Folate absorption

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Folic acid, or pteroylglutamic acid, is the parent substance of a large group of related compounds called 'folates'. All folate compounds have the same basic molecular structure consisting of three parts: pteridine, para-aminobenzoate (these two together forming a pteroyl group), and L-glutamic acid (Fig. 1). Pteroylglutamic acid itself is a stable, watersoluble compound with a molecular weight 441.4; it forms yellow, spear-shaped crystals. Though this was the form of the vitamin first crystallized from natural materials (Mitchell, Snell, and Williams, 1941), pteroylglutamic acid as such is only a minor component (less than 1%) of natural folates. The folates that do occur in plants and animals and therefore form the bulk of normal dietary folate differs from pteroylglutamic acid in that (1) they are usually reduced to the di- or tetrahydrofolate state at positions 5, 6, 7, and 8 in the pteroyl portion; (2) they usually contain a methyl or formyl group at position 5, or a formyl at position 10 in the pteroyl portion; (3) they usually contain a chain of three or more glutamic acid residues linked to each other by gamma peptide bonds (Fig. 1).

Most studies of folate absorption have been carried out using pteroylglutamic acid because this compound is stable and is commercially available in pure form, both radioactively labelled as tritiated pteroylglutamic acid, and non-labelled. It is likely that the events that take place during absorption of natural folates differ in several respects from those that occur during pteroylglutamic acid absorption. It is impossible, however, to give a clear detailed picture of the absorption process of either form of folate since there are wide disagreements in this field, both in the evidence produced by different groups and in their interpretation. These differences may stem partly from technical difficulties. For instance, many measurements are made with relatively inaccurate microbiological assays. Moreover, most folate compounds are not available pure and are unstable; the only radioactive forms of folates available are labelled with $\beta$ emitters ($^{14}$C or $^{15}$II), which means that studies of faecal excretion are difficult and studies using whole body counting are impossible. Finally, no studies have been carried out using foods containing isotopically labelled folate.

The aim of this paper will be first to summarize what is known about the absorption of pteroylglutamic acid, both in man and in experimental animals, second to describe what is known of the absorption of natural folates in man, and finally, to enumerate those clinical syndromes in which malabsorption of folate is thought to occur. Causes of folate deficiency other than malabsorption and the methods of diagnosis of folate deficiency have been the subjects of a number of recent reviews (Mollin and Waters, 1968; Chanarin, 1969; Hoffbrand and Peters, 1970; Herbert, 1970; Waxman, Corcino, and Herbert, 1970) and are not discussed here.

Absorption of Pteroylglutamic Acid

HUMAN STUDIES

Methods

Microbiological assays with Streptococcus faecalis and Lactobacillus casei are widely used in the study of folate absorption and it is therefore essential to know the growth characteristics of these bacteria on different forms of folate in order to interpret the results. Table I shows the response of these organisms to different folate compounds.

Pteroylglutamic acid absorption was originally studied by measuring urinary excretion of folate microbiologically with S. faecalis after an oral dose of the compound (Denko, Grundy, Wheeler, . . . )

![Fig. 1 Formula for folic acid (pteroylglutamic acid).

Dietary folates may contain: (1) Additional hydrogen atoms at positions 7 and 8 (dihydrofolate) or 5, 6, 7, and 8 (tetrahydrofolate). (2) A formyl group at $N_5$ or $N_{10}$ or a methyl group at $N_6$. (3) Additional glutamate moieties attached to the $\gamma$-carboxyl group of the glutamate moiety.](image-url)
Folate absorption and malabsorption

<table>
<thead>
<tr>
<th></th>
<th><em>L. casei</em></th>
<th><em>S. faecalis</em></th>
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<tbody>
<tr>
<td>Pteroylglutamic acid</td>
<td>+</td>
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<tr>
<td>5-Methyltetrahydropteroylglutamic acid</td>
<td>+</td>
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<tr>
<td>Pteroylglutamic acid</td>
<td>+</td>
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<tr>
<td>Pteroyltiglutamic acid (&gt; 3 glutamate moieties)</td>
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Table I Growth of *L. casei* and *S. faecalis* on different folate compounds

1 + = growth, - = no growth.

Henderson, Berryman, Friedemann, and Youmans, 1946; Swendsen, Bird, Brown, and Bethell, 1947. Girdwood (1953) adapted this technique to distinguish more accurately between conditions of folate malabsorption and conditions of deficiency without malabsorption by comparing excretion of folate after oral and parenteral doses of pteroylglutamic acid. A more reliable method for routine clinical purposes was subsequently developed, in which the rise in serum pteroylglutamic acid level is measured with *S. faecalis* following an oral dose of pteroylglutamic acid (Denko, 1951; Spray and Witts, 1952; Chanarin, Anderson, and Mollin, 1958). The patient is saturated with folate before the test to ensure that absorbed folate is not rapidly removed from plasma by folate-deficient tissues.

Studies in man have also been performed by using the double-lumen tube technique, i.e., perfusing a given segment of small intestine and measuring folate absorption by the difference in concentration between folate infused and withdrawn (Hepner, Booth, Cowan, Hoffbrand, and Mollin, 1968); by assessing the ability of a patient with megaloblastic anaemia due to folate deficiency to respond haematologically to physiological doses of pteroylglutamic acid given by mouth; and by measuring the rise in plasma radioactivity, and urinary and/or faecal excretion of radioactivity after an oral dose of tritiated pteroylglutamic acid (Anderson, Belcher, Chanarin, and Mollin, 1960; Kinnear, Johns, MacIntosh, Burgen, and Cameron, 1963; Klipstein, 1963; Paterson, David, and Baker, 1965; Kremen chuzky, Musso, Hoffbrand, and Rocha Viola, 1967; Yoshino, 1968a).

Findings

Pteroylglutamic acid is rapidly absorbed from the duodenum and jejunum and a rise in blood folate level occurs as soon as 15 minutes after an oral dose. Between 60% and 80% of a single dose is absorbed whether this is small (25-200 µg) or large (1-15 mg). When pteroylglutamic acid is infused at a constant rate into the jejunum, the proportion absorbed is the same until very high concentrations (10 µg/ml) are reached when the proportion absorbed falls (Hepner et al, 1968). Folate is comparatively poorly absorbed from the lower small intestine, the fall in proportion absorbed from the upper to lower jejunum being much steeper for pteroylglutamic acid than for glucose (Hepner et al, 1968). Folate is not absorbed from the large intestine (otherwise, folate-producing colonic bacteria would be able to supply the body with considerable amounts of the vitamin).

There is no definite evidence whether or not pteroylglutamic acid is absorbed by an active, energy requiring process in man. Most of a dose of 1 mg enters the portal blood unchanged (Whitehead and Cooper, 1967) and the pteroylglutamic acid released from a large dose of pteroylglutamic acid also enters the portal blood intact (Butterworth, Baugh, and Krumdieck, 1969). Possible evidence that pteroylglutamic acid is absorbed against a concentration gradient in man is that there is no inhibition of pteroylglutamic acid absorption when the blood folate is raised by parenteral administration of pteroylglutamic acid (Hepner et al, 1968), but the exact form of folate on the mucosal side of the jejunal brush border in such an experiment is uncertain.

Baker, Frank, Feingold, Ziffer, Gellene, Leevy, and Sobotka (1965) suggested that pteroylglutamic acid in doses as high as 5 mg was reduced and methylated during passage through the small intestine, but they did not exclude the possibility that the compound was absorbed unchanged and then converted into 5-methyltetrahydrofolate (the form of folate in normal plasma) by the liver or exchanged with liver 5-methyltetrahydrofolate. Most workers consider that such large doses of pteroylglutamic acid are absorbed intact. However, it is possible, though not established, that there is a minor degree of conversion of low doses of pteroylglutamic acid to 5-methyltetrahydrofolic acid during its transfer through the jejunal mucosa. As discussed below, there is stronger evidence for conversion of partly reduced folate to 5-methyltetrahydrofolic acid by the jejunum (Perry and Chanarin, 1970).

Baker, Frank, and Sobotka (1964) suggested that pteroylglutamic acid is conjugated to the triglutamate form during absorption but there has been no support for this hypothesis.

Perhaps the best evidence that absorption of pteroylglutamic acid in man involves a specific process is provided by the observations of Luhby, Eagle, Roth, and Cooperman (1961) and Lanzkowsky, Erdlandson, and Bezan (1969) of individuals with specific malabsorption of folates, including pteroylglutamic acid (see below) but, as yet, there is no definite evidence that this process requires energy.
ANIMAL STUDIES

Most studies have been performed in the rat or hamster. The jejunum is the principal site of folate absorption in these animals and absorption is maximum at a pH around 6.0 (Elsborg, 1970; Smith, Matty, and Blair, 1970). It is again uncertain whether or not pteroylglutamic acid is actively transferred across the small intestine. Burgen and Goldberg (1962) using in-vivo perfused loops, Herbert and Shapiro (1962) and Herbert (1967) using the everted sac technique, and Hepner (1969) using the in-vivo tied loop system all conclude that pteroylglutamic acid is actively transported by the jejunum at low doses and passively transferred at higher doses. On the other hand, Turner and Hughes (1962), Spencer and Bow (1964), and Yoshino (1968b) all conclude, on the basis of everted sac studies, that pteroylglutamic acid is passively absorbed across the jejunum of the rat at all doses. In a recent careful study, albeit with the everted sac preparation, Smith et al. (1970) conclude that transport of pteroylglutamic acid at concentrations ranging from 1 x 10^{-7} M to 1 x 10^{-8} M across the jejunal mucosa of the rat occurs by a saturable process, partly by passive diffusion and partly by glucose-stimulated solvent drag with water flow.

All workers agree that transport of pteroylglutamic acid across the ileum of the rat is passive at all concentrations.

On the basis of differential microbiological assays, Cohen (1965) postulated that pteroylglutamic acid was converted to 5-methyltetrahydrofolate during transport across rat jejunum, but these studies are inconclusive since bacterial synthesis of the methylfolate in the (unsterile) everted sac preparation was not excluded. A possible point in favor of Cohen’s conclusion is the observation by two groups (Burgen and Goldberg, 1962; Hepner, 1969) that methotrexate, which inhibits reduction of pteroylglutamic acid, also inhibits absorption of pteroylglutamic acid in the rat. This could be interpreted to indicate that reduction to the tetrahydrofolate form is a key, rate-limiting reaction in the pteroylglutamic acid absorption process. This is unlikely, however, in view of the convincing demonstration by Smith et al. (1970) that pteroylglutamic acid is largely unchanged after transport across the rat jejunal mucosa in vitro. Instead inhibition by methotrexate might indicate that pteroylglutamic acid and methotrexate compete for the same absorptive mechanism. Methotrexate has indeed been shown to interfere with transport of pteroylglutamic acid into haemopoietic cells (Das and Hoffbrand, 1970). Yoshino (1968b) could not demonstrate an effect of methotrexate on pteroylglutamic acid absorption in the rat, however, so no conclusion can yet be reached on this point.

Human Absorption of Natural Folates

FORMS OF FOLATE

The small intestine in the human adult is presented with folate from three sources—from the diet, from bile, and from sloughed intestinal cells.

Dietary folate

The normal adult western diet contains about 600-700 μg of folate daily and approximately three-quarters of this is in the polyglutamate form (Butterworth, Santani, and Frommeyer, 1963; Chanarin, Rothmann, Perry, and Stratfull, 1968; Hurdle, Barton, and Searles, 1968). The uncooked diet contains much larger amounts of folate but up to 100% of this may be lost when high temperatures and large amounts of water are used in cooking. Major compounds in the diet are 5-methyl-, 5-formyl-, and 10-formyl-tetrahydropteroylglutamates. The polyglutamate forms with more than three glutamate moieties are not microbiologically active until they have been hydrolysed to the simpler tri, di-, or monoglumamate forms, when they can be assayed with L. casei, and are termed ‘free’ (Table I).

Biliary folate

S. Baker, Kumar, and Swaminathan (1965) first demonstrated an enterohepatic circulation for folate in man. They found a mean folate level of 32-6 ng/ml in the duodenal juices of eight normal subjects who had a mean serum folate level of 4-6 ng/ml; apparently between 60 and 90 μg of folate enters the bile each day. Biliary folate is mainly in the form of 5-methyl-tetrahydrofolate and formylfolates (Bernstein, Gutstein, Weiner, and Efron, 1970b; Pratt and Cooper, 1971).

Folate in sloughing intestinal cells

This is extremely difficult to quantitate. Small amounts of folate are present in human jejunal mucosa and these are probably mainly in the free form of the vitamin when they reach the intestinal lumen.

ABSORPTION OF PTEROYLPOLYGLUTAMATES

Two groups have synthesized labelled pteroylpolylglutamates and studied their absorption both microbiologically and by radioactive techniques (Butterworth et al., 1969; Godwin and Rosenberg, 1970; Rosenberg and Godwin, 1971). All other studies have been carried out using microbiological assay of serum or urine after oral doses of natural folates. Early studies using S. faecalis as a test organism suggested that very little folate could be absorbed from yeast (Spray, 1952). It is now established,
Folate absorption and malabsorption

however, that significant amounts of folate are absorbed from the higher pteroylpolyglutamates of yeast and other foods.

The folate entering the blood stream after feeding pteroylpolyglutamates is active for L. casei and has been shown to be in the pteroylmonoglutamate form (Butterworth et al, 1969). This indicates that hydrolysis has occurred during absorption. Recent work from many laboratories has failed to support the earlier suggestion of Cooperman and Luhy (1965) that pteroylpolyglutamates can be absorbed intact. The proportion of a dose of yeasts pteroylpolyglutamates that can be absorbed has been estimated to be similar to that of pteroylglutamic acid at a dose of about 200 µg, while at higher doses absorption has been estimated to be only one-third that of pteroylglutamic acid (Streiff and Rosenberg, 1967; Hoffbrand and Necheles, 1968; Perry and Chanarin, 1968; Hoffbrand and Peters, 1970). It is likely that the compounds with the greatest number of glutamate moieties are least well absorbed (Butterworth et al, 1969).

Most studies of pteroylpolyglutamate absorption have been performed with compounds partly or completely purified from yeast. Retief (1969) has shown that there may be wide variations in the availability of pteroylpolyglutamates from different foods, for instance, that folate from calf's liver, peas, and spinach is better absorbed than equivalent amounts from tomato, cauliflower, and pumpkin. Earlier observations of Baumslag and Metz (1964) suggest that folate in lettuce is relatively well absorbed. Apart from differences due to variation in the chemical composition of the polyglutamates, differences may also arise because of other substances in the foods. Cellulose has been reported to impede folate absorption by complexing the vitamin in insoluble form (Luther, Santini, Brewster, Perez-Santiago, and Butterworth, 1965). There may also be 'folate conjugate inhibitors' in food and this point is discussed next.

PTEROYLGLUTAMATE HYDROLASE

The enzyme responsible for the hydrolysis of the glutamyl-γ-glutamyl peptide chain has been called 'folate conjugase' (Bird, Binkley, Blood, Emmett, and Pfiffrner, 1945) but this is an unsatisfactory name since it suggests the opposite effect to the 'deconjugation' that the enzyme carries out. The alternative names 'pteroylglutamate hydrolase' (PPH) (Hoffbrand and Peters, 1969) and 'glutamylglutamylcarboxypeptidase' (Blakley, 1969; Bernstein, Gutstein, and Weiner, 1970a) are therefore used. The enzyme, which hydrolyses the peptide chain to the monoglutamate form, has not however, been purified, and it is possible that two or more enzymes are concerned, for instance, one that hydrolyses the higher polyglutamates to the triglutamate stage, and a second which takes the tri- to the monoglutamate form (Mims and Bird, 1950). Similar enzymes exist in nature, such as that in chick pancreas, but this differs from human PPH by such characteristics as pH optimum and by producing pteroyldiglutamate as an end-product.

There is no delay in the absorption of polyglutamate forms compared to pteroylglutamic acid, implying that the hydrolysis of dietary polyglutamates to monoglutamates occurs rapidly. The exact anatomical site of the hydrolysis is, however, uncertain. Small amounts of PPH are present in saliva, bile, pancreatic, and duodenal juices, but the pH optimum of the enzyme is low (between 4-0 and 5-0) and enzyme activity in the lumen of the upper small intestine from these sources and from sloughed intestinal cells is very low (Santini, Berger, Berdasco, Sheehy, Avites, and Daisla, 1962; Klipstein, 1967; Hoffbrand and Peters, 1970). Much higher concentrations of the enzyme are present in the jejunal mucosa, and this seems a more likely site of the hydrolysis, particularly, as deconjugation of pteroylpolyglutamates has been reported during transport across everted sacs of rat intestine in vitro (Rosenberg, Streiff, Godwin, and Castle, 1969) where the luminal enzyme is presumably absent. Moreover, absorption of pteroylpolyglutamates appears to be normal in adult pernicious anaemia (see Table V) when presumably the pH of the upper small intestine is particularly unfavourable for the action of the luminal enzyme.

Surprisingly, if the mucosal enzyme does have an absorptive function, it is not situated in the brush border of the mucosal cell, the usual subcellular site for an absorptive enzyme, but is concentrated in the mucosal cell lysosomes (Hoffbrand and Peters, 1970). Furthermore, the concentration of the enzyme in jejunal and ileal mucosa in man is the same (Hoffbrand and Peters, 1970)—though Bernstein et al (1970b) report that in the guinea pig the jejunal concentration of the enzyme is higher than that of the ileum.

If polyglutamate hydrolysis does occur within the intestinal cells, the localization of the enzyme in lysosomes suggests that the compounds may be absorbed by a process of pinocytosis and are then digested in the secondary lysosomes formed by fusion of pinocytic vacuoles with primary lysosomes containing the enzyme (Hoffbrand and Peters, 1970). Alternatively, it is possible that polyglutamates are transported to the lysosomes by some process other than pinocytosis though, in view of their relatively high molecular weight (greater than 800), they would not be expected to cross the lyso-
somal membrane. The easy saturation of the absorptive process for pteroylpolyglutamates implies that entry of the compounds into the cells may be one of the limiting factors in their absorption, the location of the enzyme inside the mucosal cell and the large amounts present there making it less likely that hydrolysis within the mucosal cell could be a limiting factor.

If, on the other hand, deconjugation takes place only in the small intestinal lumen, then this could well be a limiting factor in view of the small amounts of enzyme found there. Until more is known about the mechanism of entry of all folates into the jejunal cells, however, it is difficult to know which process is so easily saturated during polyglutamate absorption.

**Inhibitors of pteroylpolyglutamate hydrolase**

The enzyme is usually estimated by incubating non-\textit{L. casei} active pteroylpolyglutamate substrate with the enzyme preparation and measuring the amount of free folate released in a given time. A more elegant technique using labelled pteroylpolyglutamates has also been described (Baugh and Krumdieck, 1969).

The presence of inhibitors of PPH in natural materials was originally suggested by Mims, Swendsen, and Bird (1947). They showed an apparent inhibition of enzyme activity by nucleic acids. Subsequent work has shown that DNA and RNA directly inhibit the growth of \textit{L. casei} which was used to assess the enzyme activity and fail to confirm that these substances inhibit the enzyme itself (A. V. Hoffbrand, C. Griffin, and T. J. Peters, unpublished observations). Glutamic acid polypeptides linked to para-aminobenzoic acid have also been reported to inhibit chick pancreas PPH (Sims and Totter, 1947) while Bernstein \textit{et al} (1970a and b) describe inhibition of human PPH by bile salts and by sulphobromophthalein. Anticonvulsant drugs and the contraceptive 'pill' have also been described as PPH inhibitors and these hypotheses are discussed further below.

**FURTHER EVENTS**

It is likely that partly or fully reduced pteroylmonoglutamates (whether ingested as such or derived by hydrolysis of partly or fully reduced pteroyl polyglutamates) are fully reduced and methylated within the jejunal mucosa before they enter portal blood (Chanarin and Perry, 1969; Perry and Chanarin, 1970). This conclusion is based on the finding of radioactive folate active for \textit{L. casei} and not for \textit{S. faecalis} in the blood stream after oral ingestion of radioactive dihydrofoleric and tetrahydrofoleric acid but not after their intravenous administration. Whitehead, Pratt, Viallet, and Cooper (1970) have also shown that ingested 5-formyltetrahydrofolic acid (folinic acid, citrovorum factor) is also converted to 5-methyltetrahydrofolate during its transfer across the jejunal mucosa in man.

Two enzyme systems must be concerned in these conversions: (1) dihydrofolate reductase (DHFR) which reduces dihydrofolate (and much less readily pteroylglutamic acid) to tetrahydrofolate; (2) at least two enzyme(s) concerned with methylation of tetrahydrofolate at the N\textsubscript{5} position (eg, serine hydroxymethyltransferase and 5,10-methylenetetrahydrofolate reductase).

Darzynkiewicz, Rogers, Barnard, Wong, and Werkheiser (1966) using autoradiographic techniques localized DHFR to the apical position of the jejunal mucosal cells in the mouse. In our own studies, we have found the enzyme to be localized principally in the soluble (cell sap) fraction of the jejunal cells in the guinea-pig (Table II), so that if both deconjugation and reduction do take place in the mucosal cell they must take place in different parts (lysosomes and cell cytoplasm) of the cell.

<table>
<thead>
<tr>
<th>Protein (n mol/mg/hr)(^{1})</th>
<th>Whole homogenate</th>
<th>Brush borders and nuclei</th>
<th>Mitochondria and lysosomes</th>
<th>Microsomes</th>
<th>Cell sap</th>
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<tr>
<td></td>
<td>88±3</td>
<td>8±2</td>
<td>-</td>
<td>17±1</td>
<td>101±5</td>
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</table>

Table II **Subcellular localization of dihydrofolate reductase in guinea-pig intestinal mucosa\(^{2}\)**

\(^{1}\)Assay method of Bertiino and Fischer (1964).
\(^{2}\)Mean of four experiments.

**CONCLUSIONS**

Dietary pteroylpolyglutamates are hydrolysed to pteroylmonoglutamate form, probably in the jejunal mucosa. If the compounds are already partly reduced, it is likely that they are fully reduced and methylated in the mucosa so that the major compound entering portal blood after ingestion of a wide variety of dietary folate compounds is 5-methyltetrahydropteroylmonoglutamate. Pteroylglutamic acid itself is largely absorbed unchanged but whether this is by an active or passive process and whether partial reduction and methylation of pteroylglutamic acid also occurs is uncertain. In both animals and man, however, a saturable process is involved in transfer of pteroylglutamic acid across the upper small intestinal mucosa.

**Malabsorption of Folate**

The most usual cause of folate deficiency in the western hemisphere is inadequate dietary intake of the vitamin. Many severely folate-deficient patients...
Folate absorption and malabsorption

also suffer from a condition which accelerates folate depletion, eg, haemolytic anaemia or myelosclerosis; the most common factor precipitating folate deficiency throughout the world, however, is pregnancy. There are only three diseases in which malabsorption of folate is considered the major cause of the deficiency—tropical sprue, coeliac disease, and the extremely rare disease of specific malabsorption of folate (Table III). In a number of other conditions, malabsorption of folate may play a part, at least in some of the patients, in causing folate deficiency though in these conditions, inadequate intake and/or excess utilization of the vitamin are probably more important factors in patients with severe deficiency (Table IV). The conditions in which there is still dispute about whether folate malabsorption occurs at all, or where folate malabsorption has been reported in only one study of pteroylglutamic acid absorption are given in Tables Va and b.

Established Causes of Malabsorption of Folate

Tropical sprue and coeliac disease

Malabsorption of folate is now known to be a consistent feature of untreated tropical sprue and of untreated coeliac disease, both in children and adults. In these diseases there is structural and functional damage to the jejunal mucosa. Folate deficiency may be due not only to malabsorption of dietary folate but also to failure of absorption of biliary folate, and of folate from sloughed intestinal cells. Loss of folate from the latter source may well be excessive in these syndromes (Croft, Loehry, and Creamer, 1968). Reduced folate intake may also be a factor in many of the patients. Particularly in tropical sprue absorption of dietary folate appears to be more impaired than absorption of pteroylglutamic acid (Sheehy, Rubini, Perez-Santiago, Santini, and Haddock, 1961; Jeejeebhoy, Desai, Borkar, Deshpande, and Pathase, 1968; Hoffbrand, Necheles, Maldonado, Horta, and Santini, 1969; Klipstein, 1969), even though PPH concentrations in the succus entericus (Klipstein, 1967) and in the jejunal mucosa in both tropical sprue and coeliac disease (Hoffbrand et al, 1969; Hoffbrand, Douglas, Fry, and Stewart, 1970) are normal.

It has been proposed that the selective malabsorption of pteroylglutamates in sprue is due to inhibition of jejunal PPH. No definite inhibitors have been identified in this disease, though a naturally occurring anti-folate compound (Butterworth, 1968) and a bacterial or viral inhibitor (Klipstein, 1968), have been proposed to have this action. More recently, Bernstein et al (1970b) have suggested that products of bile salt degradation by bacteria may be

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<th>Table III</th>
<th>Conditions in which malabsorption of folate has been established</th>
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<td>Disputed conditions in which malabsorption may occur</td>
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<td>Intestinal stigmoid-loop syndrome</td>
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<td>Congestive heart failure</td>
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<td>Treated pernicious anaemia</td>
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<th>Table Va</th>
<th>Serum folate levels (ng/ml) in control subjects and in patients with pernicious anaemia receiving maintenance vitamin B₁₂ therapy for at least one year</th>
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<th>Table Vb</th>
<th>Peak rise in serum folate level in five adult patients with treated pernicious anaemia following an oral dose of 200 μg (0-45 μ mole) pteroylglutamic acid and 200 μg (0-45 μ mole, folate equivalent) of a semi-purified preparation of pteroylglutamates prepared from yeast</th>
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<td>Patients</td>
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<td>Fasting Serum Folate (ng/ml)</td>
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<td>3</td>
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<td>4</td>
<td>12-1</td>
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<td>5</td>
<td>15-4</td>
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| Lower limit in normal subjects | 7-5 | 7-4 | |

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jejunal resection, deficiency folate folate remaining portion of the glutamic acid and (Chanarin to a found and folate cause bacteria the claving pteroylpolyglutamates (Hoffbrand 1968). Indeed, up to 70% of a 200 µg oral dose of pteroylglutamic acid may be absorbed in untreated coeliac disease (Kremenchuzky et al, 1967). Sufficient folate can usually be absorbed from large doses of pteroylglutamic acid (5 mg or more) to cause a satisfactory haematological response and to saturate body folate stores in patients with either tropical sprue or coeliac disease. Baker, Frank, Ziffer, and Feingold (1968), however, have made the remarkable, and as yet unconfirmed, observation that severe malabsorption of pteroylglutamic acid occurs in tropical sprue and adult coeliac disease which can be corrected by feeding the pteroylglutamic acid with lyophilized calf jejunum. Apparently the postulated factor in calf jejunum which enhances absorption of pteroylglutamic acid in these diseases was not a protein since it could not be destroyed by autoclaving the calf jejunum preparation. These remarkable findings remain totally unexplained.

Dermatitis herpetiformis
Patients with this skin disease may show a jejunal mucosal lesion resembling that of coeliac disease and in a proportion of patients the jejunal abnormality responds to a gluten-free diet. Absorption of pteroylglutamic acid is usually normal but nevertheless the patients show folate deficiency and malabsorption of pteroylpolyglutamates (Hoffbrand et al, 1970).

Jejunal resection
Malabsorption of pteroylglutamic acid and of dietary folate occurs if the jejunum is resected (Chanarin and Bennett, 1962; Baker, Thomson, and Feingold, 1969). Pavesio (1965) studied 30 children and found a slightly reduced absorption of pteroylglutamic acid in six patients with an ileal resection but marked reduction in nine patients with a jejunal resection. Sufficient absorption usually occurs from the remaining portion of the small intestine after jejunal resection, however, to protect patients from severe folate deficiency unless reduced intake of folate occurs.

Crohn's disease
Folate deficiency occurs frequently in patients with active Crohn's disease. A number of factors are involved—poor diet, excess utilization of the vitamin, and probably malabsorption in some patients. This is probably partly due to involvement of the jejunum by the disease but may also be due to impaired function of the small intestine not actually involved by the disease (Hoffbrand, Stewart, Booth, and Mollin, 1968).

Partial gastrectomy
Mild folate deficiency occurs frequently in post-gastrectomy patients; when megaloblastic anaemia due to the deficiency occurs, however, the predominant factor is always poor diet. A minor degree of malabsorption of pteroylglutamic acid and of dietary folate has been reported in a proportion of the patients (Cox, Meynell, Cooke, and Gaddie, 1958; Hoffbrand, Hines, Harrison, and Mollin, 1967; Markkanen, 1968; Chanarin, 1969). The cause of this may be excessively rapid passage of the vitamin from the gastric remnant to the lower jejunum, and possibly alterations in pH in the upper jejunum. Severe malabsorption of folate after partial gastrectomy may be due to occult coeliac disease.

Specific malabsorption of folate
This disease has been reported in two children (Lubby et al, 1961; Lubby and Cooperman, 1969) and one girl of 18 (Lanzkowsky et al, 1969; Lanzkowsky, 1970). The patients showed relapsing megaloblastic anaemia requiring therapy with large (10 mg or more) doses of pteroylglutamic acid by mouth, mental retardation and epileptic convulsions, and other neurological features, eg, ataxia or choreoathetotic movements. Absorption of all forms of folate tested was impaired, including absorption of pteroylglutamic acid, 5-methyltetrahydrofolate, and of pteroylpolyglutamates. Thus, lack of enzymes for deconjugation, reduction, or methylation could not be the explanation. The patient of Lanzkowsky responded well to a small parenteral dose of pteroylglutamic acid.

It is of interest that the patient of Lanzkowsky was also shown to have defective transport of folate into the cerebrospinal fluid and this may account for the neurological disturbances. It also suggests that folate absorption and folate transport into the cerebrospinal fluid may occur by related processes, and, since cerebrospinal fluid folate is normally three times that of plasma (Herbert and Zalusky, 1961), this is indirect evidence that both processes may be active.

POSSIBLE CAUSE OF MALABSORPTION OF FOLATE
In these conditions (Table IV) malabsorption of folate has been described but is not yet established as a significant cause of folate deficiency.
Intestinal blind-loop syndrome
Malabsorption of pteroylglutamic acid has been reported in a few patients with this syndrome (Cooke, Cox, Fone, Meynell, and Gaddie, 1963; Barrett and Holt, 1967; Wakisaka, 1968). These workers postulated that colonic bacteria in the upper jejunum might render folate unavailable for absorption. Hoffbrand, Tabaqchali, Booth, and Mollin (1971) have demonstrated lactobacilli capable of consuming folate in the small intestine of a patient with this syndrome. The predominant effect of an abnormal upper intestinal flora in the stagnant-loop syndrome, however, is to raise serum folate by producing folate that is absorbed (Hoffbrand, Tabaqchali, and Mollin, 1966; Klipstein and Lipton, 1970; Hoffbrand et al, 1971). Even though exceptional patients with large numbers of faecal organisms in the jejunum show folate deficiency and malabsorption of pteroylglutamic acid, there is as yet no conclusive evidence that the deficiency in these particular patients is due to malabsorption of dietary folate rather than to inadequate dietary intake of the vitamin (Hoffbrand et al, 1971).

Anticonvulsant drug therapy
Folate deficiency occurs frequently in patients receiving the anticonvulsant drugs, diphenylhydantoin (phenytoin) and primidone. A number of theories have been proposed to explain this deficiency. (1) Inhibition of enzymes concerned with folate metabolism but no such inhibition has been convincingly demonstrated. (2) Displacement of folate from its transport protein (Klipstein, 1964)—but cell uptake of folate is not affected by phenytoin in plasma (Das and Hoffbrand, 1970; Corcino, Waxman, and Herbert, 1971). (3) Induction of an enzyme concerned with folate metabolism causing excess folate utilization (Richens and Waters, 1971); this is unlikely since barbiturates are much more powerful enzyme inducers than phenytoin or primidone yet the latter two drugs are much more likely to cause folate deficiency. (4) Inhibition of PPH causing selective malabsorption of pteroylpolyglutamates (Hoffbrand and Necheles, 1968; Rosenberg, Godwin, Streiff, and Castle, 1968); other workers (Baugh and Krumdieck, 1969; Bernstein et al, 1970b) have not confirmed this. (5) Malabsorption of pteroylglutamic acid; this has been reported in the rat (Hepner, 1969) and in humans (Meynell, 1966; Dahlke and Mertens-Roesler, 1967). Hepner, Gerson, Hepner, Brown, Cohen, Herbert, and Janowitz (1970) suggested that the drugs do this by inhibiting intestinal Na-K ATPase. Aledort, Gerson, Cohen, Herbert, and Janowitz (1970) and Benn, Swan, Cooke, Blair, Matty, and Smith, (1971) have produced evidence for an alternative mechanism for phenytoin-induced malabsorption of pteroyl-glutamic acid. They demonstrated higher pH values in the upper small intestine of three patients on long-term anticonvulsant therapy who had developed folate-deficient megaloblastic anaemia than in control subjects. They also found that feeding either phenytoin or sodium bicarbonate with pteroylglutamic acid caused 'flat' absorption curves for the vitamin in normal volunteers. On the basis of these findings, they conclude that phenytoin causes malabsorption of pteroylglutamic acid by raising the pH in the lumen of the duodenum and jejunum above the optimum for pteroylglutamic acid absorption. Elsborg (1970) has indeed shown (in the rat) that phenytoin only causes malabsorption of pteroylglutamic acid if it raises the pH of the luminal contents, the drug being highly alkaline in solution, and as mentioned earlier, pteroylglutamic acid absorption is pH dependent with an optimum, at least in the rat, around pH 6-0.

It is necessary, however, to establish that the flat plasma curves after feeding pteroylglutamic acid with phenytoin reflect true malabsorption of pteroylglutamic acid rather than simply delayed absorption since other workers have found normal absorption of pteroylglutamic acid in drug-treated epileptics. Moreover, if alkalization of the upper small intestine does cause serious malabsorption of folate, it is surprising that in pernicious anaemia, a condition in which the upper small intestine may well be more alkaline than normal due to gastric achlorhydria, the mean serum folate level in the treated state is not significantly different from that of a control group and folate absorption is normal (Table V).

Contraceptive pill therapy
Some workers (Shojania, Hornady, and Barnes, 1968 and 1969), though not all (Spray, 1968; McLean, Heine, Held, and Streiff, 1968), have found low serum folate levels in women taking the 'pill'. Streiff (1970) reports that oral contraceptives cause selective malabsorption of pteroylglutamates by inhibiting intestinal PPH (Streiff and Green, 1970). In view of the findings of McLean, Heine, Held, and Streiff (1970) of similar absorption of pteroylglutamic acid and pteroylglutamates in pregnancy, these findings in patients receiving the synthetic oestrogens and progesterones are difficult to understand, and require confirmation. Coeliac disease was not excluded by jejunal biopsy as the cause of malabsorption in Streiff's folate-deficient cases, nor in two similar cases reported by Necheles and Snyder (1970).

Folate deficiency
The relation between folate deficiency and folate
absorption is complex. It is well known that folate therapy improves the structure and absorptive function of the small intestine in tropical sprue and improvement in intestinal absorption of xylose and vitamin B₁₂ has also been reported in anticonvulsant megaloblastic anaemia (Reynolds, Hallpike, Phillips, and Matthews, 1965). On the other hand, in nutritional folate deficiency jejunal structure has been reported both as normal (Winawer, Sullivan, Herbert, and Zamcheck, 1965) and megaloblastic (Bianchi, Chipman, Dreskin, and Rosensweig, 1970). Absorption of xylose and glucose is usually normal though malabsorption of vitamin B₁₂ occurs frequently, suggesting that ileal absorptive function is impaired. Tests of folate absorption are difficult to perform in the folate-deficient state since all tests of folate absorption except measurement of the faecal excretion of labelled folates require presaturation of the body with folate and this will of necessity correct folate deficiency of intestinal cells. It is likely, however, that folate deficiency contributes to malabsorption of folate in humans at least with tropical sprue and that the absorption of folate in these patients, like that of other nutrients, improves with folate therapy. It is also probable that folate absorption is reduced in other situations where the small bowel suffers general nonspecific damage as in kwashiorkor and starvation.

In each of the remaining conditions, only one group has reported patients showing malabsorption of pteroylglutamic acid.

Alcohol
A number of mechanisms are responsible for folate deficiency in alcoholics—poor nutrition, liver damage, and possibly inhibition of folate coenzymes. Halsted, Griggs, and Harris (1967) reported flat plasma curves after feeding ³H-pteroylglutamic acid in some chronic alcoholics. Total urinary excretion of radioactivity was normal, however, so absorption may well have been delayed rather than impaired.

Congestive heart failure
This is a recently recognized cause of folate deficiency (Hyde and Loehry, 1968; Brody, Soltys, and Zinsser, 1969) and the major factor is probably excess urinary folate excretion (Retief and Huskisson, 1969). Hyde and Loehry (1968) reported malabsorption of pteroylglutamic acid in seven of 25 patients tested, malabsorption being most frequent in those with long-standing heart failure.

Lymphosarcoma and chronic lymphocytic leukaemia
Pitney, Joske, and MacKinnon (1960) found reduced absorption of pteroylglutamic acid in eight of 10 patients with a lymphoma and five of eight with chronic lymphocytic leukaemia. They considered that these abnormalities were largely due to involvement of the small intestine by the disease since other tests of intestinal absorption were abnormal in most of the patients.

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