mtDNA CN in granulosa cells.

Log (mean mtDNA CN) was significantly reduced in PCOS cases compared with controls, with an overall SMD of -0.36 (95% CI -0.53 to -0.18 , $p < 0.0001$). Considering the subgroup analyses, log (mean mtDNA CN) was shown to be significantly reduced in both blood and granulosa cells with SMD values of -0.30 (95% CI -0.50 to -0.11) and -0.60 (95% CI -1.00 to -0.19), respectively. The value of $I^2 = 82\%$ does, however, indicate significant between-study heterogeneity.

Quality assessment

Quality assessment was carried out for all included studies, with each study being assigned an overall quality score (Supplement 8). The majority of papers (6/15, 40%) were found to be of moderate quality, while the minority of papers (4/15, 26.7%) were deemed to be of poor quality. An intermediate number of papers (5/15, 33.3%) were regarded as good quality.

The highest scoring category was 'rationale for study' (median 6, IQR 4–7), while the two poorest performing categories included 'other sources of bias' (median 2, IQR 1–3), and 'sample size and power' (median 2, IQR 1–2).

DISCUSSION

A current gap in literature exists relating to a systematic review of the associations between mitochondrial genetics and PCOS. As such, this study aimed to assess genetic variants in both the coding and non-coding regions of the mitochondrial genome, and changes in mitochondrial genetic content in women with PCOS, while evaluating the quality of evidence overall.

Polymorphisms in coding regions of the mitochondrial genome

In the majority of controls without PCOS, the 9 bp sequence (CCCCCTCTA) in region V of the mitochondrial genome was repeated twice in tandem. Many women with PCOS, however, showed deletions in this region, leaving them with only one copy of this sequence. These deletions are suspected to have arisen from DNA replication slippage errors made by DNA polymerase-gamma, which possesses underdeveloped repair mechanisms in mtDNA.^{18 26} Interestingly, Hu *et al* found these deletions to be significantly associated with higher serum levels of glucose during oral glucose tolerance testing, and lower insulin sensitivity indices,¹⁶ suggesting that the deletions may be an underlying factor contributing towards insulin-resistant states in patients with PCOS.

The ND5 T12811C and T12338C polymorphisms, as well as the A6 G8584A and C8684T polymorphisms, were detected only in PCOS cases, and not in controls. These polymorphisms were found to cause amino acid substitutions,^{17 21} and could therefore bear important implications. Amino acid changes result in altered primary, and potentially secondary/tertiary/quaternary structures of proteins. This could subsequently impact the ETC, which itself is a series of proteins, and could then lead to inefficiencies in electron transfer and higher levels of ROS. These genetic variants, given that they were only found in cases, may therefore contribute to the widespread metabolic dysfunction witnessed in PCOS.

Interestingly, all four single nucleotide polymorphisms (SNPs) listed above have been associated with Leber's hereditary optic neuropathy syndrome,^{27–29} with the ND5 T12338C mutation

also being linked to neonatal deafness.³⁰ Both of these disorders showcase failures in energy metabolism, and are known to be mitochondrially, and therefore maternally, inherited. This may suggest that some characteristics of PCOS, having shown such similarities in mitochondrial genetics and energy disruptions (although to a lesser extent), may also be attributed to a more maternal pattern of inheritance, by way of the mitochondrial genome.

Genes coding for tRNA and rRNA

Polymorphisms in genes coding for tRNAs and rRNAs were present and much more frequent in PCOS cases compared with controls. This too could bear consequences for women with PCOS, considering the essential roles of tRNAs and rRNAs in the translation of mitochondrial-encoded proteins. The majority of detected variants occurred in regions that remain highly conserved among vertebrate species, eluding to their ability to alter pivotal mitochondrial processes, and to result in biochemical changes and oxidative stress, as seen in PCOS.

Some polymorphisms reported in women with PCOS, including the tRNA^{Gln} T4395C and tRNA^{Leu} A3302G polymorphisms,^{17 23} occurred in the acceptor arms of tRNA molecules—the regions which attach to amino acids in the process of aminoacylation. Such genetic variants may therefore possess the ability to disrupt amino acid-tRNAs binding, subsequently impacting the assembly of polypeptide chains. Other polymorphisms, such as the tRNA^{Asp} A7543G, and tRNA^{Leu} T4363C polymorphisms, occurred in the anticodon stem of tRNA molecules,²⁰ and could potentially disrupt base-pairing with mRNA transcripts,²⁰ and cause inefficiencies in the link between transcription and translation of mitochondrial-encoded proteins. Other SNPs, including the tRNA^{Lys} A8343G, tRNA^{Arg} T10454C and tRNA^{Glu} A14693G SNPs,²⁰ were localised to the T-loop of tRNA molecules, which is an important recognition site that allows tRNA molecules to bind to mitochondrial ribosomes, which themselves appeared to bear genetic polymorphisms in women with PCOS.

Current data suggest that these polymorphisms confer structural and functional changes in the mitochondrial protein synthesising machinery, and result in destabilisation of tRNA and rRNA molecules,³¹ loss of nucleotide modifications and deficient aminoacylation,^{32 33} all of which may affect tRNA-amino acid binding, and tRNA-ribosome complex formation.³⁴ Such alterations in protein synthesis may once again affect complexes that make up the ETC, and may offer possible explanations for the altered enzyme levels seen in women with PCOS.³² As evidence of this, women carrying these mutations were found to have lower levels of superoxide dismutase, a key enzyme and hence protein, as well as higher levels of oxidative stress markers, such as 8-hydroxy-2'-deoxyguanosine and malondialdehyde.²⁰ Women with these tRNA mutations also showed lower mtDNA CNs, eluding to disturbances in mitochondrial equilibrium.²⁰

mt-tRNA gene polymorphisms also demonstrated links to other mitochondrially inherited diseases, with similar biochemical and metabolic profiles to those of PCOS. In a familial study, Ding *et al* suggested that the tRNA^{Leu} C3275T, tRNA^{Gln} T4363C and tRNA^{Lys} A8343G SNPs were associated with maternally inherited MetS,¹⁹ while in another familial study it was suggested that the tRNA^{Leu} A3302G SNP was associated with maternally inherited IR.²³ This evidence further supports that the pathology presented in PCOS may be partly rooted in mitochondrial genetics.

Polymorphisms in the non-coding region of the mitochondrial genome

Unlike the above polymorphisms, the D-loop C150T polymorphism and, more often than not, the T146C polymorphism, occurred at higher frequencies in controls, suggesting that these polymorphisms may have a somewhat protective element that contributes to a slightly lowered risk of PCOS.³⁵ Interestingly, the T146C polymorphism has also been associated with lowered risk of endometriosis,^{36,37} indicating a potentially key polymorphism in wider female reproduction. In contrast, the A263G polymorphism occurred at similar frequencies in both cases and controls, with some studies indicating a significant relationship and others not, summing to show no overall effect. Inconsistencies between studies could have arisen due to variations in the prevalence of this SNP, owing to ethnic or environmental differences.

While genetic variants in the D-loop do not affect mitochondrial-encoded proteins directly, they have the ability to alter gene expression regulation, and to bring about changes in mtDNA replication and transcription, given that the D-loop contains the origins of replication of the mitochondrial genome.³⁸ As such, genetic polymorphisms in the D-loop have the capacity to influence mitochondrial dynamics, and may therefore contribute further to ROS production in PCOS.

As an example of this, women with PCOS carrying the D-loop A189G and D310 polymorphisms were shown to have significantly lower mtDNA CNs than those not carrying these polymorphisms.^{6,39} The D310 region is located within one of the key conserved regions of the genome (CSB-II) essential for mtDNA replication,^{6,40} offering a potential explanation as to why PCOS cases with D310 variants show lowered mtDNA CNs, and subsequently experience oxidative stress.

Apart from regulating the mitochondrial genome, the D-loop also enables nuclear-mitochondrial crosstalk. Multiple nuclear hormone receptors, including the glucocorticoid receptors, thyroid receptors (T3) and oestrogen receptors, bind to regions of the D-loop as part of their normal physiological function,^{24,41} creating scope for genetic variants in these regions to influence hormone profiles in women with PCOS, and the endocrine system at large.

Mitochondrial DNA content

Compared with controls, PCOS cases showed significantly lowered mtDNA CNs, indicating a reduction in both mitochondrial number and mass overall. This is likely to be a result of autophagy,⁴² a process by which cells degrade their damaged organelles, including mitochondria, by lysosomal action.⁴³ This decrease may be testament to the effects of harbouring these various genetic variants, which act to increase ROS production to the point of cell death. The reduction of mitochondrial content in both blood and granulosa cells suggests that this mitochondrial dysfunction affects cells of the ovary, and bears more widespread consequences in women with PCOS, causing this syndrome to present with multiorgan and multisystem involvement.

Studies have shown that maintenance of mtDNA CNs is essential for preservation of mitochondrial function and cell growth.⁴⁴ A decrease in mitochondrial content translates to a subsequent decrease in the expression of mitochondrial-encoded genes and proteins. This means that a lowered number of mitochondria and mtDNA-encoded enzymes are left to manage the raised level of ROS after damaged mitochondria are removed, predisposing the remaining mitochondria to further damage, oxidative stress

and inevitable autophagy. Therefore, a diminished mtDNA CN in PCOS may be a result of increased ROS generation, and may contribute further towards it, by feeding into the 'vicious cycle' of ROS-induced ROS release.⁴⁵

Patients with PCOS with lowered mtDNA CNs have also shown to have higher degrees of IR, as well as lowered levels of sex hormone binding globulin,⁴⁶ suggesting that mtDNA depletion could be involved in the pathogenesis of this syndrome, linking changes in genetic expression to hormonal and homeostatic imbalances.

These findings are further supported by multiple animal studies, which showed abnormalities in mitochondrial structure and function, and reported alterations in expression of mtDNA-encoded genes as a result of lower mtDNA CNs.^{47,48} Reduced mtDNA content has also been linked to poor oocyte quality,⁴⁹ and as such, could contribute to subfertility or infertility in women with PCOS.

The mitochondrial genome in women with PCOS

Overall, women with PCOS showcased polymorphisms in both coding and non-coding regions of the mitochondrial genome, and showed a reduction in the amount of mtDNA present. These genetic variants have the ability to affect mitochondrial function on multiple levels, be it at the level of replication and transcription through D-loop polymorphisms or slippage errors, or at the level of translation, by way of polymorphisms in genes coding for proteins, tRNAs and rRNAs.

Based on the above findings, this study proposes that this 'multilevel' genetic dysregulation sets up a cycle of events that then predisposes to further genetic damage and mitochondrial destruction, with the cycle unfolding as follows: initial genetic variants contribute to impairment of ETC function,³² which subsequently leads to less efficient electron transfer, and therefore increased rates of ROS generation, bringing about oxidative stress. This oxidative stress then burdens cells to the point of apoptosis,^{42,43} resulting in lowered mitochondrial content, which subsequently leaves a reduced pool of mitochondria in an environment of increased oxidative stress, thus leading to further mtDNA damage (figure 4).⁵⁰⁻⁵⁴ This proposed cycle draws parallels with the concept of ROS-induced ROS release,⁴⁵ but additionally accounts for the presence of aberrant genetic variants in the mitochondrial genomes of women with PCOS.

Quality assessment and critical appraisal of studies

Studies scored highly in the category entitled 'rationale for study', which subsequently translated to the majority of studies possessing clear hypotheses. Studies also benefitted from having well-defined eligibility criteria for cases, with all but two studies characterising PCOS cases as per the revised 2003 Rotterdam criteria.⁵⁵ Studies also showcased well-structured methodology, and use of accredited statistical software, showing good a priori planning of analyses. Studies achieving the highest scores were those which documented better genetic techniques, for example, those which detailed efforts to normalise expression of target genes relative to housekeeping genes (β -globin/ β -actin/GAPDH).

However, studies appeared to be insufficiently powered by generally small sample sizes, and did not appropriately account for possible sources of bias. Many studies recruited cases from hospitals or fertility clinics. Therefore, in order to be considered as cases, women would have had to experience subfertility/infertility, or symptoms considerable enough to seek professional intervention, arguably representing only the most severe cases of

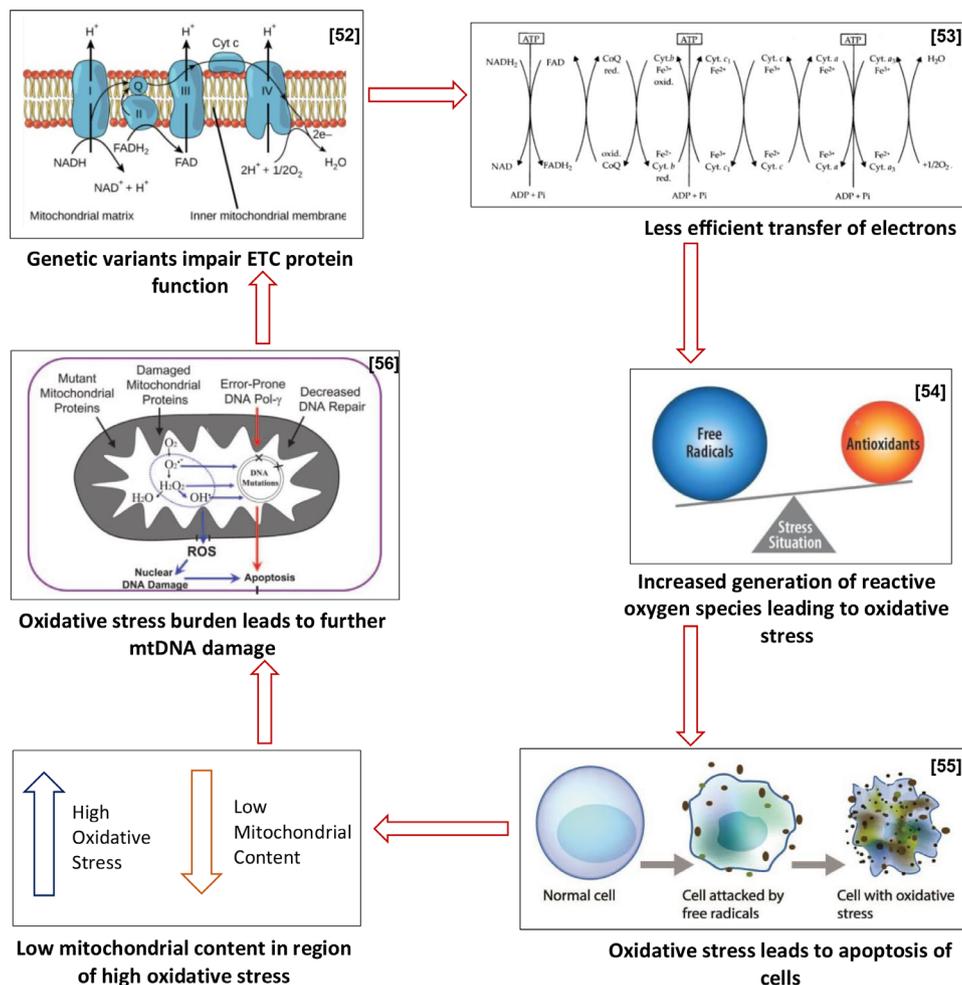


Figure 4 Proposed cycle of events in women with polycystic ovary syndrome (PCOS). ETC, electron transport chain; mtDNA, mitochondrial DNA; ROS, reactive oxygen species.

PCOS. By only considering women at one end of the spectrum of what is a largely heterogeneous disorder, studies were made vulnerable to selection bias, as cases sampled may not be representative of wider PCOS populations.

Similarly, while studies recruited age-matched and BMI-matched controls, other important characteristics, such as smoking—a factor known to cause genetic mutagenesis, and diet—a factor important in insulin-resistant PCOS states, were less regulated, potentially introducing heterogeneity between participants.

A minority of studies also failed to show records of all collected data raising concerns around publication biases. Owing to the limited number of papers, however, the use of funnel plots to assess risks of publication biases was deemed inappropriate for this study.

Moreover, only eight studies were fit for inclusion in various meta-analyses, with other studies showcasing insufficient data to undergo appropriate statistical analysis. Some studies successfully reported mtDNA CNs, but failed to report suitable measures of spread to be included in forest plots. Other studies documented an array of genetic variants, but failed to report numerical data that allowed for the frequencies of polymorphisms to be calculated. As such, much of the extracted data either could not be analysed to their full potential, or were limited by their qualitative nature. While the outcomes of such

papers were still assessed qualitatively, these findings could not be quantified in a manner that allowed for statistical validation of results.

Overall, only 5/15 (33.3%) of studies were found to be of good quality. This has potential implications on the outcomes of this review, as it becomes difficult to gauge the true relationship between the mitochondrial genome and PCOS.

Strengths and limitations

To the authors' knowledge, this is the first systematic review to explore the relationship between the mitochondrial genome and PCOS. This study addresses a gap in previous literature, and focuses on a newly emerging concept, making it significant to ongoing and future research. However, by restricting the search to English-only results, and by only accounting for literature published on online platform, some otherwise relevant studies may have been excluded from this review. By its design, this review was also based on the assumption that data presented in primary studies were valid and accurate. Moreover, this review, much like its included studies, considered genetic variants in the mitochondrial genomes of women who had already obtained a diagnosis of PCOS, making it difficult to distinguish cause from effect, and to gauge the true association between the mitochondrial genome and PCOS.

Future implications

In clinical practice, women with PCOS are treated with metformin to encourage normalised glucose usage in cells.⁵⁶ This study has shown that if this regulation is not achieved, blood leucocytes will experience oxidative stress and undergo apoptosis, leading to reduced mtDNA CN. In theory, this could mean that mtDNA CNs measured in blood samples could be used as a minimally invasive way to monitor metformin treatment and efficacy in women with PCOS, with lower mtDNA CN indicating higher rates of cell death and less effective treatment. Women are also commonly advised to incorporate foods with lower glycaemic indices into their diets. Given the capacity of oxidative stress to disrupt mitochondrial equilibrium and genetics, women with PCOS may also be advised to increase intake of foods rich in anti-oxidants.

Future recommendations

Due to the aforementioned data limitations, this study proposes the implementation of a standardised protocol for reporting in genetic association studies. Abiding by such a protocol will allow uniform reporting of data that can then be processed to quantify the relationship between the mitochondrial genome and PCOS more accurately.

Future research should also aim to explore different aspects of the mitochondria in the context of PCOS. While many studies considered mitochondrial content in blood leucocytes, only two were found to consider mitochondrial content in granulosa cells, with only one being suitable for meta-analysis. To allow for larger subgroup analyses, and to uncover a more holistic relationship, it is recommended that more studies investigate mitochondrial genomes in granulosa cells, and in other cell types involved PCOS, such as beta-pancreatic cells given their role in insulin disturbances. It is also worth noting that the majority of the studies were conducted in Asian populations, and it is necessary to explore this relationship in women of other ethnicities and genetic backgrounds.

More familial studies, carrying out genetic screening of maternal mitochondrial genomes, could help decipher whether mitochondrial genetic variants are germline or acquired changes, and may help determine a cause-effect relationship.

Moreover, the mitochondrial genome, although distinct from the nuclear genome, does not function independently of nuclear-encoded proteins. Future research should therefore aim to explore mitochondrial genetics in women with PCOS in a more expansive manner to account for nuclear-mitochondrial crosstalk.

These recommendations would provide valuable insight on the topic of mitochondrial genetics and PCOS, and would allow for this relationship to be put into context with respect to wider bodily function and inheritance patterns.

CONCLUSION

Women with PCOS appear to harbour genetic variants in both coding and non-coding regions of their mitochondrial genomes, including 9 bp deletions and an array of specific SNPs. These are thought to contribute to aberrant protein synthesis of mitochondrial elements, and to result in oxidative stress in cells, leading to autophagy, and thus, lowered mitochondrial content. This is likely to feed into a cycle of

ROS-induced ROS release, predisposing to further mitochondrial dysfunction.

Given that only 33.3% of analysed studies were found to be of good quality, however, further research, reporting more quantitative data regarding different cell types and women of different ethnicities, is required to unveil the true association between mitochondrial genetics and PCOS, and to understand future management of this complex syndrome.

Handling editor Mona El-Bahrawy.

Twitter Helen Claire O'Neill @DrHelenO'Neill

Acknowledgements The authors would like to thank Dr Aviva Petrie for her professional advice regarding data management and statistical analyses, and for her educational support. The authors would also like to thank the authors of the various studies included in this systematic review, for providing the data and framework on which this study was based.

Contributors Conception and design of the work; HCO'N. Acquisition, analysis and interpretation of data for the work: HCO'N, AM and MG. Drafting the work or revising it critically for important intellectual content: AM and HCO'N. Final approval of the version to be published: HCO'N.

Funding The authors have not declared a specific grant for this research from any funding agency in the public, commercial or not-for-profit sectors.

Competing interests None declared.

Patient consent for publication Not applicable.

Ethics approval Not applicable.

Provenance and peer review Not commissioned; externally peer reviewed.

Supplemental material This content has been supplied by the author(s). It has not been vetted by BMJ Publishing Group Limited (BMJ) and may not have been peer-reviewed. Any opinions or recommendations discussed are solely those of the author(s) and are not endorsed by BMJ. BMJ disclaims all liability and responsibility arising from any reliance placed on the content. Where the content includes any translated material, BMJ does not warrant the accuracy and reliability of the translations (including but not limited to local regulations, clinical guidelines, terminology, drug names and drug dosages), and is not responsible for any error and/or omissions arising from translation and adaptation or otherwise.

Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, appropriate credit is given, any changes made indicated, and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>.

ORCID iD

Helen Claire O'Neill <http://orcid.org/0000-0003-3481-3459>

REFERENCES

- McGowan MP. Polycystic ovary syndrome: a common endocrine disorder and risk factor for vascular disease. *Curr Treat Options Cardiovasc Med* 2011;13:289–301.
- Sirmans SM, Pate KA. Epidemiology, diagnosis, and management of polycystic ovary syndrome. *Clin Epidemiol* 2013;6:1–13.
- Sagvekar P, Dadachanji R, Patil K, et al. Pathomechanisms of polycystic ovary syndrome: multidimensional approaches. *Front Biosci* 2018;10:384–422.
- Marshall JC, Dunaif A. Should all women with PCOS be treated for insulin resistance? *Fertil Steril* 2012;97:18–22.
- Navarro A, Boveris A. The mitochondrial energy transduction system and the aging process. *Am J Physiol Cell Physiol* 2007;292:C670–86.
- Reddy TV, Govatati S, Deenadayal M, et al. Impact of mitochondrial DNA copy number and displacement loop alterations on polycystic ovary syndrome risk in South Indian women. *Mitochondrion* 2019;44:35–40.
- Taanman JW. The mitochondrial genome: structure, transcription, translation and replication. *Biochim Biophys Acta* 1999;1410:103–23.
- Bolden A, Noy GP, Weissbach A. DNA polymerase of mitochondria is a gamma-polymerase. *J Biol Chem* 1977;252:3351–6.
- Wolstenholme DR. Animal mitochondrial DNA: structure and evolution. *Int Rev Cytol* 1992;141:173–216.
- Page MJ, McKenzie JE, Bossuyt PM, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *BMJ* 2021;372:n71.
- PRISMA. Preferred reporting items for systematic reviews and meta-analyses, 2021. Available: <http://www.prisma-statement.org/> [Accessed 7 Jul 2021].

- 12 Sohani ZN, Meyre D, de Souza RJ, *et al.* Assessing the quality of published genetic association studies in meta-analyses: the quality of genetic studies (Q-Genie) tool. *BMC Genet* 2015;16:50.
- 13 Q-Genie. Quality Assessment-of-Genetic-Association-Studies, 2015. Available: <https://fhs.mcmaster.ca/pgp/documents/Q-Geniev.1.1.pdf> [Accessed 7 Jul 2021].
- 14 Rabøl R, Svendsen PF, Skovbro M, *et al.* Skeletal muscle mitochondrial function in polycystic ovarian syndrome. *Eur J Endocrinol* 2011;165:631–7.
- 15 Yang P-K, Chou C-H, Chang C-H, *et al.* Changes in peripheral mitochondrial DNA copy number in metformin-treated women with polycystic ovary syndrome: a longitudinal study. *Reprod Biol Endocrinol* 2020;18:69.
- 16 ZP H, Wang Y, Mao WW. Single nucleotide polymorphisms (SNPs) and variable number tandem repeats (VNTRs) in mtDNA D-loop and CO II- tRNALys intergenic region with PCOS. *Journal of Reproduction and Contraception* 2011;22:129–38.
- 17 Zhuo G, Ding Y, Feng G, *et al.* Analysis of mitochondrial DNA sequence variants in patients with polycystic ovary syndrome. *Arch Gynecol Obstet* 2012;286:653–9.
- 18 Zhuo G, Feng G, Leng J, *et al.* A 9-bp deletion homoplasmism in women with polycystic ovary syndrome revealed by mitochondrial genome-mutation screen. *Biochem Genet* 2010;48:157–63.
- 19 Ding Y, Xia B-H, Zhang C-J, *et al.* Mitochondrial tRNA^{Leu(UUR)} C3275T, tRNA^{Gln} T4363C and tRNA^{Lys} A8343G mutations may be associated with PCOS and metabolic syndrome. *Gene* 2018;642:299–306.
- 20 Ding Y, Xia B-H, Zhang C-J, *et al.* Mutations in mitochondrial tRNA genes may be related to insulin resistance in women with polycystic ovary syndrome. *Am J Transl Res* 2017;9:2984–96.
- 21 Ding Y, Zhuo G, Zhang C, *et al.* Point mutation in mitochondrial tRNA gene is associated with polycystic ovary syndrome and insulin resistance. *Mol Med Rep* 2016;13:3169–72.
- 22 Saeed NAAAH, Hamzah IH, Al-Gharrawi SAR. Polycystic ovary syndrome dependency on mtDNA mutation; copy number and its association with insulin resistance. *BMC Res Notes* 2019;12:455.
- 23 Ding Y, Zhuo G, Zhang C. The Mitochondrial tRNA^{Leu(UUR)} A3302G Mutation may be Associated With Insulin Resistance in Woman With Polycystic Ovary Syndrome. *Reprod Sci* 2016;23:228–33.
- 24 Shukla P, Mukherjee S, Patil A. Identification of variants in mitochondrial D-loop and oriL region and analysis of mitochondrial DNA copy number in women with polycystic ovary syndrome. *DNA Cell Biol* 2020;39:1458–66.
- 25 Wang J, Wu X. The effects of mitochondrial dysfunction on energy metabolism switch by HIF-1 α signalling in granulosa cells of polycystic ovary syndrome. *Endokrynol Pol* 2020;71:134–45.
- 26 Schlötterer C, Tautz D. Slippage synthesis of simple sequence DNA. *Nucleic Acids Res* 1992;20:211–5.
- 27 Huoponen K, Lamminen T, Juvonen V, *et al.* The spectrum of mitochondrial DNA mutations in families with Leber hereditary optic neuropathy. *Hum Genet* 1993;92:379–84.
- 28 Cai W, Fu Q, Zhou X, *et al.* Mitochondrial variants may influence the phenotypic manifestation of Leber's hereditary optic neuropathy-associated ND4 G11778A mutation. *J Genet Genomics* 2008;35:649–55.
- 29 Liu X-L, Zhou X, Zhou J, *et al.* Leber's hereditary optic neuropathy is associated with the T12338C mutation in mitochondrial ND5 gene in six Han Chinese families. *Ophthalmology* 2011;118:978–85.
- 30 Lu J, Qian Y, Li Z, *et al.* Mitochondrial haplotypes may modulate the phenotypic manifestation of the deafness-associated 12S rRNA 1555A>G mutation. *Mitochondrion* 2010;10:69–81.
- 31 Suzuki T, Nagao A, Suzuki T. Human mitochondrial tRNAs: biogenesis, function, structural aspects, and diseases. *Annu Rev Genet* 2011;45:299–329.
- 32 Ding Y, Xia B, Yu J, *et al.* Mitochondrial DNA mutations and essential hypertension (review). *Int J Mol Med* 2013;32:768–74.
- 33 Kelley SO, Steinberg SV, Schimmel P. Functional defects of pathogenic human mitochondrial tRNAs related to structural fragility. *Nat Struct Biol* 2000;7:862–5.
- 34 Ashraf SS, Sochacka E, Cain R, *et al.* Single atom modification (O \rightarrow S) of tRNA confers ribosome binding. *RNA* 1999;5:188–94.
- 35 Deng X, Ji D, Li X, *et al.* Polymorphisms and haplotype of mitochondrial DNA D-loop region are associated with polycystic ovary syndrome in a Chinese population. *Mitochondrion* 2021;57:173–81.
- 36 Shukla P, Mukherjee S. Mitochondrial dysfunction: an emerging link in the pathophysiology of polycystic ovary syndrome. *Mitochondrion* 2020;52:24–39.
- 37 Govatati S, Deenadayal M, Shivaji S, *et al.* Mitochondrial displacement loop alterations are associated with endometriosis. *Fertil Steril* 2013;99:1980–6.
- 38 Yasukawa T, Kang D. An overview of mammalian mitochondrial DNA replication mechanisms. *J Biochem* 2018;164:183–93.
- 39 Ilie IR. Chapter Four - Advances in PCOS Pathogenesis and Progression—Mitochondrial Mutations and Dysfunction. In: Makowski GS, ed. *Advances in clinical chemistry*. Elsevier, 2018: 127–55.
- 40 Mambo E, Gao X, Cohen Y, *et al.* Electrophile and oxidant damage of mitochondrial DNA leading to rapid evolution of homoplasmic mutations. *Proc Natl Acad Sci U S A* 2003;100:1838–43.
- 41 Lee J, Sharma S, Kim J, *et al.* Mitochondrial nuclear receptors and transcription factors: who's minding the cell? *J Neurosci Res* 2008;86:961–71.
- 42 Li H, Slone J, Fei L, *et al.* Mitochondrial DNA variants and common diseases: a mathematical model for the diversity of age-related mtDNA mutations. *Cells* 2019;8:608.
- 43 Fan Y-J, Zong W-X. The cellular decision between apoptosis and autophagy. *Chin J Cancer* 2013;32:121–9.
- 44 Jeng J-Y, Yeh T-S, Lee J-W, *et al.* Maintenance of mitochondrial DNA copy number and expression are essential for preservation of mitochondrial function and cell growth. *J Cell Biochem* 2008;103:347–57.
- 45 Zandalinas SI, Mittler R. ROS-Induced ROS release in plant and animal cells. *Free Radic Biol Med* 2018;122:21–7.
- 46 Lee S-H, Chung D-J, Lee H-S, *et al.* Mitochondrial DNA copy number in peripheral blood in polycystic ovary syndrome. *Metabolism* 2011;60:1677–82.
- 47 Mishra JS, More AS, Kumar S. Elevated androgen levels induce hyperinsulinemia through increase in INS1 transcription in pancreatic beta cells in female rats. *Biol Reprod* 2018;98:520–31.
- 48 Wang H, Wang X, Zhu Y, *et al.* Increased androgen levels in rats impair glucose-stimulated insulin secretion through disruption of pancreatic beta cell mitochondrial function. *J Steroid Biochem Mol Biol* 2015;154:254–66.
- 49 Jia L, Li J, He B, *et al.* Abnormally activated one-carbon metabolic pathway is associated with mtDNA hypermethylation and mitochondrial malfunction in the oocytes of polycystic GILT ovaries. *Sci Rep* 2016;6:19436.
- 50 Lumenlearning. Oxidative phosphorylation: Boundless biology, 2021. Available: <https://courses.lumenlearning.com/boundless-biology/chapter/oxidative-phosphorylation/> [Accessed 8 Jul 2021].
- 51 Learn CBSE. NCERT exemplar class 11 biology solutions respiration electron transport chain, 2018. Available: <https://www.learncbse.in/ncert-exemplar-class-11-biology-solutions-respiration-in-plants/> [Accessed 8 Jul 2021].
- 52 Hamann J. Oxidative stress: impact on dairy health and immune function, 2015. Available: <https://en.engormix.com/dairy-cattle/articles/oxidative-stress-impact-dairy-t36491.htm> [Accessed 12 Jul 2021].
- 53 Knapp S. Oxidative Stress - The Definitive Guide, 2020. Available: <https://biologydictionary.net/oxidative-stress/> [Accessed 8 Jul 2021].
- 54 Loeb LA, Wallace DC, Martin GM. The mitochondrial theory of aging and its relationship to reactive oxygen species damage and somatic mtDNA mutations. *Proc Natl Acad Sci U S A* 2005;102:18769.
- 55 Rotterdam ESHRE/ASRM-Sponsored PCOS consensus workshop group. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome (PCOS). *Hum Reprod* 2004;19:41–7.
- 56 NHS. Polycystic ovary syndrome - Treatment, 2019. Available: <https://www.nhs.uk/conditions/polycystic-ovary-syndrome-pcos/treatment/> [Accessed 8 Jul 2021].

SUPPLEMENT 1 – SEARCH TERMS

Search terms relating to “Mitochondrial Genetics”

Association analysis	Haplotype
Candidate gene	Heavy strand
Circular DNA	HVR-1
Coding region	HVR-2
Copy number	Hypervariable
Copy number variation	Hypervariable region
D-loop	Hypervariable region-1
Deep sequencing	Hypervariable region-2
Deletion	Insertion
Displacement loop	Light strand
DNA	Messenger RNA
DNA polymerase-gamma	Microsatellite repeat
DNA sequence	Mitochondrial DNA
Electron transport chain	Mitochondrial genome
ETC	Mitochondrial-encoded
Gen* linkage	Mitoribosome
Gen* mapping	mtDNA
Gen* mutation	Multifactorial inheritance
Gene expression	Mutation
Gene expression profiling	Next generation sequencing
Gene expression regulation	NGS
Gene frequency	Non-coding region
Gene* profile	Nucleotide sequencing
Gene regulatory network	Organelle
Gene sequencing	Oxidative stress
Gene* variant	PCR
Genes	Phenotype
Genetic analysis	Polygenic
Genetic association	Polymerase chain reaction
Genetic association study	Polymorphism
Genetic background	Promoter region
Genetic defect	Reactive oxygen species
Genetic epidemiology	Respiratory chain
Genetic factor	Respiratory chain complex
Genetic influence	ROS
Genetic linkage	Single nucleotide polymorphism
Genetic marker	SNP
Genetic polymorphism	Southern blot
Genetic predisposition to disease	Tandem repeat
Genetic stud*	Transcript*
Genetic susceptibility	Transcription factor
Genetic techniqu*	Translation
Genetic test*	Trinucleotide repeat
Genetic variation	Western blot
Genetic*	Whole exome sequencing
Genome	
Genome analysis	
Genome-wide	
Genome-wide association	
Genomics	

Search terms relating to PCOS

Multicystic ovary
Polycystic ovary
PCO
PCOS
Polycystic disease of the ovary
Polycystic Ovarian Disease
Polycystic Ovarian Syndrome
Polycystic ovaries
Polycystic ovary disease
Polycystic ovary morphology
Polycystic Ovary Syndrome
POS
Stein-Leventhal Syndrome
Stein Leventhal Syndrome

SUPPLEMENT 2 – STUDIES INCLUDED IN SYSTEMATIC REVIEW

Table showing studies included in systematic review

	Study ID	Title of Publication	Location	Patient characteristics	Available at
INCLUDED IN META-ANALYSIS	Hu et al. (2011)[14]	Single Nucleotide Polymorphisms (SNPs) And Variable Number Tandem Repeats (VNTRs) In MtDNA D-Loop and CO II- tRNA Lys Intergenic Region With PCOS	China	Women aged 19-37 diagnosed with PCOS according to the revised 2003 Rotterdam Criteria and control group matched for age	http://dx.doi.org/10.1016/S1001-7844%2812%2960008-X
	Zhuo et al. (2012)[15]	Analysis of Mitochondrial DNA Sequence Variants in Patients with Polycystic Ovary Syndrome	China	Women 22-34 diagnosed with PCOS according to the revised 2003 Rotterdam Criteria and control group matched for age and BMI	https://dx.doi.org/10.1007/s00404-012-2358-7
	Ding et al. (2017)[18]	Mutations in Mitochondrial tRNA Genes May Be Related to Insulin Resistance in Women with Polycystic Ovary Syndrome	China	Women aged 18-37 diagnosed with insulin-resistant subtype PCOS according to the revised 2003 Rotterdam Criteria and control group matched for age and BMI	http://www.ajtr.org/files/ajtr0048638.pdf
	Reddy et al. (2019)[6]	Impact of Mitochondrial DNA Copy Number and Displacement Loop Alterations on Polycystic Ovary Syndrome Risk in South Indian Women	India	Women aged 22-36 diagnosed with PCOS according to the revised 2003 Rotterdam Criteria and control group matched for age and BMI	https://dx.doi.org/10.1016/j.mito.2017.12.010
	Saeed et al. (2019)[20]	Polycystic Ovary Syndrome Dependency on MtDNA Mutation; Copy Number and Its Association with Insulin Resistance	Iraq	Women aged 20-30 diagnosed with both diabetes and PCOS according to the revised 2003 Rotterdam Criteria and diabetic control group matched for age	https://dx.doi.org/10.1186/s13104-019-4453-3
	Shukla et al. (2020)[22]	Identification of Variants in Mitochondrial D-Loop and OriL Region and Analysis of Mitochondrial DNA Copy Number	India	Women aged 15-40 diagnosed with PCOS according to the	https://dx.doi.org/10.1

		in Women with Polycystic Ovary Syndrome		revised 2003 Rotterdam Criteria and control group matched for age	089/dna.2019.5323
	Wang et al. (2020)[23]	The Effects of Mitochondrial Dysfunction on Energy Metabolism Switch By HIF-1 α Signaling in Granulosa Cells of Polycystic Ovary Syndrome	China	Women aged 21-33 diagnosed with PCOS according to the revised 2003 Rotterdam Criteria and control group matched for age	http://dx.doi.org/10.5603/EP.a2020.0002
	Deng et al. (2021)[33]	Polymorphisms and Haplotype of Mitochondrial DNA D-Loop Region Are Associated with Polycystic Ovary Syndrome in A Chinese Population	China	Women aged 24-31 diagnosed with PCOS according to the revised 2003 Rotterdam Criteria and control group matched for age	http://dx.doi.org/10.1016/j.mito.2020.12.006
INSUFFICIENT DATA FOR META-ANALYSIS	Zhuo et al. (2010)[16]	A 9-Bp Deletion Homoplasmy In Women with Polycystic Ovary Syndrome Revealed by Mitochondrial Genome-Mutation Screen	China	Women diagnosed with PCOS according to the revised 2003 Rotterdam Criteria and control group matched for age and BMI	https://dx.doi.org/10.1007/s10528-009-9308-5
	Lee et al. (2011)[43]	Mitochondrial DNA Copy Number in Peripheral Blood in Polycystic Ovary Syndrome	Korea	Women aged 24-33 diagnosed with PCOS according to the revised 2003 Rotterdam Criteria and control group matched for age and BMI	https://dx.doi.org/10.1016/j.metabol.2011.04.010
	Rabol et al. (2011)[49]	Skeletal Muscle Mitochondrial Function in Polycystic Ovarian Syndrome	Denmark	Lean and obese women aged 26-33 diagnosed with PCOS according to the revised 2003 Rotterdam Criteria and lean and obese control groups matched for age and BMI	https://dx.doi.org/10.1530/EJE-11-0419
	Ding et al. (2016a)[21]	The Mitochondrial tRNA ^{Leu} (UUR) A3302G Mutation May Be Associated with Insulin Resistance in Women with Polycystic Ovary Syndrome	China	First and second generation family members of a Han	https://dx.doi.org/10.1177/19337191156027

				Chinese family with maternally inherited insulin resistance	77
Ding et al. (2016b)[19]	Point Mutation in Mitochondrial tRNA Gene Is Associated with Polycystic Ovary Syndrome and Insulin Resistance	China		A female patient (age, 31 years) diagnosed with PCOS according to the revised 2003 Rotterdam Criteria and control group matched for age	https://dx.doi.org/10.3892/mmr.2016.4916
Ding et al. (2018)[17]	Mitochondrial TRN ^{Leu} (UUR) C3275T, TRN ^{AGln} T4363C And TRN ^{Lys} A8343G Mutations May Be Associated with PCOS And Metabolic Syndrome	China		Three-generation pedigree with maternally transmitted Metabolic syndrome, including a case of PCOS diagnosed according to the revised 2003 Rotterdam Criteria and control group matched for age and BMI	https://dx.doi.org/10.1016/j.gene.2017.11.049
Yang et al. (2020)[50]	Changes in Peripheral Mitochondrial DNA Copy Number in Women with Polycystic Ovary Syndrome: A Longitudinal Study	Taiwan		Women aged 20-28 diagnosed with PCOS according to the revised 2003 Rotterdam Criteria treated with Metformin for 1 year	http://dx.doi.org/10.1186/s12958-020-00629-5

SUPPLEMENT 3 – POLYMORPHISMS IN CODING REGIONS OF THE MITOCHONDRIAL GENOME

Study ID	Gene	Position
Zhuo 2010	A6	8584
Zhuo 2010	A6	8701
Zhuo 2010	A6	8745
Zhuo 2010	A6	8794
Zhuo 2010	A6	9053
Zhuo 2010	A6	9101
Zhuo 2010	A6	9128
Zhuo 2010	A6	9180
Zhuo 2012	A6	8584
Zhuo 2012	A6	8684
Zhuo 2012	A6	8745
Zhuo 2012	A6	8793
Zhuo 2012	A6	8860
Zhuo 2012	A6	9053
Zhuo 2012	A6	9126
Zhuo 2012	A6	9128
Ding 2016	A6	8585
Ding 2016	A6	8860
Ding 2017	A6	8584
Ding 2017	A6	8684
Zhuo 2010	A8	8392
Zhuo 2010	A8	8414
Zhuo 2010	A8	8459
Zhuo 2010	A8	8473
Zhuo 2012	A8	8392
Zhuo 2012	A8	8414
Zhuo 2012	A8	8459
Zhuo 2012	A8	8473
Ding 2016	A8	8414
Zhuo 2010	COX1	6392
Zhuo 2010	COX1	6962
Ding 2016	COX1	6392
Ding 2016	COX1	7028
Ding 2018	COX1	7028
Zhuo 2012	COX2	6221
Zhuo 2012	COX2	6272
Zhuo 2012	COX2	6338
Zhuo 2012	COX2	6392
Zhuo 2012	COX2	6455
Zhuo 2012	COX2	6962
Zhuo 2012	COX2	7028
Zhuo 2012	COX2	7142
Zhuo 2010	COX2	7853
Ding 2016	COX2	8020

Study ID	Gene	Position
Zhuo 2010	COX3	9536
Zhuo 2010	COX3	9540
Zhuo 2010	COX3	9548
Zhuo 2010	COX3	9824
Zhuo 2012	COX3	9242
Zhuo 2012	COX3	9336
Zhuo 2012	COX3	9377
Zhuo 2012	COX3	9540
Zhuo 2012	COX3	9559
Zhuo 2012	COX3	9824
Ding 2016	COX3	9540
Ding 2016	COX3	9824
Zhuo 2010	CYTB	14783
Zhuo 2010	CYTB	15038
Zhuo 2010	CYTB	15043
Zhuo 2010	CYTB	15244
Zhuo 2010	CYTB	15346
Zhuo 2010	CYTB	15301
Zhuo 2012	CYTB	14783
Zhuo 2012	CYTB	15043
Zhuo 2012	CYTB	15244
Zhuo 2012	CYTB	15301
Zhuo 2012	CYTB	15326
Zhuo 2012	CYTB	15460
Zhuo 2012	CYTB	15724
Ding 2016	CYTB	14766
Ding 2016	CYTB	15301
Ding 2016	CYTB	15326
Ding 2018	CYTB	14766
Ding 2018	CYTB	15326
Ding 2018	CYTB	15535
Zhuo 2010	ND1	3316
Zhuo 2010	ND1	3394
Zhuo 2010	ND1	3497
Zhuo 2010	ND1	3970
Zhuo 2010	ND1	4086
Zhuo 2010	ND1	4248
Zhuo 2012	ND1	3316
Zhuo 2012	ND1	3394
Zhuo 2012	ND1	3423
Zhuo 2012	ND1	3497
Zhuo 2012	ND1	3552
Zhuo 2012	ND1	3834
Zhuo 2012	ND1	3970

Study ID	Gene	Position
Zhuo 2012	ND1	4086
Ding 2016	ND1	3553
Ding 2016	ND1	4047
Ding 2017	ND1	3394
Zhuo 2010	ND2	5147
Zhuo 2010	ND2	5178
Zhuo 2010	ND2	5301
Zhuo 2010	ND2	5417
Zhuo 2010	ND2	5460
Zhuo 2012	ND2	4688
Zhuo 2012	ND2	4769
Zhuo 2012	ND2	4883
Zhuo 2012	ND2	5046
Zhuo 2012	ND2	5108
Zhuo 2012	ND2	5147
Zhuo 2012	ND2	5178
Zhuo 2012	ND2	5231
Zhuo 2012	ND2	5263
Zhuo 2012	ND2	5301
Zhuo 2012	ND2	5417
Zhuo 2012	ND2	5460
Ding 2018	ND2	4715
Ding 2018	ND2	4820
Zhuo 2010	ND3	10310
Zhuo 2010	ND3	10397
Zhuo 2010	ND3	10398
Zhuo 2010	ND3	10400
Zhuo 2012	ND3	10097
Zhuo 2012	ND3	10238
Zhuo 2012	ND3	10310
Zhuo 2012	ND3	10398
Zhuo 2012	ND3	10400
Ding 2016	ND3	10398
Ding 2016	ND3	10400
Zhuo 2012	ND4	10873
Zhuo 2012	ND4	10915
Zhuo 2012	ND4	11084
Zhuo 2012	ND4	11335
Zhuo 2012	ND4	11440
Zhuo 2012	ND4	11719
Zhuo 2012	ND4	11944
Zhuo 2012	ND4	12026

Study ID	Gene	Position
Zhuo 2010	ND4	10609
Zhuo 2010	ND4	10873
Zhuo 2010	ND4	11084
Zhuo 2010	ND4	11914
Zhuo 2010	ND4	11944
Zhuo 2010	ND4	12026
Ding 2016	ND4	10873
Ding 2016	ND4	11719
Ding 2018	ND4	11719
Zhuo 2010	ND5	12406
Zhuo 2010	ND5	12705
Zhuo 2010	ND5	12882
Zhuo 2010	ND5	13759
Zhuo 2010	ND5	13824
Zhuo 2010	ND5	13928
Zhuo 2010	ND5	13942
Zhuo 2010	ND5	14067
Zhuo 2012	ND5	12406
Zhuo 2012	ND5	12705
Zhuo 2012	ND5	12811
Zhuo 2012	ND5	13145
Zhuo 2012	ND5	13263
Zhuo 2012	ND5	13434
Zhuo 2012	ND5	13708
Zhuo 2012	ND5	13759
Zhuo 2012	ND5	13928
Ding 2016	ND5	12705
Ding 2016	ND5	13928
Ding 2018	ND5	13928
Ding 2017	ND5	12811
Ding 2016	ND5	12338
Zhuo 2010	ND6	14668
Zhuo 2012	ND6	14199
Zhuo 2012	ND6	14272
Zhuo 2012	ND6	14365
Zhuo 2012	ND6	14569
Zhuo 2012	ND6	14668
Ding 2018	ND6	14470

SUPPLEMENT 4 - POLYMORPHISMS IN GENES CODING FOR MITOCHONDRIAL tRNAs**All polymorphisms detected in genes coding for mt-tRNAs**

Study ID	Gene	Position	PCOS sample size	PCOS with SNP	PCOS without SNP	Frequency in PCOS (%)	Control sample size	Controls with SNP	Controls without SNP	Frequency in Controls (%)
Zhuo 2012	tRNA Arg	10454	57	1	56	1.7	38	0	38	0
Ding 2017	tRNA Arg	10454	80	2	78	2.5	50	0	50	0
Zhuo 2012	tRNA Asp	7543	57	1	56	1.7	38	0	38	0
Ding 2017	tRNA Asp	7543	80	1	79	1.25	50	0	50	0
Zhuo 2012	tRNA Cys	5821	57	1	56	1.7	38	0	38	0
Ding 2017	tRNA Cys	5821	80	3	77	3.75	50	1	49	2
Zhuo 2010	tRNA Gln	14693	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Ding 2018	tRNA Gln	4363	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Ding 2017	tRNA Gln	4363	80	4	76	5	50	0	50	0
Ding 2017	tRNA Gln	4395	80	1	79	1.25	50	0	50	0
Zhuo 2012	tRNA Glu	4395	57	17	40	29.8	38	0	38	0
Ding 2017	tRNA Glu	14693	80	1	79	1.25	50	0	50	0
Ding 2017	tRNA Met	4454	80	0	80	0	50	2	48	4
Ding 2016	tRNA Leu	3302	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Ding 2018	tRNA Leu	3275	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Ding 2017	tRNA Leu	3302	80	1	79	1.25	50	0	50	0
Ding 2017	tRNA Leu	3275	80	2	78	2.5	50	0	50	0
Saeed 2019	tRNA Leu	3157	70	2	68	2.86	59	0	59	0
Saeed 2019	tRNA Leu	3162	70	0	70	0	59	0	59	0
Saeed 2019	tRNA Leu	3203	70	3	67	4.29	59	1	58	1.69
Saeed 2019	tRNA Leu	3282	70	2	68	2.86	59	0	59	0
Saeed 2019	tRNA Leu	3285	70	2	68	2.86	59	0	59	0
Saeed 2019	tRNA Leu	3302	70	2	68	2.86	59	0	59	0
Saeed 2019	tRNA Leu	3275	70	4	66	5.71	59	0	59	0

Saeed 2019	tRNA Leu	4225	70	1	69	1.43	59	0	59	0
Saeed 2019	tRNA Leu	5206	70	3	67	4.29	59	0	59	0
Saeed 2019	tRNA Leu	8434	70	1	69	1.43	59	3	56	5.08
Zhuo 2012	tRNA Lys	8343	57	1	56	1.7	38	0	38	0
Ding 2018	tRNA Lys	8343	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Ding 2017	tRNA Lys	8343	80	2	78	2.5	50	0	50	0
Ding 2017	tRNA Ser	7492	80	1	79	1.25	50	0	50	0
Ding 2016	tRNA Ser	7492	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Ding 2017	tRNA Thr	15900	80	0	80	0	50	1	49	2

Dataset used in meta-analysis

Mitochondrial-encoded tRNA	Frequency of SNP in PCOS (%)	Frequency of SNP in Control (%)
Cys	2.9	1.1
Leu	2.7	0.6
Glu	13.1	0.0
Gln	3.1	0.0
Lys	2.2	0.0
Arg	1.9	0.0
Asp	1.5	0.0

SUPPLEMENT 5 – POLYMORPHISMS IN GENES CODING FOR MITOCHONDRIAL rRNAs

Study ID	Gene	Position
Zhuo 2010	12S rRNA	663
Zhuo 2010	12S rRNA	709
Zhuo 2010	12S rRNA	752
Zhuo 2010	12S rRNA	827
Zhuo 2010	12S rRNA	1438
Zhuo 2012	12S rRNA	663
Zhuo 2012	12S rRNA	709
Zhuo 2012	12S rRNA	750
Zhuo 2012	12S rRNA	752
Zhuo 2012	12S rRNA	827
Zhuo 2012	12S rRNA	1382
Zhuo 2012	12S rRNA	1438
Ding 2016	12S rRNA	750
Ding 2016	12S rRNA	1438
Ding 2018	12S rRNA	750
Ding 2018	12S rRNA	827
Ding 2018	12S rRNA	1438
Zhuo 2010	16S rRNA	1736
Zhuo 2010	16S rRNA	3010
Zhuo 2012	16S rRNA	1736
Zhuo 2012	16S rRNA	2706
Zhuo 2012	16S rRNA	3010
Ding 2016	16S rRNA	1734
Ding 2016	16S rRNA	2706
Ding 2018	16S rRNA	2706
Ding 2018	16S rRNA	3109

SUPPLEMENT 6 – POLYMORPHISMS IN THE D-LOOP OF THE MITOCHONDRIAL GENOME**All polymorphisms detected in the D-Loop of the mitochondrial genome**

Study ID	Polymorphism	PCOS sample size	PCOS with SNP	PCOS without SNP	Frequency in PCOS (%)	Control sample size	Control with SNP	Control without SNP	Frequency in controls (%)
Zhuo 2012	A73G	57	10	47	17.5	38	8	30	21.6
Zhuo 2012	T146C	57	1	56	1.7	38	1	37	2.7
Zhuo 2012	C150T	57	6	51	10.5	38	3	35	8.1
Zhuo 2012	A153G	57	4	53	7.0	38	2	36	5.4
Zhuo 2012	T195A	57	5	52	8.7	38	2	36	5.4
Zhuo 2012	A235G	57	3	54	5.3	38	2	36	5.4
Zhuo 2012	A263G	57	8	49	14.0	38	7	31	18.9
Zhuo 2012	G316A	57	1	56	1.7	38	0	38	0
Zhuo 2012	T489A	57	4	53	7.0	38	3	35	8.1
Zhuo 2012	G16129A	57	2	55	3.5	38	2	36	5.4
Zhuo 2012	T16140C	57	3	54	5.3	38	0	38	0
Zhuo 2012	T16189C	57	6	51	10.5	38	4	34	10.8
Zhuo 2012	C16223T	57	9	48	15.8	38	2	36	5.4
Zhuo 2012	A16316G	57	6	51	10.5	38	5	33	13.5
Zhuo 2012	T16357C	57	1	56	1.7	38	0	38	0
Zhuo 2012	T16519C	57	2	55	3.5	38	1	37	2.7
Reddy 2019	A73G	118	75	43	63.6	114	75	39	65.8
Reddy 2019	A93G	118	7	111	5.9	114	4	110	3.5
Reddy 2019	T146C	118	8	110	6.8	114	12	102	10.5
Reddy 2019	T152C	118	23	95	19.5	114	23	91	20.2
Reddy 2019	A189G	118	31	87	26.3	114	15	99	13.2
Reddy 2019	T195A	118	8	110	6.8	114	7	107	6.1
Reddy 2019	A263G	118	72	46	61.0	114	76	38	66.7
Reddy 2019	316C	118	71	47	60.1	114	68	46	59.6
Reddy 2019	T489C	118	25	93	21.2	114	14	100	12.3
Reddy 2019	522CA	118	15	103	12.7	114	7	107	6.1

Reddy 2019	G16129A	118	9	109	7.6	114	11	103	9.6
Reddy 2019	T16172C	118	10	108	8.5	114	11	103	9.6
Reddy 2019	T16189C	118	14	104	11.9	114	15	99	13.2
Reddy 2019	C16223T	118	44	74	37.3	114	52	62	45.6
Reddy 2019	T16311C	118	13	105	11.0	114	16	98	14.0
Reddy 2019	G16319A	118	12	106	10.2	114	5	109	4.4
Reddy 2019	T16362C	118	14	104	11.9	114	8	106	7.0
Reddy 2019	T16519C	118	63	55	53.4	114	54	60	47.4
Reddy 2019	D310	118	64	54	54.2	114	33	81	28.9
Deng 2021	C150T	421	79	342	18.76	409	103	306	25.18
Deng 2021	G207A	421	11	410	2.61	409	28	381	6.85
Deng 2021	A263G	421	418	3	99.29	409	399	10	97.56
Deng 2021	16036G	421	24	397	5.70	409	42	367	10.27
Deng 2021	16036GG	421	2	419	0.48	409	22	387	5.38
Deng 2021	16049G	421	2	419	0.48	409	36	373	8.80
Deng 2021	C16234T	421	17	404	4.04	409	30	379	7.33
Deng 2021	T16362C	421	169	252	40.14	409	196	213	47.92
Hu 2011	16094T/C	77	7	70	9.09	45	4	41	8.89
Hu 2011	C16173T	77	67	10	87.01	45	38	7	84.40
Hu 2011	16181 – 16195	77	76	1	98.70	45	44	1	97.78
Hu 2011	A2C12	77	6	71	7.89	45	1	44	2.27
Hu 2011	A3C11	77	9	68	11.84	45	8	37	18.18
Hu 2011	A4C9T	77	48	29	63.16	45	31	14	70.45
Hu 2011	A4C8T2	77	13	64	17.11	45	4	41	9.09
Hu 2011	T16225C	77	24	53	31.17	45	14	31	31.11
Hu 2011	T16300C	77	9	68	11.69	45	8	37	17.78
Hu 2011	T16306C	77	10	67	12.99	45	4	41	8.89
Hu 2011	T16313C	77	10	67	12.99	45	7	38	15.56
Hu 2011	G16321A	77	9	68	11.69	45	8	37	17.78
Hu 2011	T16364C	77	28	49	36.36	45	18	27	40.00

Hu 2011	T146C	77	9	68	11.69	45	5	40	11.11
Hu 2011	C150T	77	62	15	80.52	45	39	6	80.00
Hu 2011	T152C	77	19	58	24.68	45	9	36	20.00
Hu 2011	C195T	77	66	11	85.71	45	42	3	93.33
Hu 2011	A248	77	17	60	22.08	45	9	36	20.00
Hu 2011	T491C	77	48	29	62.34	45	22	23	48.89
Hu 2011	303-317	77	76	1	98.70	45	44	1	97.78
Hu 2011	C8TC6	77	41	36	53.95	45	18	27	40.91
Hu 2011	C9TC6	77	11	66	14.47	45	5	40	11.36
Hu 2011	C7TC6	77	24	53	31.58	45	21	24	47.73
Zhuo 2010	T146C	-	-	-	-	-	-	-	-
Zhuo 2010	T150C	-	-	-	-	-	-	-	-
Zhuo 2010	T152C	-	-	-	-	-	-	-	-
Zhuo 2010	C195T	-	-	-	-	-	-	-	-
Zhuo 2010	A200G	-	-	-	-	-	-	-	-
Zhuo 2010	G207A	-	-	-	-	-	-	-	-
Zhuo 2010	A235G	-	-	-	-	-	-	-	-
Zhuo 2010	A263G	-	-	-	-	-	-	-	-
Zhuo 2010	T489C	-	-	-	-	-	-	-	-
Zhuo 2010	G16129A	-	-	-	-	-	-	-	-
Zhuo 2010	T16172C	-	-	-	-	-	-	-	-
Zhuo 2010	T16189C	-	-	-	-	-	-	-	-
Zhuo 2010	C16223T	-	-	-	-	-	-	-	-
Zhuo 2010	C16290T	-	-	-	-	-	-	-	-
Zhuo 2010	G16319A	-	-	-	-	-	-	-	-
Zhuo 2010	T16362C	-	-	-	-	-	-	-	-
Zhuo 2010	T16519C	-	-	-	-	-	-	-	-
Ding 2016	A73G	-	-	-	-	-	-	-	-
Ding 2016	T152C	-	-	-	-	-	-	-	-
Ding 2016	D310	-	-	-	-	-	-	-	-

Ding 2016	T16189C	-	-	-	-	-	-	-	-
Ding 2016	T16519C	-	-	-	-	-	-	-	-
Ding 2018	A73G	-	-	-	-	-	-	-	-
Ding 2018	C150T	-	-	-	-	-	-	-	-
Ding 2018	C328T	-	-	-	-	-	-	-	-
Ding 2018	T16142C	-	-	-	-	-	-	-	-
Ding 2018	T16189C	-	-	-	-	-	-	-	-
Ding 2018	T16224C	-	-	-	-	-	-	-	-

Dataset used in meta-analysis

Polymorphism	Study ID	PCOS sample size	PCOS with SNP	PCOS without SNP	Frequency in PCOS (%)	Control sample size	Control with SNP	Control without SNP	Frequency in controls (%)
C150T	Hu 2011	77	62	15	80.52	45	39	6	80.00
	Zhuo 2012	57	6	51	10.5	38	3	35	8.1
	Deng 2021	421	79	342	18.76	409	103	306	25.18
T146C	Hu 2011	77	9	68	11.69	45	5	40	11.11
	Zhuo 2012	57	1	56	1.7	38	1	37	2.7
	Reddy 2019	118	8	110	6.8	114	12	102	10.5
A263G	Zhuo 2012	57	8	49	14.0	38	7	31	18.9
	Reddy 2019	118	72	46	61.0	114	76	38	66.7
	Deng 2021	421	418	3	99.29	409	399	10	97.56

SUPPLEMENT 7 – MtDNA COPY NUMBERS

	Study ID	Cells	Total Sample Size (n)	PCOS Sample Size	Control Sample Size	PCOS Log (mean mt CN)	Control Log (mean mt CN)	PCOS SEM	Control SEM	PCOS SD	Control SD
Included in meta-analysis	Ding et al. 2017	Blood	130	80	50	0.29	0.84	0.31	0.88	0.44	0.79
	Reddy et al. 2019	Blood	232	118	114	1.36	1.50	0.44	0.56	4.78	5.98
	Shukla et al. 2020	Blood	60	30	30	1.22	1.64	0.33	0.35	1.81	1.92
	Wang et al. 2020	Granulosa	107	39	68	0.761	1.021	0.05	0.06	0.31	0.49
Insufficient data for meta-analysis	Lee et al. 2011	Blood	110	50	60	1.44	1.88	-	-	-	-
	Ding et al. 2016	Blood	-	-	-	-	-	-	-	-	-
	Ding et al. 2018	Blood	-	-	-	-	-	-	-	-	-
	Saeed et al. 2019	Blood	63	38	25	0.88	1.00	-	-	-	-
	Yang et al. 2020	Blood	88	0	-	-	-	-	-	-	-

SUPPLEMENT 8 – QUALITY ASSESSMENT SCORES

Study ID	Rationale for study	Selection and definition of outcome of interest	Selection and comparability of comparison group	Technical classification of the exposure	Non-technical classification of the exposure	Other sources of bias	Sample size and power	A priori planning of analyses	Statistical methods and control for confounding	Testing of assumptions and inferences for genetic analyses	Appropriateness of inferences drawn from results	Total score	Quality status
Zhuo 2010	5	4	4	3	1	1	2	2	1	3	3	29	Poor
Hu 2011	4	3	4	5	3	3	2	5	6	4	3	42	Moderate
Lee 2011	6	5	6	6	5	7	2	5	6	6	6	60	Good
Rabol 2011	7	4	5	3	2	1	1	5	3	4	5	40	Moderate
Zhuo 2012	6	5	5	3	3	2	2	3	5	5	4	43	Moderate
Ding 2016a	2	2	2	2	1	2	1	2	1	3	1	19	Poor
Ding 2016b	1	2	1	1	1	1	1	2	1	1	1	13	Poor
Ding 2017	6	6	6	6	6	2	3	6	4	7	6	58	Good
Ding 2018	3	2	2	3	1	2	1	3	2	4	2	25	Poor
Reddy 2019	7	6	7	7	7	3	3	7	7	6	3	63	Good
Saeed 2019	4	2	4	6	4	3	2	6	5	1	2	39	Moderate
Yang 2020	7	2	-	6	3	6	2	7	5	5	6	49	Good
Shukla 2020	6	3	6	6	3	1	2	5	2	5	5	44	Moderate
Wang 2020	5	4	4	7	3	1	1	4	3	5	5	42	Moderate
Deng 2021	7	3	5	3	3	1	4	7	7	6	4	50	Good
Median	6	3	4	5	3	2	2	5	4	5	4		
Q1	4	2	2	3	1	1	1	3	2	3	2		
Q3	7	5	6	6	4	3	2	6	6	6	5		
IQR	4-7	2-5	2-6	3-6	1-4	1-3	1-2	3-6	2-6	3-6	2-5		